

Molecular evolution and phylogenetic tree

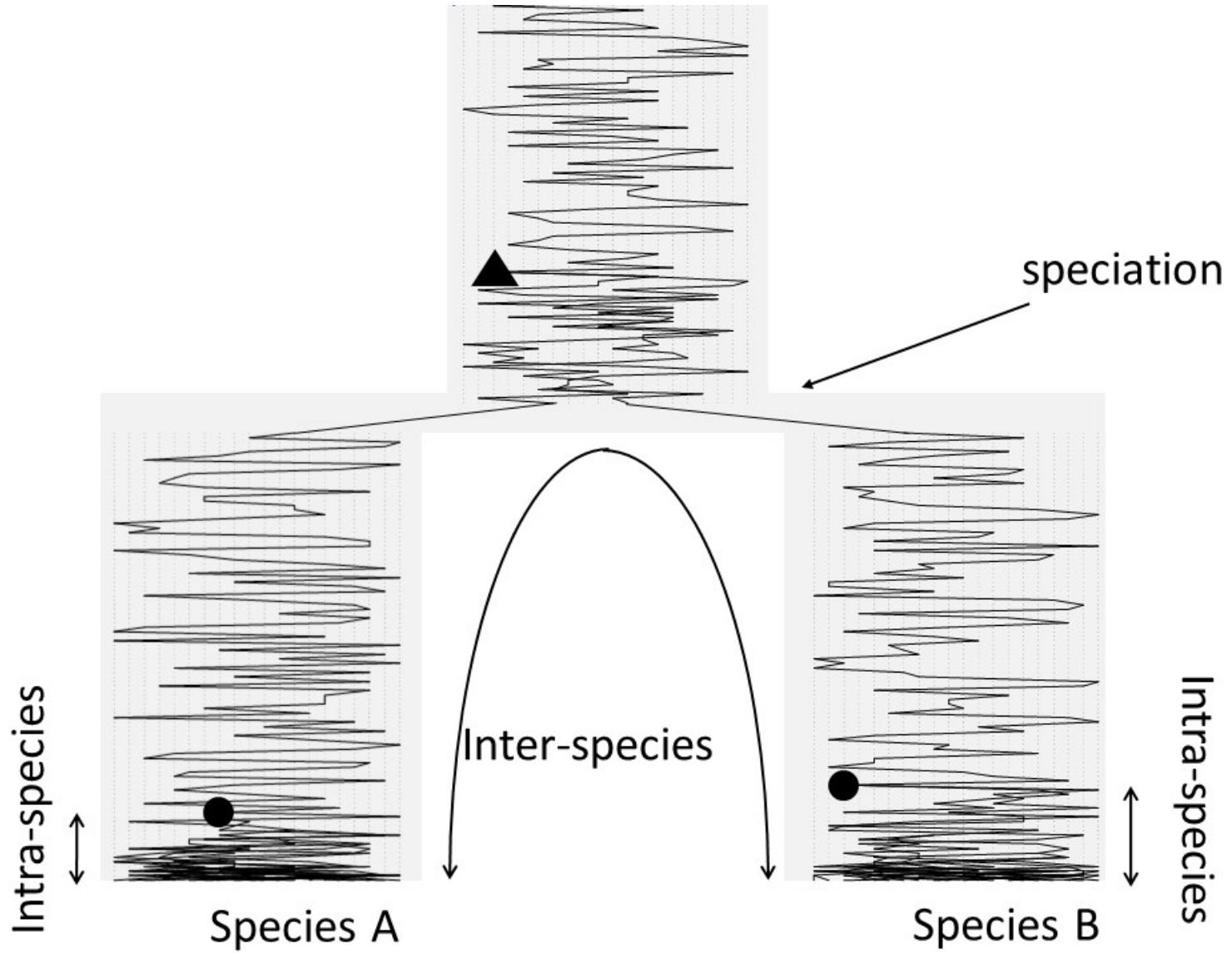
Intra-species polymorphism

- monomorphic
 - △ weakly polymorphic
 - polymorphic

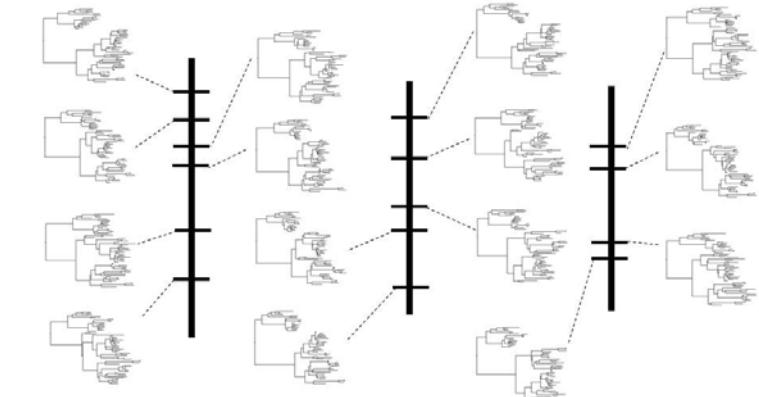
association

	●	△	●	●	O	●	●	●	●	O	●	●	phenotype
...	T	T	A	G	A	C	C	T	G	A	G	A	G
...	T	T	A	G	A	C	C	T	G	A	G	A	G
...	T	T	A	G	A	C	C	T	G	A	G	A	G
...	T	T	A	G	A	T	C	T	G	A	G	G	A
...	T	T	A	G	A	C	C	T	G	A	G	G	A
...	T	T	A	G	A	T	C	T	G	A	G	A	G
...	T	T	A	G	A	C	C	T	G	A	G	A	G
...	T	●	A	G	A	C	C	T	G	A	G	G	A
...	T	T	A	G	A	T	C	T	G	A	G	G	A
...	T	T	A	G	A	T	C	T	G	A	G	A	G
...	T	T	A	G	A	C	C	T	G	A	G	A	G
...	T	T	A	G	A	T	C	T	G	A	G	A	G
...	T	T	A	G	A	C	C	T	G	A	G	A	G

Inter-species diversity and intra-species polymorphism

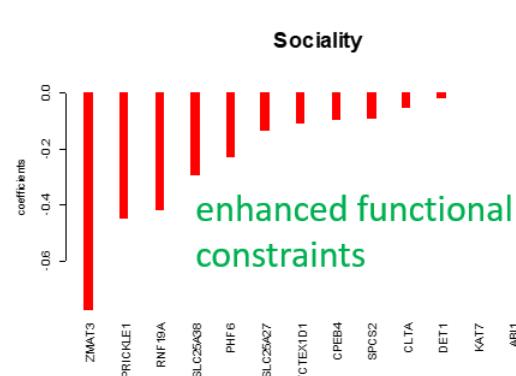
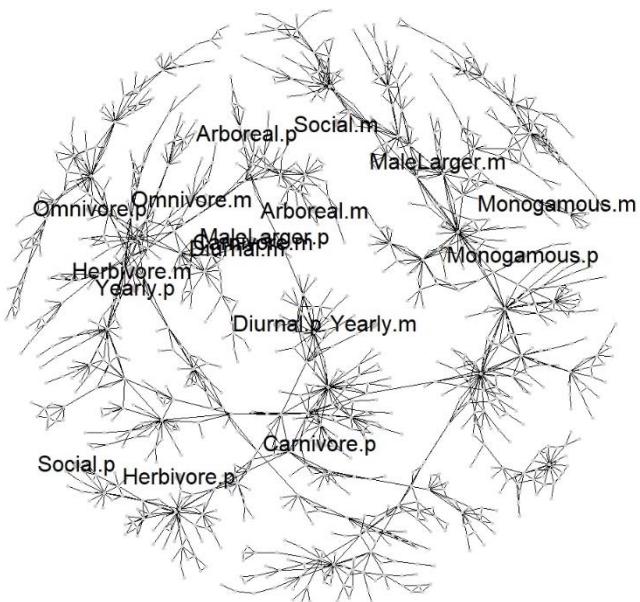


