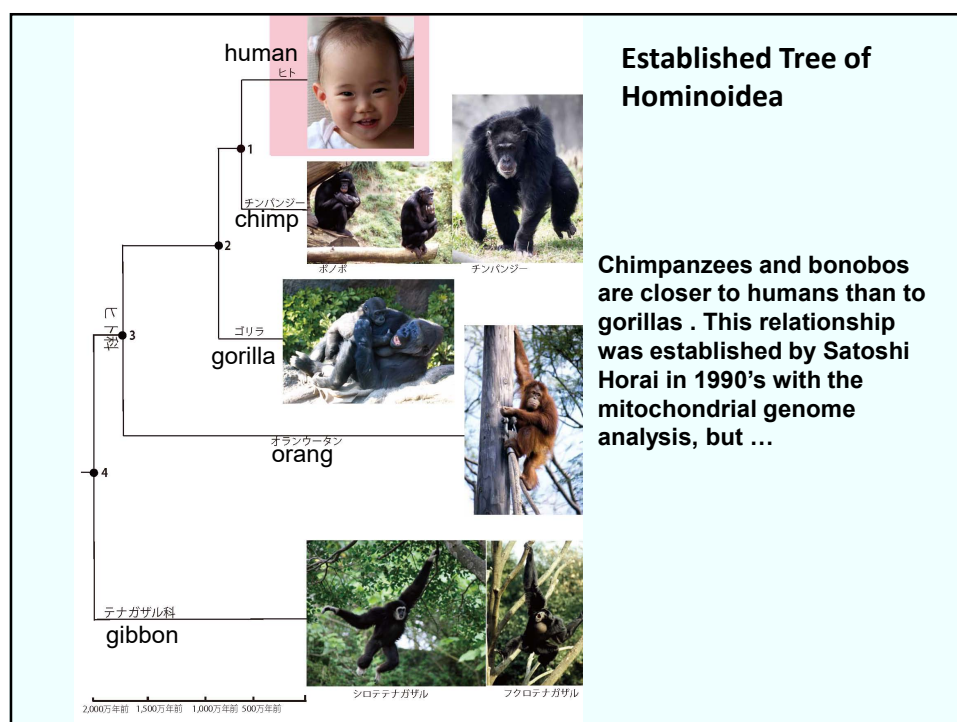


Importance of Population Genetics

Gene trees do not necessarily reflect phylogenetic relationships among species (species tree or population tree).

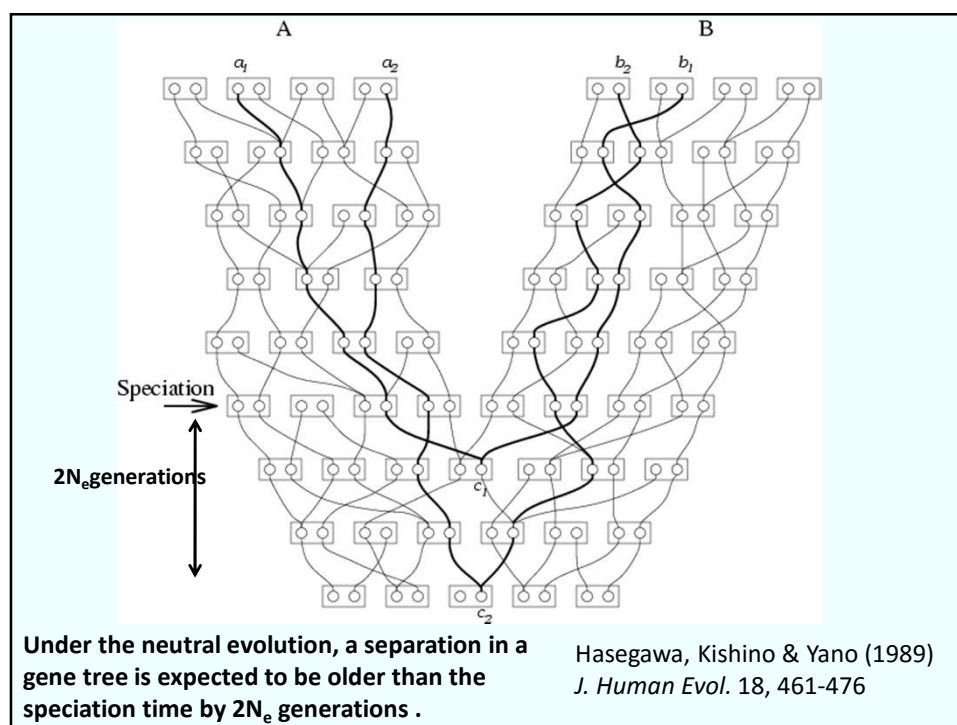


Number of genes significantly ($PP > 0.95$) supporting the 3 topologies among human, chimp and gorilla

	Gene	%
((Human,Chimp),Gorilla)	3,814	79.3
((Chimp,Gorilla),Human)	504	10.5*
((Human,Gorilla),Chimp)	492	10.2*

*: Incongruent trees (20.7%)

Modified from Table 3 of Ebersberger et al. (2007, Mol. Biol. Evol. 24, 2266-2276)



Molecular evolution rate

Evolution rate: v (substitutions/site/year)

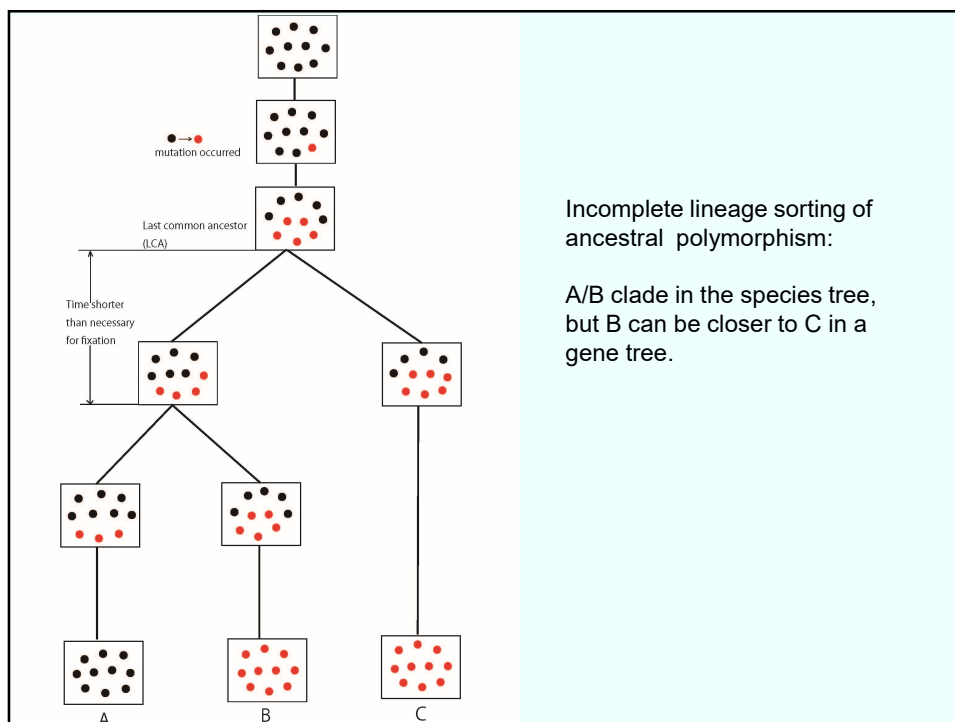
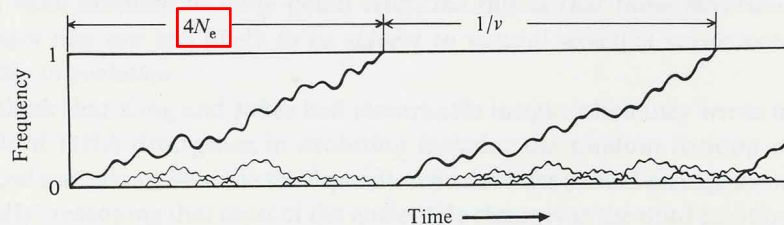
Effective population size: N_e

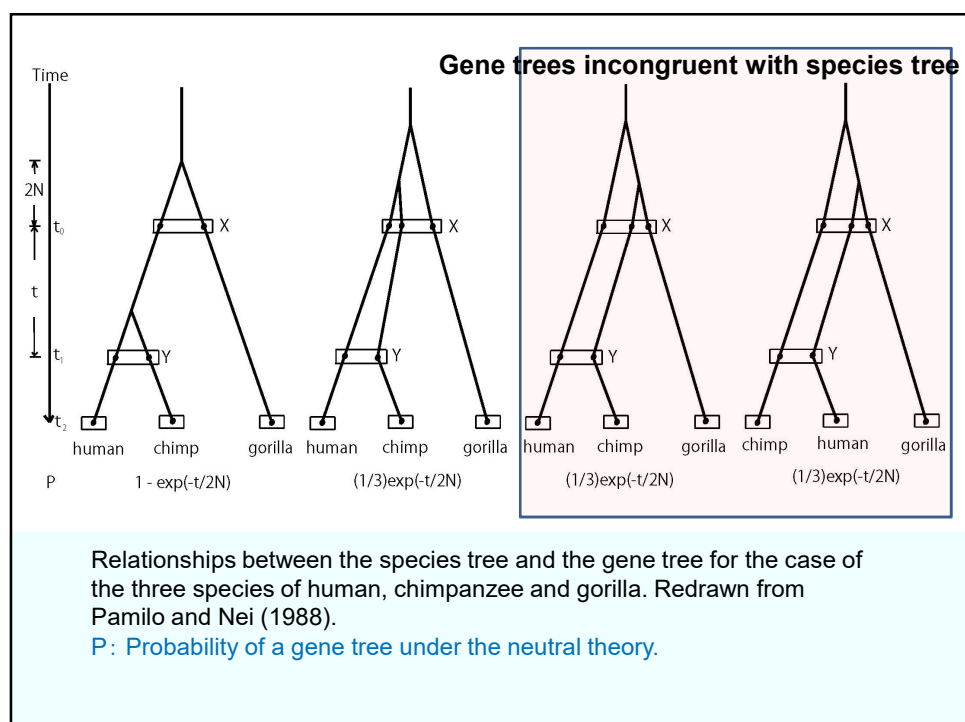
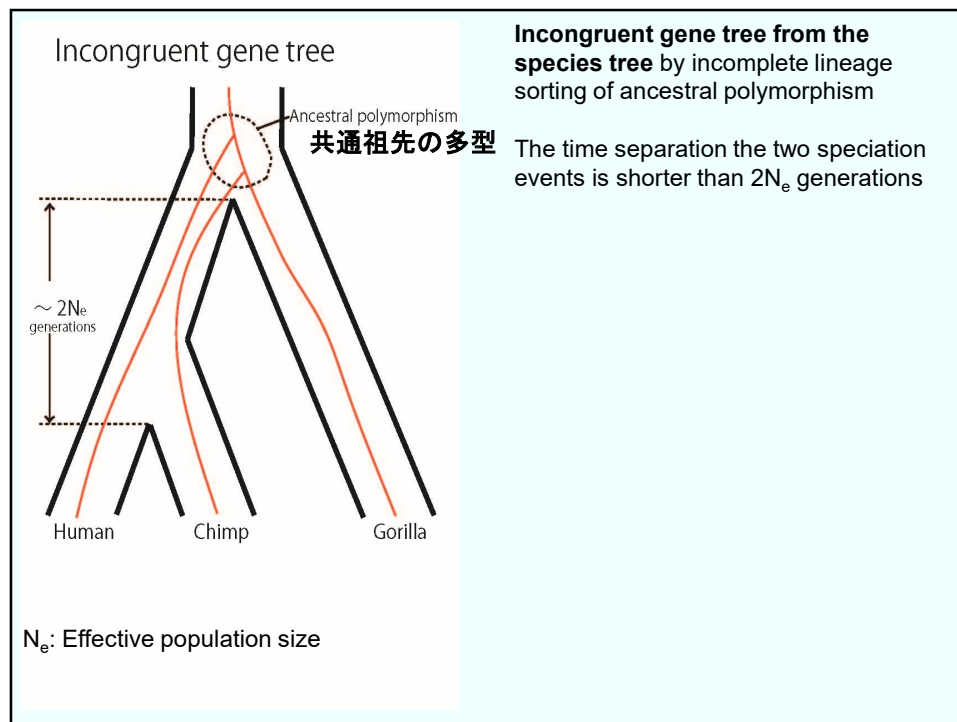
Mutation rate: μ