Bridging molecular evolution and phenotypic evolution

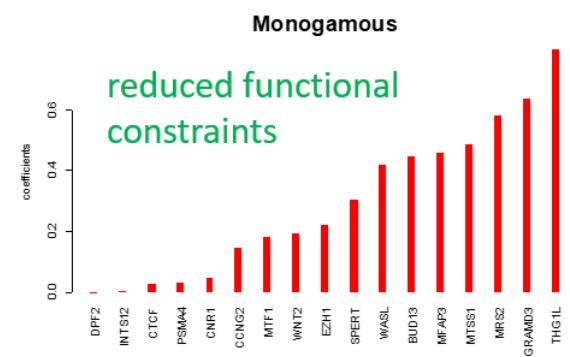


+

	Social	Diurnal	Yearly	Arboreal	Monogamous
Giant_panda	0	0	0.5	0	0
Night_monkey	0	0	0	0	0
Minke_whale	0	0	0	0	0
American_bison	1	0.5	1	0	0
Yak	0	0	0	0	0
Cattle	1	0	0	0	1
Buffalo	1	0	0	0	1
.....					



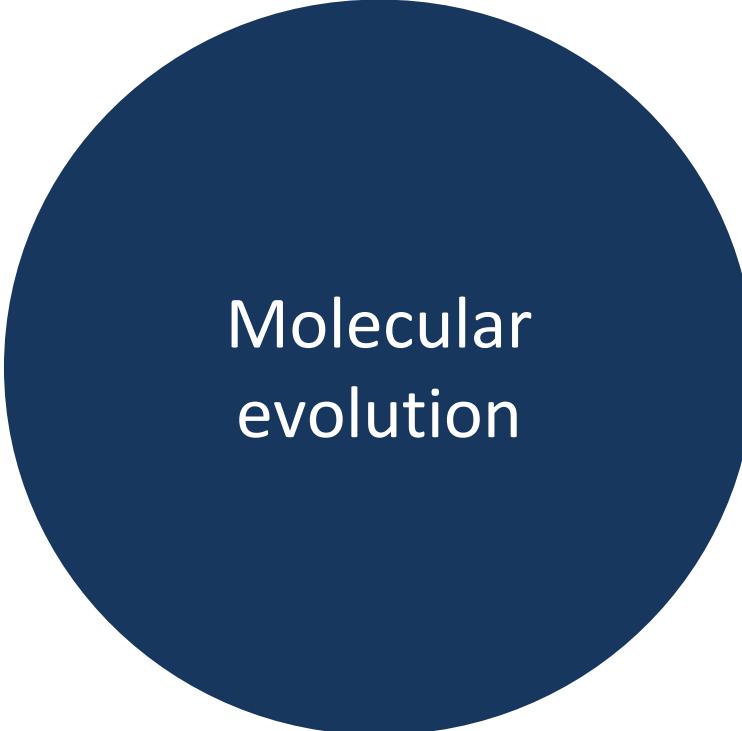
enhanced functional constraints



Wu et al (2017) Current Biology 27:
3025-3033.

Schedule of this course

Dates	Contents
15 May	Molecular evolution and phylogenetic tree
22 May	Rates of molecular evolution
29 May	Population structure and adaptation
5 May	Inferring traits evolution and selection

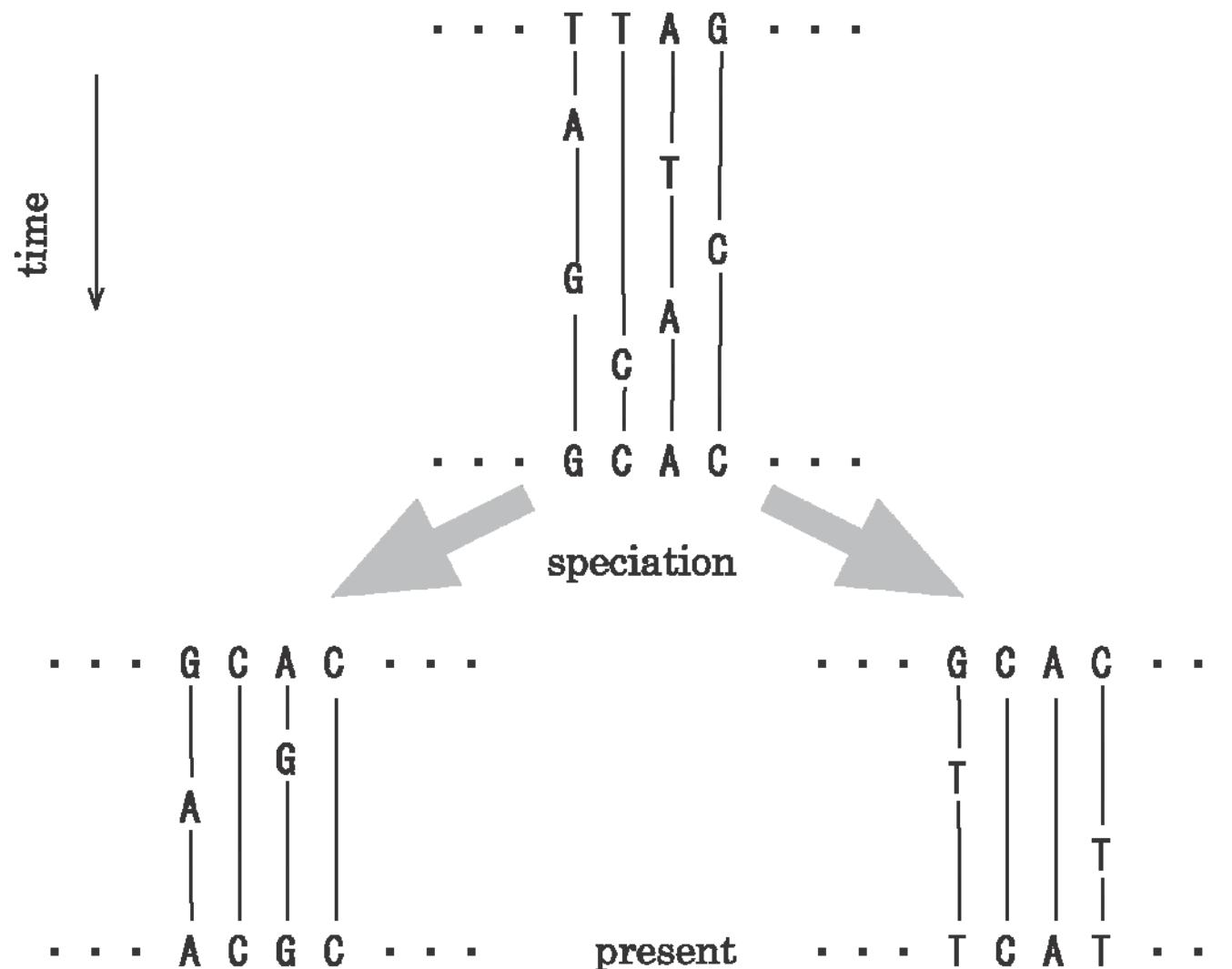


Molecular
evolution

The cost of natural selection

Unless selection is very intense, the number of deaths needed to secure the substitution, by natural selection, of one gene for another at a locus, is independent of the intensity of selection. It is about 30 times the number of organisms in a generation. It is suggested that, in horotelic evolution, the mean time taken for each gene substitution is about 300 generations. This accords with the observed slowness of evolution.