Probability of fixation of the gene: u

$v = 2N_e\mu u = \mu$: Under the neutral theory ($u = 1/2N_e$)

Fig. 3.1. Behavior of mutant genes following their appearance in a finite population. Courses of change in the frequencies of mutants destined to fixation are depicted by thick paths. N_e stands for the effective population size and v is the mutation rate.





Probabilities of gene trees

$T/2N_e$	((human, chimp), gorilla)	(human,(chimp,gorilla)) + ((human,gorilla),chimp)
	$1 - e^{-T/2N_e}$	$(2/3)e^{-T/2N_e}$
0.5	59.6%	40.4%
1	75.5%	24.5%
2	91.0%	9.0%
4	98.8%	1.2%

Number of genes significantly (PP>0.95) supporting the 3 topologies among human, chimp and gorilla

	Gene	%
((Human,Chimp),Gorilla)	3,814	79.3
((Chimp,Gorilla),Human)	504	10.5*
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*: Incongruent trees (20.7%)

Modified from Table 3 of Ebersberger et al. (2007, *Mol. Biol. Evol.* 24, 2266-2276)

Ebersberger et al. (2007)

Human Genetic Ancestry 2271

% of gene trees
rejecting the
human/chimp clade

Estimated effective
population size of the
human/chimp common
ancestor

Effective population size of
X-chromosome is $\frac{1}{4}$ of
those of autosomes, ...

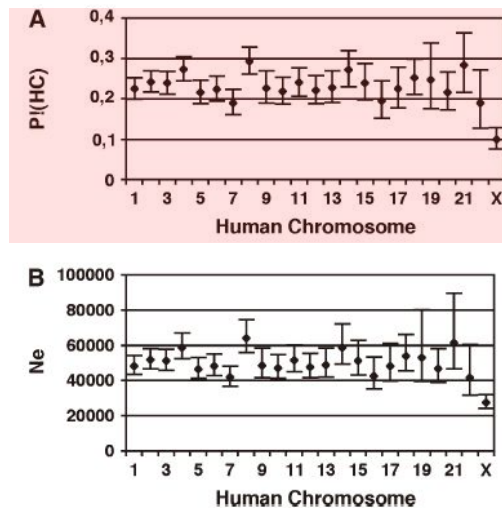


FIG. 3.—(A) Variation of the probability to observe a sequence tree significantly rejecting the human–chimp sister group ($P!(HC)$) among the human chromosomes. (B) Variation of the effective population size estimate of the human–chimp ancestral species among the human chromosomes. Bars denote the 95% CI.

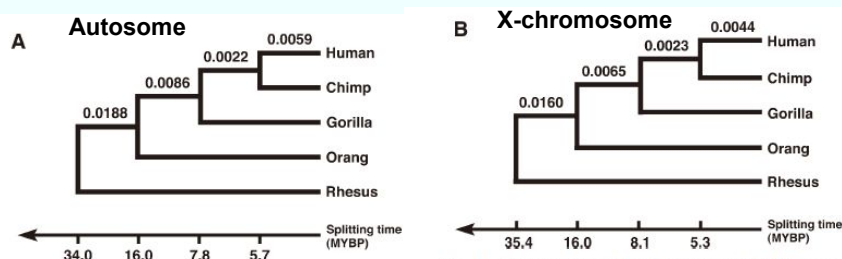
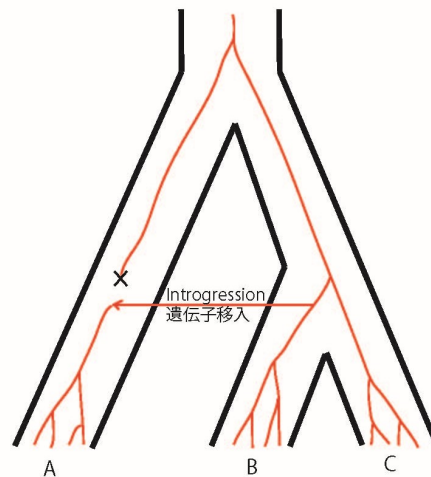


FIG. 4.—Sequence tree of the 5 species under study reconstructed from the autosomal sequences (A) and X chromosomal sequences (B), respectively. Splitting times were dated assuming the split of the orangutan lineage 16 MYBP. Trees are not drawn to scale.