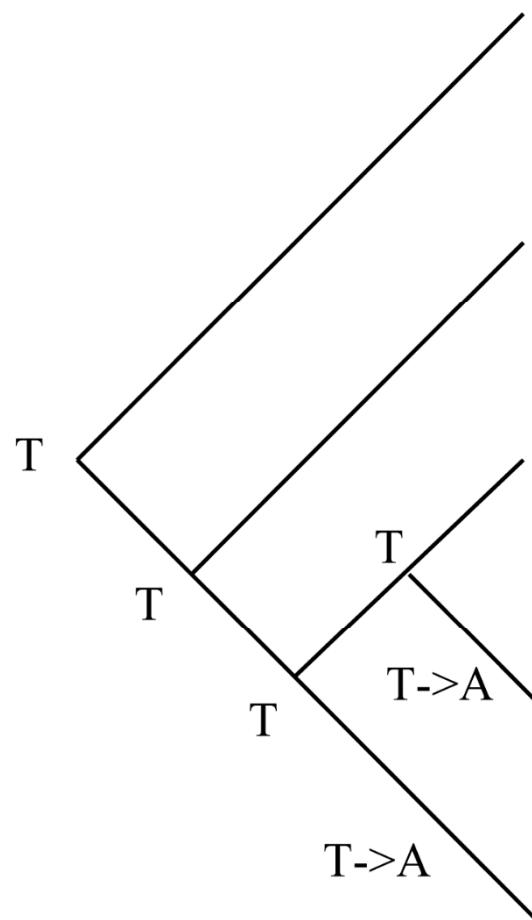
Large collection of molecular evolution provides unbiased estimate of species trees



Evolutionary Rate at the Molecular Level

Calculating the rate of evolution in terms of nucleotide substitutions seems to give a value so high that many of the mutations involved must be neutral ones.

Reocnstructing the evolutionary history: Criteria of minimum evolution



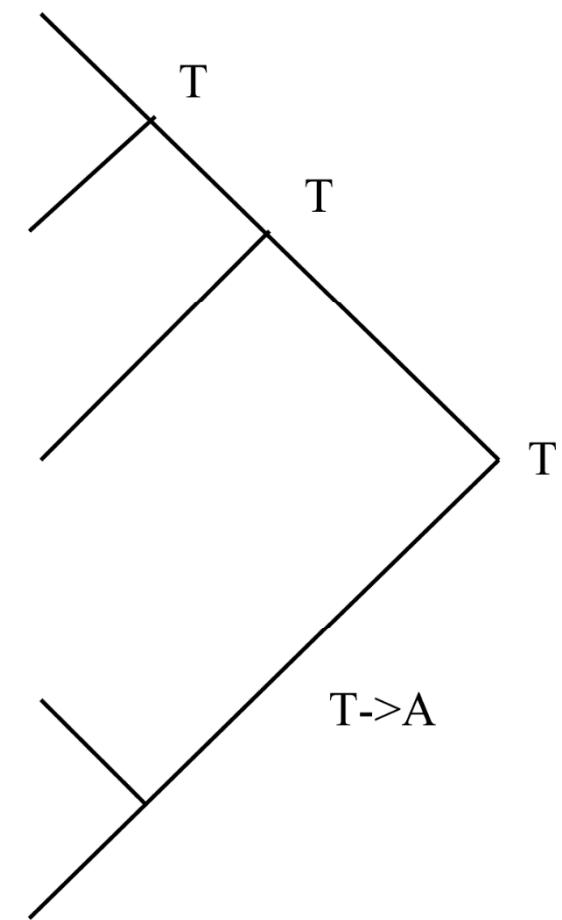
Species 1 : ... T ...

Species 2 : ... T ...

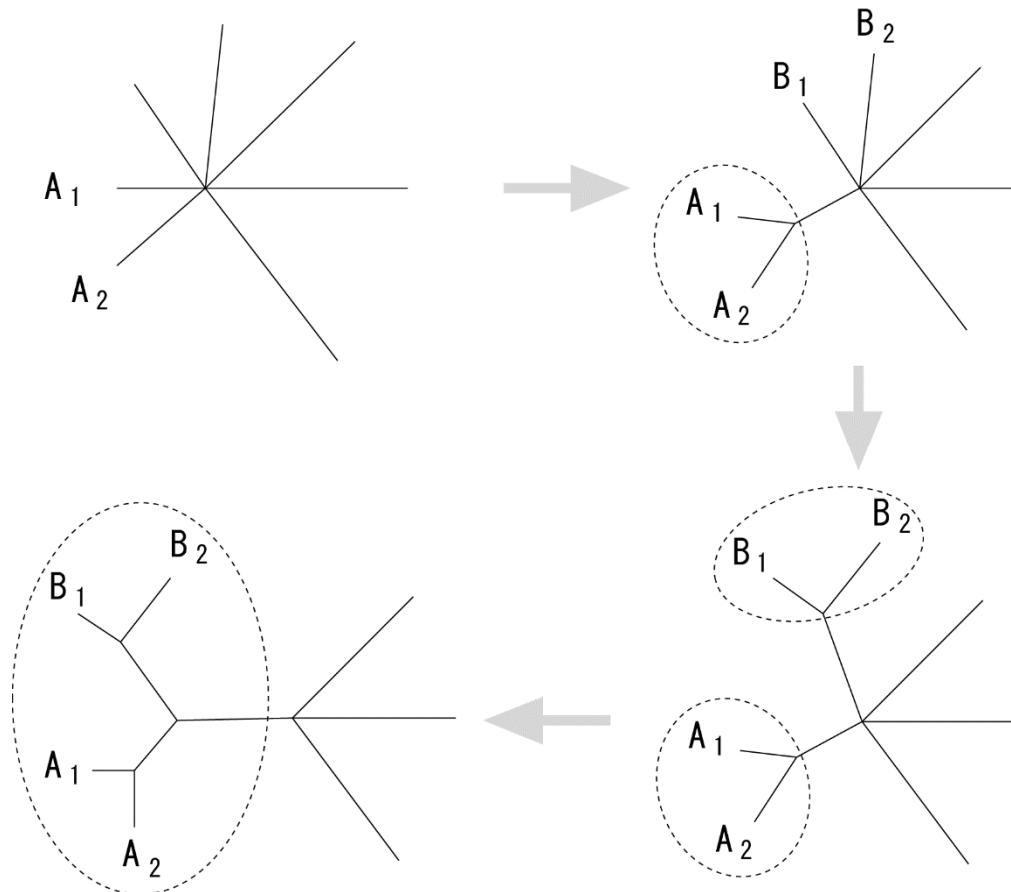
Species 3 : ... T ...

Species 4 : ... A ...

Species 5 : ... A ...



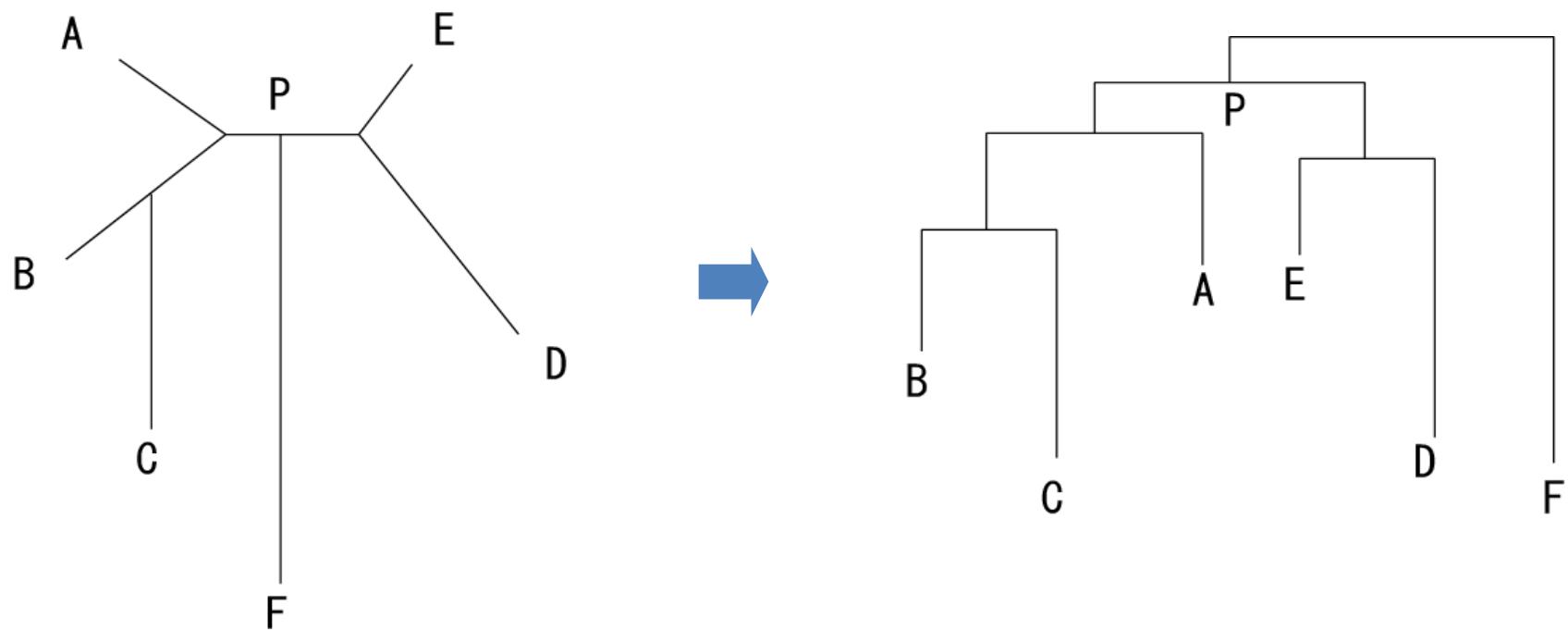
Neighbor joining method



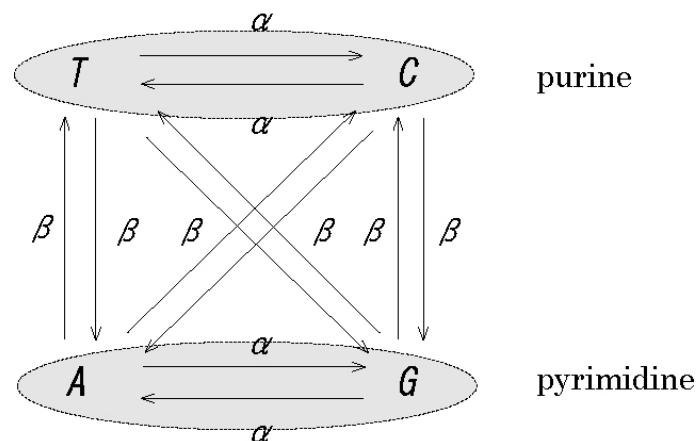
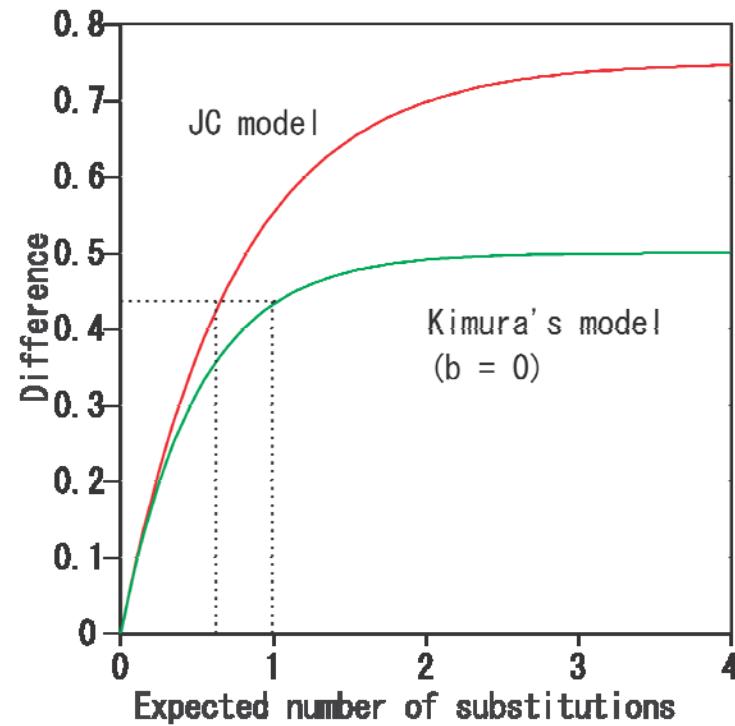
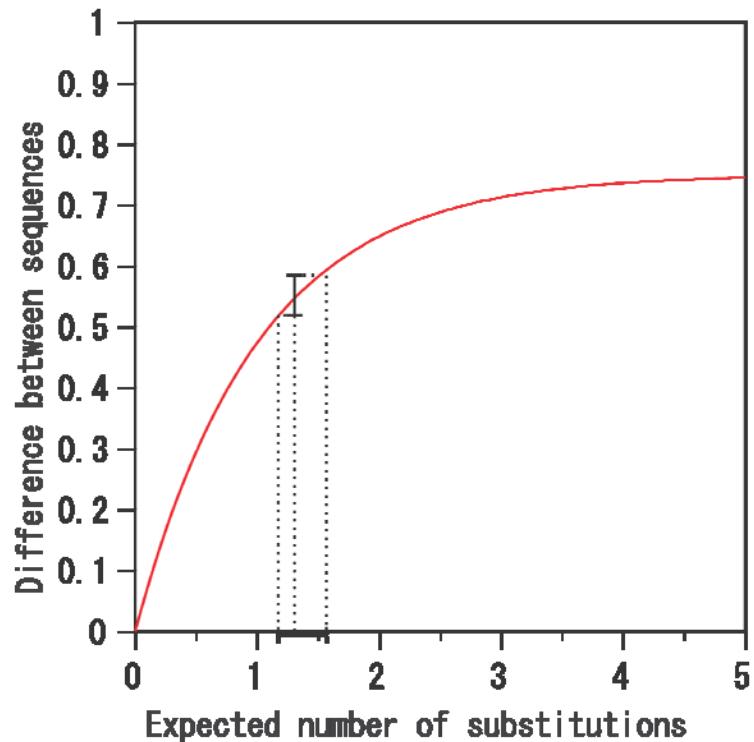
Minimizing the total branch length at each step

Saitou and Nei (1987) Mol Biol Evol. 4:
406-425.