Ebersberger et al. (2007)

The branching date estimated from X-chromosome is younger than that from autosomes (closer to species branching?)
→ Effective population size of mitochondria is $\frac{1}{4}$ of that of autosome, and therefore the branching date estimated from mitogenome may be closer to species divergence than those estimated from nuclear genes, but there is another problem...

Another possibility is introgression by hybridization



Effective population size of mitochondria is $\frac{1}{4}$ of autosomal genes, and therefore the introgressed haplotype lineage can be fixed more easily in mitochondrial genomes than in nuclear ones.

Caprinae

Himalayan tahr (*Hemitragus jemlahicus*)

Capra

Mitochondria introgression

bharal (*Pseudois nayaur*)

aoudad (*Ammotragus lervia*)

ibex (*Capra ibex*)

markhor (*Capra falconeri*)

goat (*Capra hircus*)

Ovis

mouflon (*Ovis montanus*)

bighorn (*Ovis canadensis*)

sheep (*Ovis aries*)

Cap

While Himalayan tahr is a sister-group to *Capra* in the mtDNA tree, it is close to bharal in the nuclear gene tree.

Merit and demerit of mitochondrial tree

- Effective population size of mtDNA is $\frac{1}{4}$ of those of nuclear autosomal genes (haploid and maternal inheritance)
 - Incomplete lineage sorting of ancestral polymorphism is less of a concern for mtDNA tree than for nuclear gene trees.
- On the other hand, small effective population size of mtDNA may facilitate fixation of introgressed haplotype lineage.

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Speciation of two gobioid species, *Pterogobius elapoides* and *Pterogobius zonoleucus* revealed by multi-locus nuclear and mitochondrial DNA analyses☆

Akihito ^a, Fumihito Akishinonomiya ^{b,c}, Yuji Ikeda ^d, Masahiro Aizawa ^d, So Nakagawa ^{e,i}, Yumi Umehara ^e, Takahiro Yonezawa ^{f,g}, Shuhei Mano ^g, Masami Hasegawa ^{f,g}, Tetsuji Nakabo ^h, Takashi Gojobori ^{e,j,*}