Unrooted tree and rooting by an outgroup



Distance based on statistical model of substitutions



Estimating the pattern of substitutions from sequence comparison

..... **G T T C A C G G A T A C T G G T C T A C**

..... **A T T C A T G A A T A C C C G C C T G C**



$$p_{TT}, p_{TC}, p_{TA}, p_{TG}, p_{TT}, p_{CC}, p_{CA}, p_{CG}, p_{AA}, p_{AG}, p_{GG}$$

Modeling the proportions of the pattern of sites

$$p_{ij} = p_i \times p_{i \rightarrow j}(t)$$

Felsenstein (1981)

$$p_{ii}(t) = \exp(-ut) + (1 - \exp(-ut)) \pi_i$$

$$p_{ij}(t) = (1 - \exp(-ut)) \pi_j \quad (i \neq j) ,$$

Kimura (1981)

$$\begin{aligned} p_{TT}(t) &= p_{CC}(t) = p_{AA}(t) = p_{GG}(t) \\ &= \frac{1}{4} + \frac{1}{4} \exp(-4\beta t) + \frac{1}{2} \exp(-2(\alpha + \beta)t) \end{aligned}$$

$$\begin{aligned} p_{TC}(t) &= p_{CT}(t) = p_{AG}(t) = p_{GA}(t) \\ &= \frac{1}{4} + \frac{1}{4} \exp(-4\beta t) - \frac{1}{2} \exp(-2(\alpha + \beta)t) \end{aligned}$$

$$\begin{aligned} p_{TA}(t) &= p_{TG}(t) = p_{CA}(t) = p_{CG}(t) = p_{AT}(t) = p_{AC}(t) = p_{GT}(t) = p_{GC}(t) \\ &= \frac{1}{4} - \frac{1}{4} \exp(-4\beta t) . \end{aligned}$$

Models of nucleotide substitutions

Original	Mutant			
	A	T	C	G
1. Jukes-Cantor model:				
A	...	λ	λ	λ
T	λ	...	λ	λ
C	λ	λ	...	λ
G	λ	λ	λ	...
2. Felsenstein model:				
A	...	$\pi_T \lambda$	$\pi_C \lambda$	$\pi_G \lambda$
T	$\pi_A \lambda$...	$\pi_C \lambda$	$\pi_G \lambda$
C	$\pi_A \lambda$	$\pi_T \lambda$...	$\pi_G \lambda$
G	$\pi_A \lambda$	$\pi_T \lambda$	$\pi_C \lambda$...
3. Kimura model:				
A	...	β	β	α
T	β	...	α	β
C	β	α	...	β
G	α	β	β	...
4. Hasegawa <i>et al.</i> model:				
A	...	$\pi_T \beta$	$\pi_C \beta$	$\pi_G \alpha$
T	$\pi_A \beta$...	$\pi_C \alpha$	$\pi_G \beta$
C	$\pi_A \beta$	$\pi_T \alpha$...	$\pi_G \beta$
G	$\pi_A \alpha$	$\pi_T \beta$	$\pi_C \beta$...
5. Tamura-Nei model:				
A	...	$\pi_T \beta$	$\pi_C \beta$	$\pi_G \alpha_1$
T	$\pi_A \beta$...	$\pi_C \alpha_2$	$\pi_G \beta$
C	$\pi_A \beta$	$\pi_T \alpha_2$...	$\pi_G \beta$
G	$\pi_A \alpha_1$	$\pi_T \beta$	$\pi_C \beta$...
6. Rzhetsky-Nei model:				
A	...	β_2	β_3	α_4
T	β_1	...	α_3	β_4
C	β_1	α_2	...	β_4
G	α_1	β_2	β_3	...

Modeling the pattern of molecular evolution and the transition probabilities

Felsenstein (1981)

$$p_{ii}(t) = \exp(-ut) + (1 - \exp(-ut)) \pi_i$$

$$p_{ij}(t) = (1 - \exp(-ut)) \pi_j \quad (i \neq j) ,$$

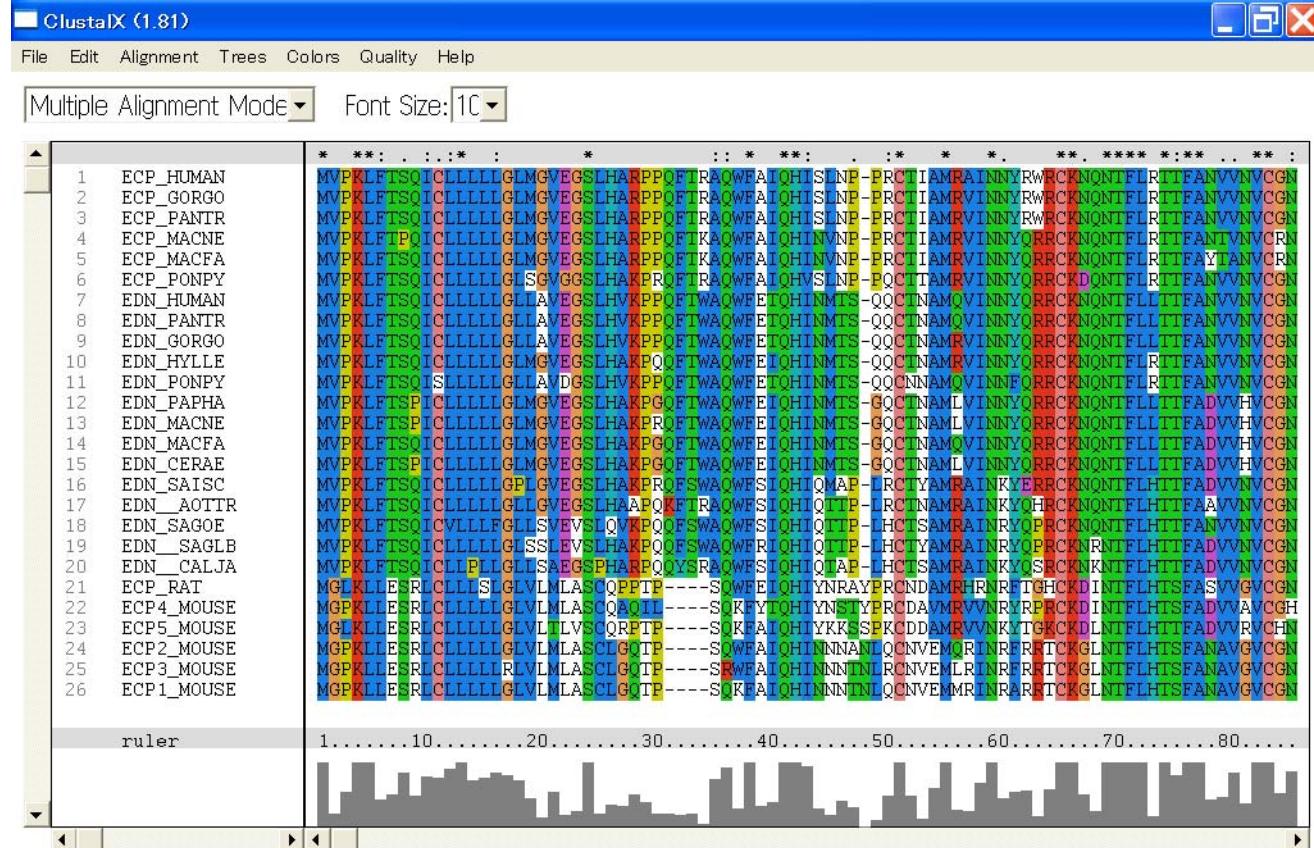
Kimura (1980)

$$\begin{aligned} p_{TT}(t) &= p_{CC}(t) = p_{AA}(t) = p_{GG}(t) \\ &= \frac{1}{4} + \frac{1}{4} \exp(-4\beta t) + \frac{1}{2} \exp(-2(\alpha + \beta)t) \end{aligned}$$

$$\begin{aligned} p_{TC}(t) &= p_{CT}(t) = p_{AG}(t) = p_{GA}(t) \\ &= \frac{1}{4} + \frac{1}{4} \exp(-4\beta t) - \frac{1}{2} \exp(-2(\alpha + \beta)t) \end{aligned}$$

$$\begin{aligned} p_{TA}(t) &= p_{TG}(t) = p_{CA}(t) = p_{CG}(t) = p_{AT}(t) = p_{AC}(t) = p_{GT}(t) = p_{GC}(t) \\ &= \frac{1}{4} - \frac{1}{4} \exp(-4\beta t) . \end{aligned}$$

Multiple alignment identifies rate heterogeneity among sites



CLUSTAL-Alignment file created



A note on maximum likelihood and parsimony: the case of complete observation

n_1, n_2, \dots, n_k : occurrence (1) / non-occurrence (0) of evolutionary events along the branches 1,2,...,k

- Assuming no multiple evolutionary events along a branch
- Likelihood

$$L(b|n_1, n_2, \dots, n_k) = \prod_{i=1}^k b^{n_i} (1-b)^{1-n_i}$$

$b_1 = b_2 = \dots = b_k = b$: probability of occurrence along a branch

- Assuming rare occurrence: $\lambda = kb$

$$L(\lambda|n_T) = e^{-\lambda} \frac{\lambda^{n_T}}{n_T!} \quad n_T = n_1 + n_2 + \dots + n_k:$$

the number of evolutionary events assuming tree T

A note on maximum likelihood and parsimony: the case of complete observation

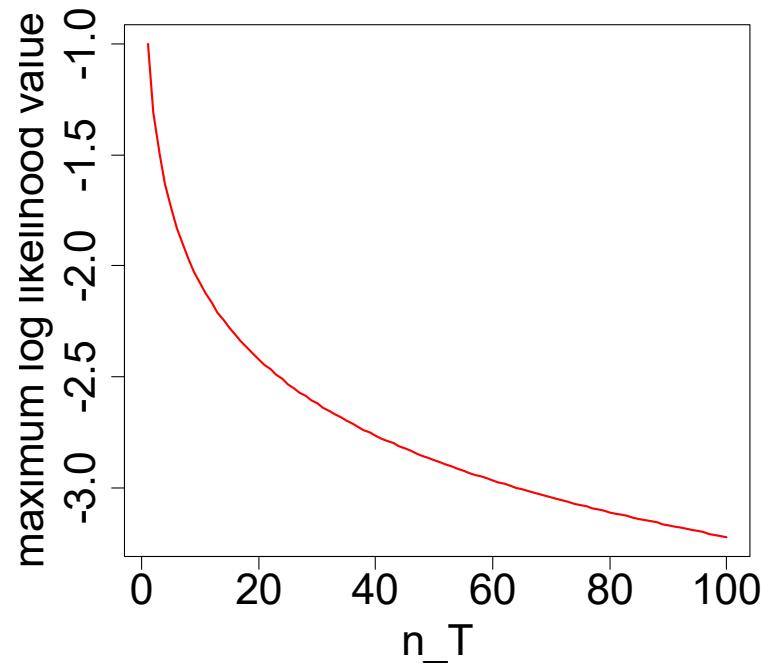
- Maximum likelihood estimate and parsimony

$$\hat{\lambda} = n_T$$

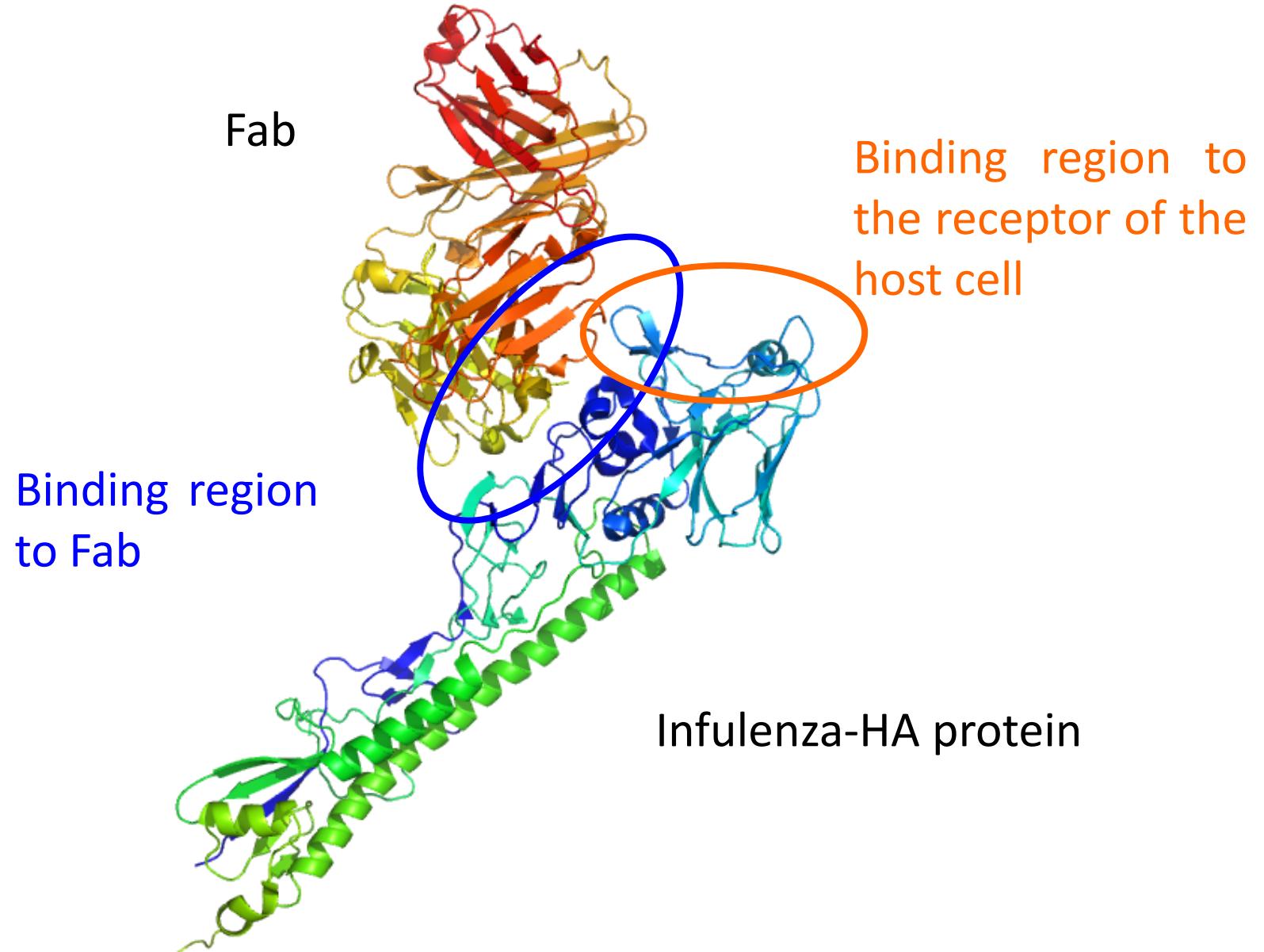
$$\text{logL}(\hat{\lambda}|n_T) = -\hat{\lambda} + n_T \log \hat{\lambda} - \sum_{i=1}^{n_T} \log i$$