Pterogobius elapoides
キヌバリ

Pterogobius zonoleucus
チャガラ

Pterogobius zonoleucus
チャガラ

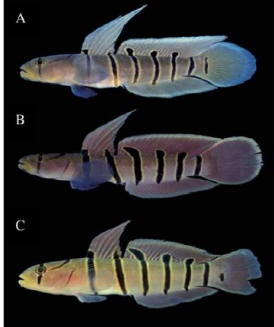

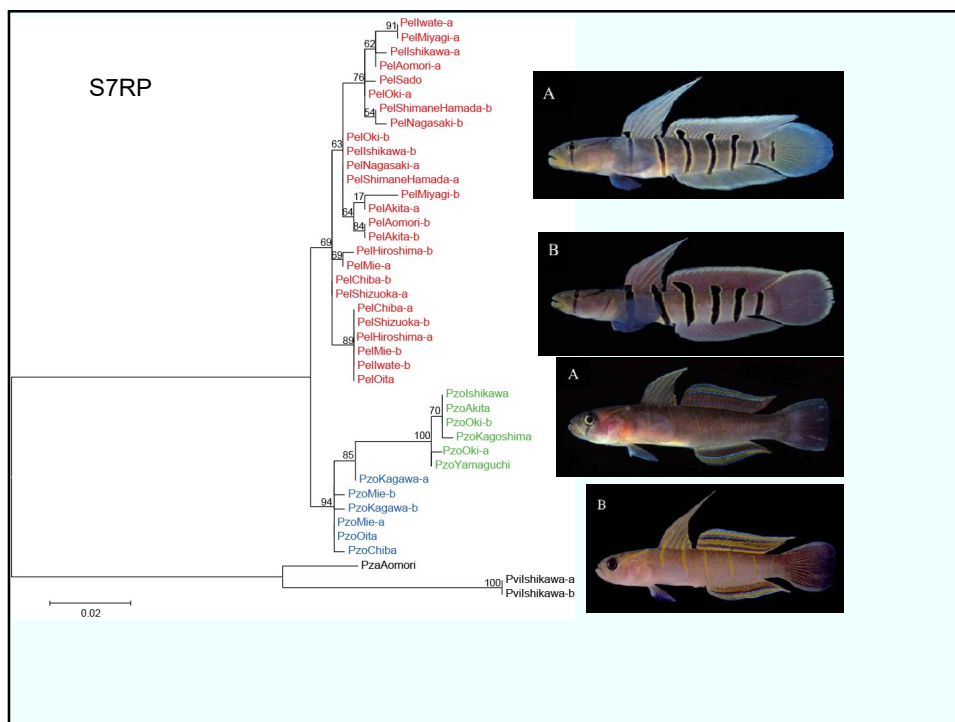
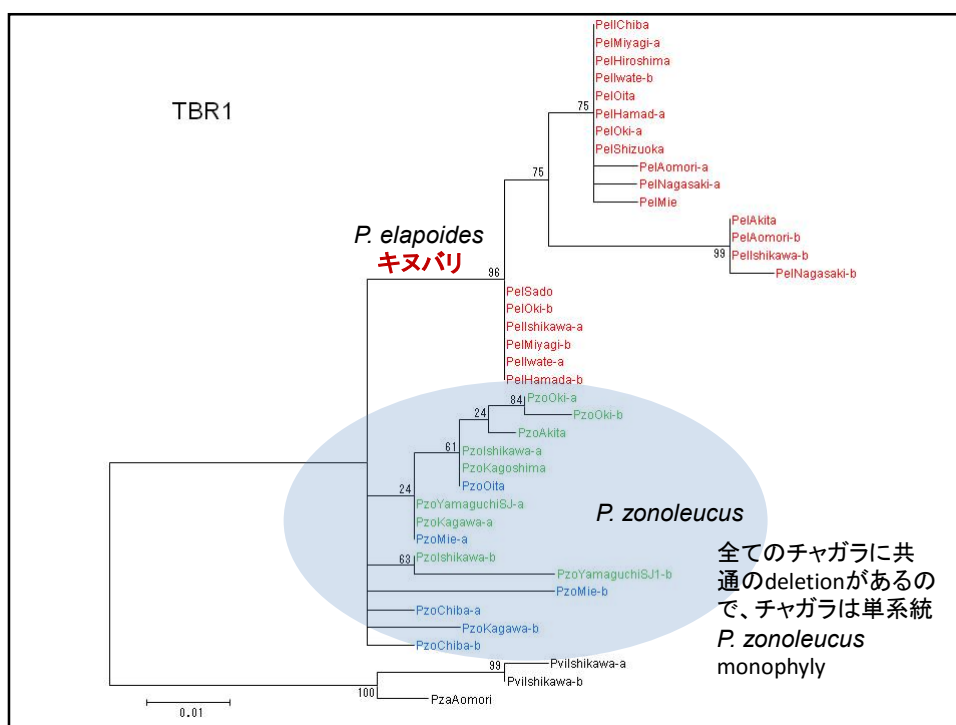
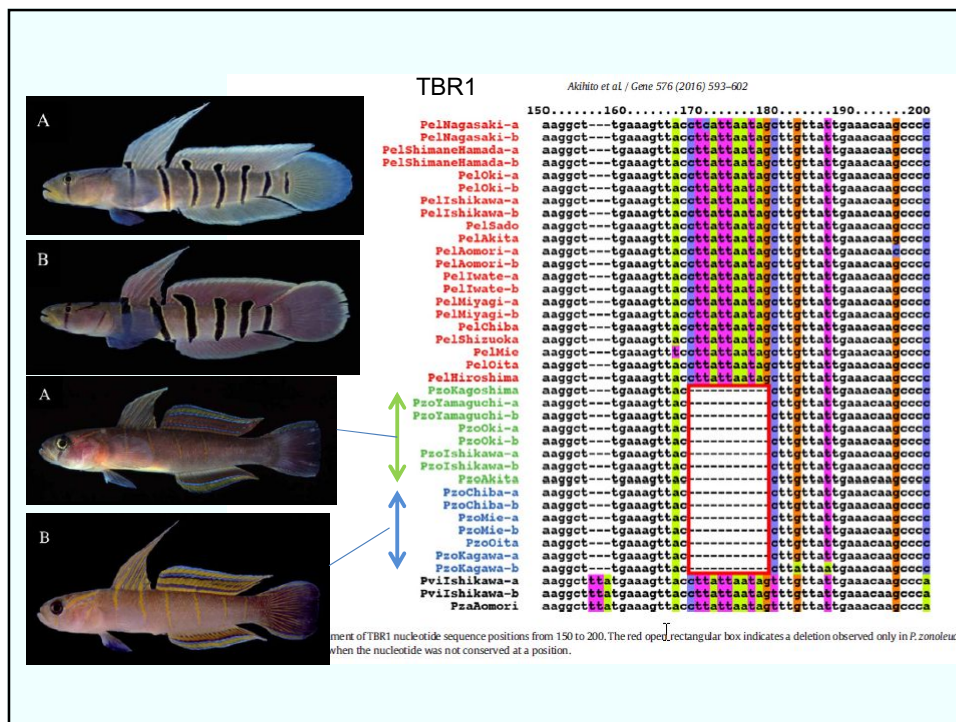



Fig. 1. *Pterogobius elapoides*: (A) The Sea of Japan, Ishikawa Prefecture (BLH20020439); (B) The Pacific Ocean, Chiba Prefecture (BLH20030460); (C) The Inland Sea, Hiroshima Prefecture (BLH20030461).

Fig. 2. *Pterogobius zonoleucus*: (A) The Sea of Japan, Ishikawa Prefecture (BLH20050505); (B) The Pacific Ocean, Chiba Prefecture (BLH20060883).