$$= -n_T + n_T \log n_T - \sum_{i=1}^{n_T} \log i$$

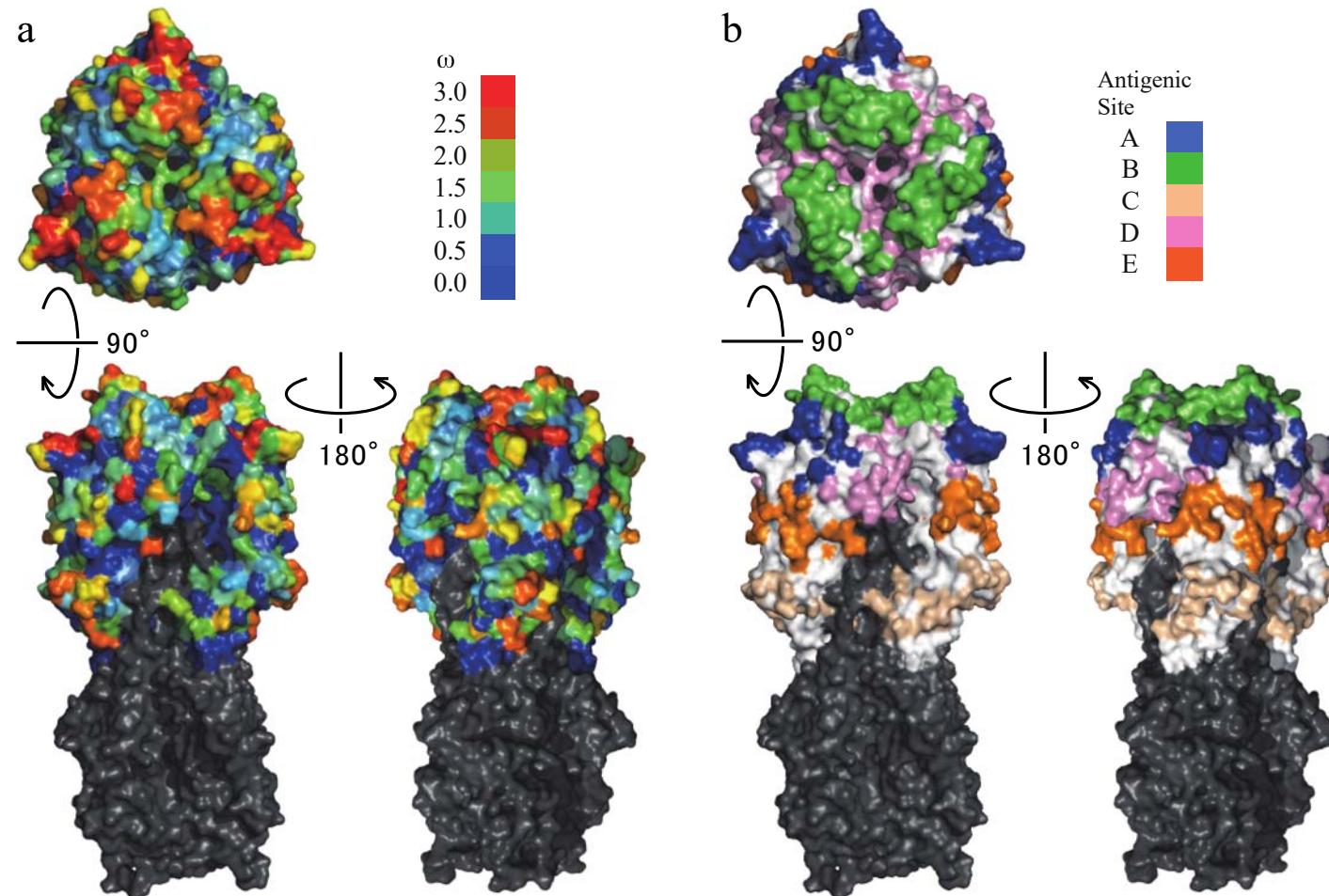
Likelihood approach is consistent with the criterion of minimum evolution



Selection pressure varies among the region of the protein



Regions under the diversifying selection and antigenic sites



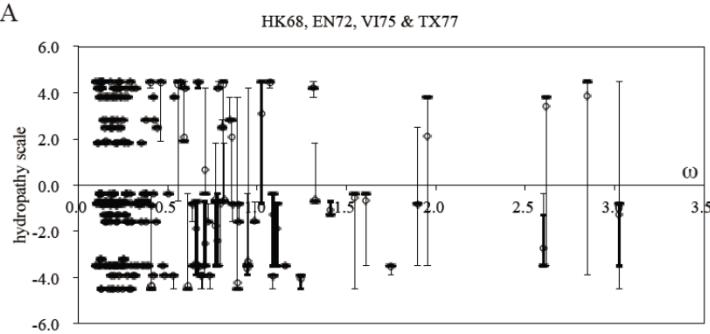
The spatial distribution of selection pressure

The distributions of the antigenic sites A to E.

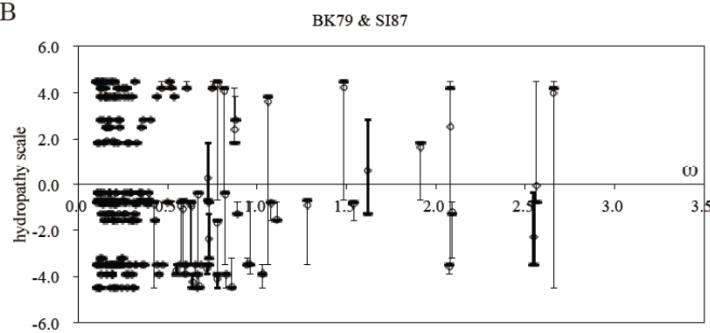
Watabe and Kishino (2013) Mol Biol Evol. 30: 2714-2722.