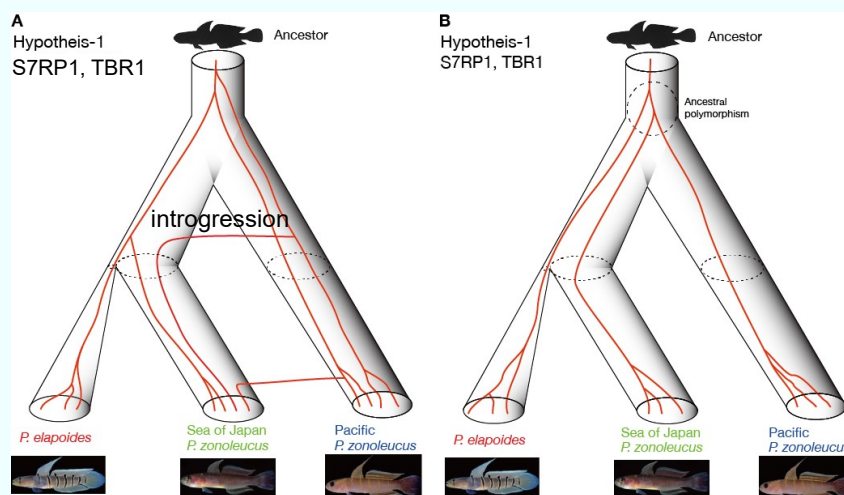




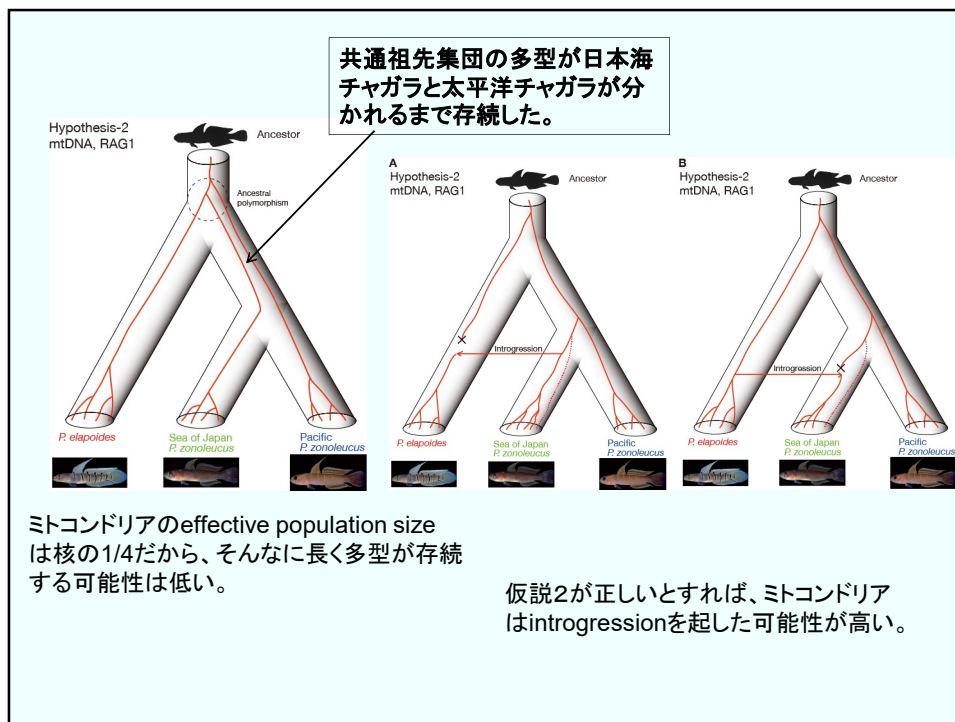
Two possible hypotheses

- Hypothesis-1: In the population tree, *P. elapoides* evolved from the Japan Sea population of *P. zonoleucus*.
- Hypothesis-2: In the population tree, the two populations (Sea of Japan & Pacific) of *P. zonoleucus* are monophyletic, and *P. elapoides* is a sister-group of these two.

How the incongruence of the gene trees can be explained by these hypotheses?



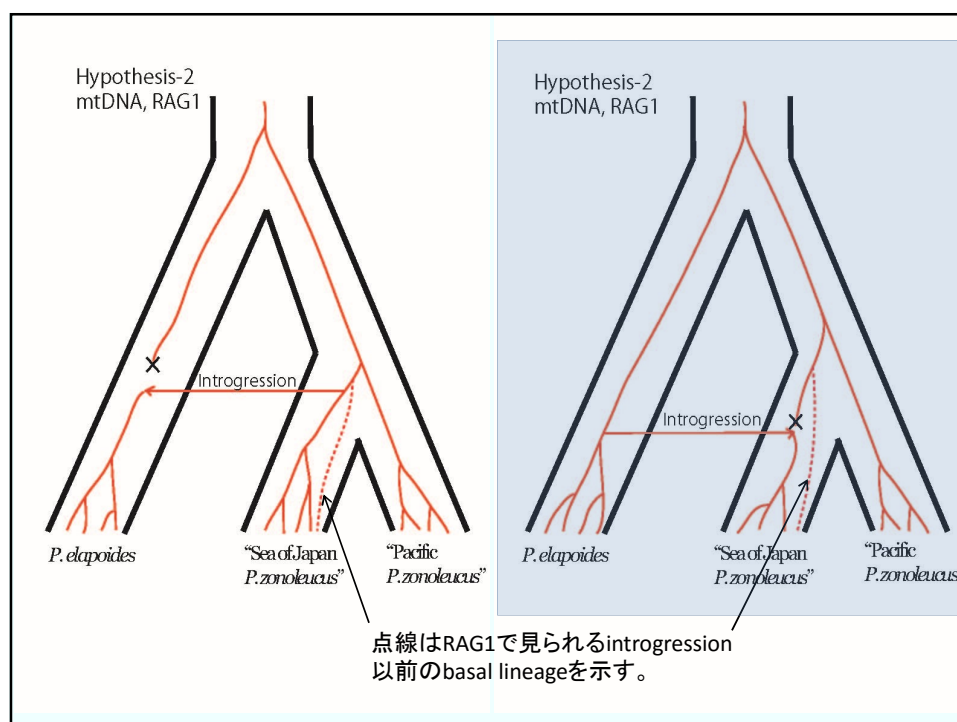
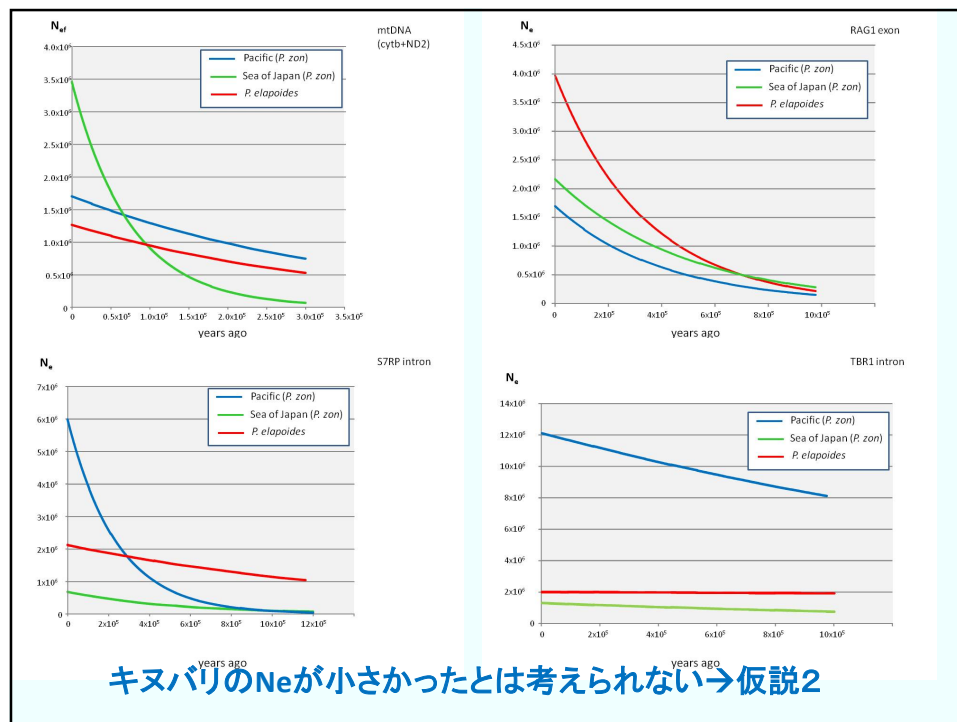
If Hypothesis-1 is true, it is expected that the population size of *P. elapoides* was expected to be smaller than that of Sea of Japan *P. zonoleucus*, because a new species usually evolve from a local population of the mother species.

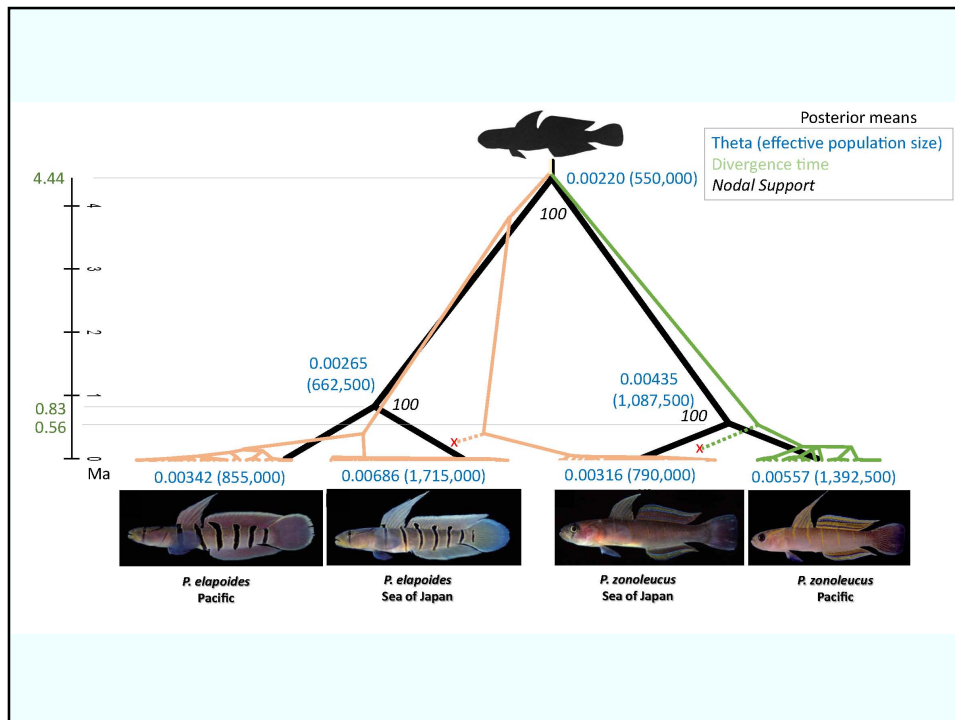


Suppl. Table 1. Estimates of $\theta = 4uN_e$ and $\beta = 2rN_e$, where N_e , u and r are effective population size, mutation rate per sequence per generation and exponential growth rate, respectively. Only for the mtDNA, $\theta = 2uN_{ef}$ and $\beta = rN_{ef}$, where N_{ef} is effective female population size.

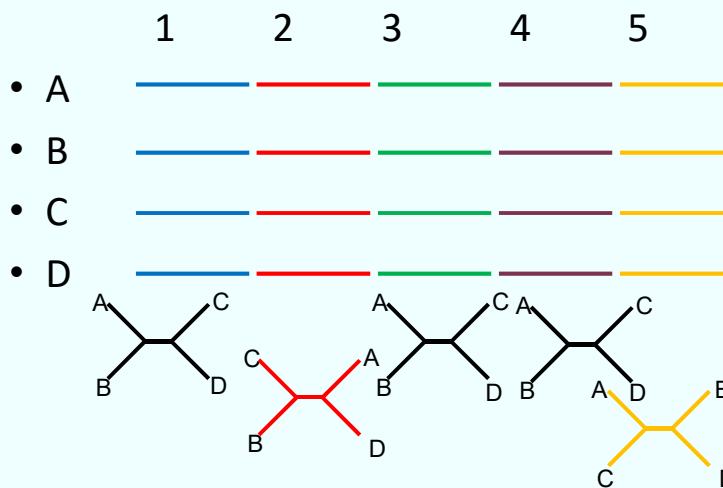
	mtDNA		RAG1 exon		S7RP intron		TBR1 intron	
Sequence length	2178 bp		1542 bp		853 bp		351 bp	
	θ	β	θ	β	θ	β	θ	β
Pacific <i>P. zon.</i>	74.0	4.7	10.4	8.3	20.4	50.3	17.0	10.0
SJapan <i>P. zon.</i>	150.5	46.2	13.4	9.1	2.3	2.6	1.8	1.4
<i>P. elapoides</i>	55.0	3.7	24.5	23.5	7.2	2.6	2.8	0.2

↑
現存集団では日本海チャガラはキヌバリのものよりも大きい、
種分化直後では逆転していた。





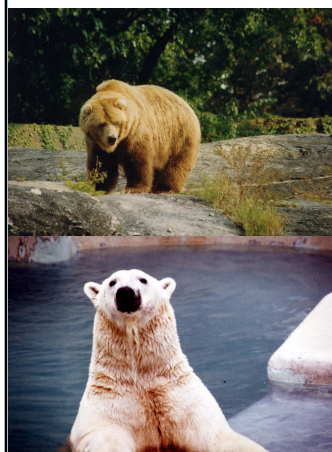
Analysis of concatenated sequences



Since gene trees can be incongruent among different genes, it cannot be justified to analyze concatenated sequences.

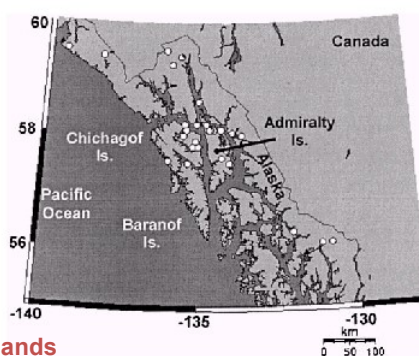
Phylogeography of Mitochondrial DNA Variation in Brown Bears and Polar Bears

Gerald F. Shields, Deborah Adams, Gerald Garner,^{*,†} Martine Labelle, Jacy Pietsch, Malcolm Ramsay,[†] Charles Schwartz,[‡] Kimberly Titus,[§] and Scott Williamson



Journal of Biology and Wildlife, 1999, Alaska Science Center, Fairbanks, Alaska 99775, Soldotna, Alaska 99686, and Wildlife Conservation Society, New York, New York 10024.

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ABC Islands

FIG. 1. Locations (open circles) on Admiralty, Baranof, and Chichagof islands and on the coastal mainland of southeast Alaska where tissues of brown bears were obtained.

		MITOCHONDRIAL DNA OF BEARS																323	
		nucleotide position																	
		1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
		5	5	5	5	5	5	5	5	5	5	5	5	5	5	5	5		
		2	2	2	3	3	3	4	4	4	4	4	4	4	5	5	5		
		6	7	9	3	4	6	0	1	2	5	5	6	0	1				
		6	2	9	5	1	2	4	0	8	3	9	4	9	9				
lineage		1	G	T	T	C	A	C	A	C	G	A	G	T	C	T		Brown Bears, ABC Islands (n=30)	←
		2	•	•	•	•	G	•	•	T	•	•	•	•	T	•		Polar Bears: Canadian Arctic Hudson Bay, Siberia (n=42)	
		3	A	•	•	•	G	•	•	T	•	•	•	•	T	•		Polar Bears, Hudson Bay (n=9)	
		4	•	•	•	•	G	•	•	T	•	•	•	•	C	T	•	Polar Bears: Canadian Arctic, Siberia (n=4)	
		5	•	C	•	T	G	T	C	T	A	G	A	•	T	C		Brown Bears, Southeast mainland, Kenai (n=5)	
		6	•	C	C	T	G	T	C	T	A	G	A	•	T	C		Brown Bears, Southeast mainland (n=7)	
		7	•	C	•	T	G	T	•	T	•	G	A	•	T	C		Brown Bears, Kenai (n=19)	

FIG. 7. Condensed dot matrix of variable nucleotide positions among 286 nucleotides (15243 to 15529) of the middle portion of the cytochrome *b* gene of bears of this study.

MtDNA of brown bear from ABC Islands is closer to those of polar bears than to those of brown bears from other areas.