Changes in chemico-physical feature and the ω values of substitutions

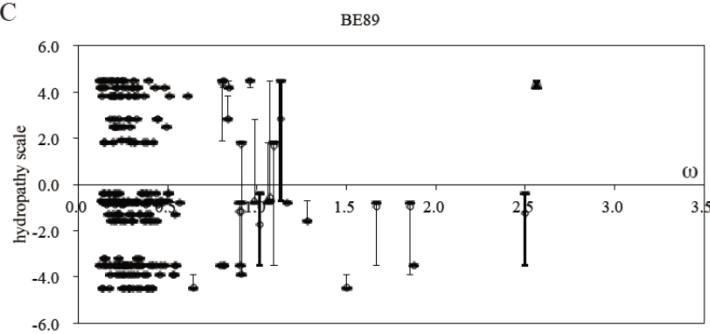
A



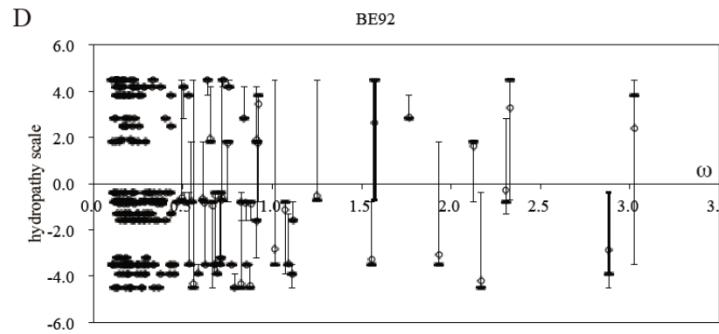
B



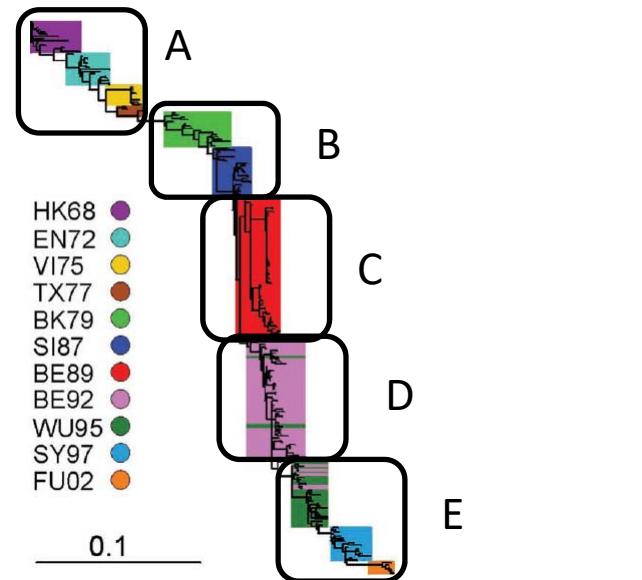
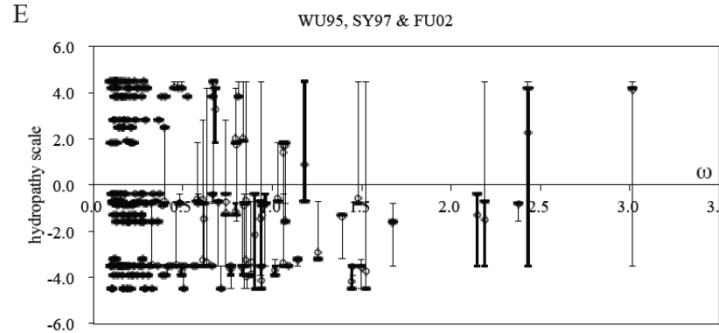
C



D



E



Rate of molecular evolution, mutation rate, and fixation rate

- Fitness of mutations: advantageous (rare), neutral, deleterious
- Mutations deleted from the populations are not observed by comparing the genomes
- Molecular evolution: mutations fixed to the populations

$$r = \mu \times 2N \times f$$

neutral

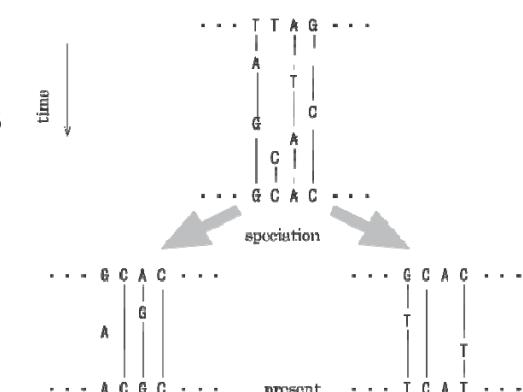
p

$$= \mu \times 2N \times p \times \frac{1}{2N}$$

deleterious

$1 - p$

$$= \mu p$$

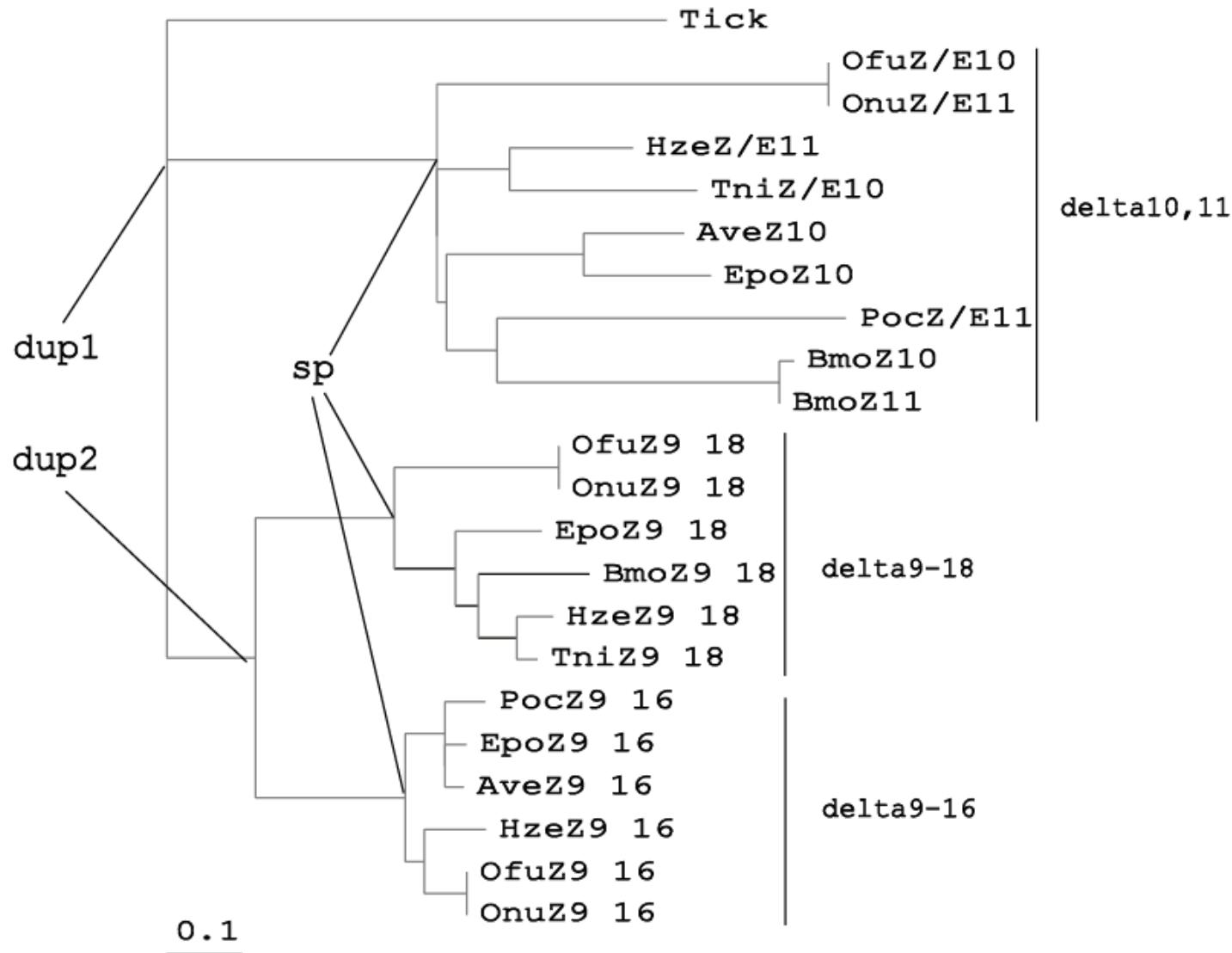


- Rate of molecular evolution is the product of **mutation rate** and **proportion of neutral mutations**

Generation length, mutagens
(affects on the whole genome)

Functional constraints, diversifying selection
(vary among genes in the genome)

Desaturase and the origin of sex pheromone



Analyzing
desaturase
sequences

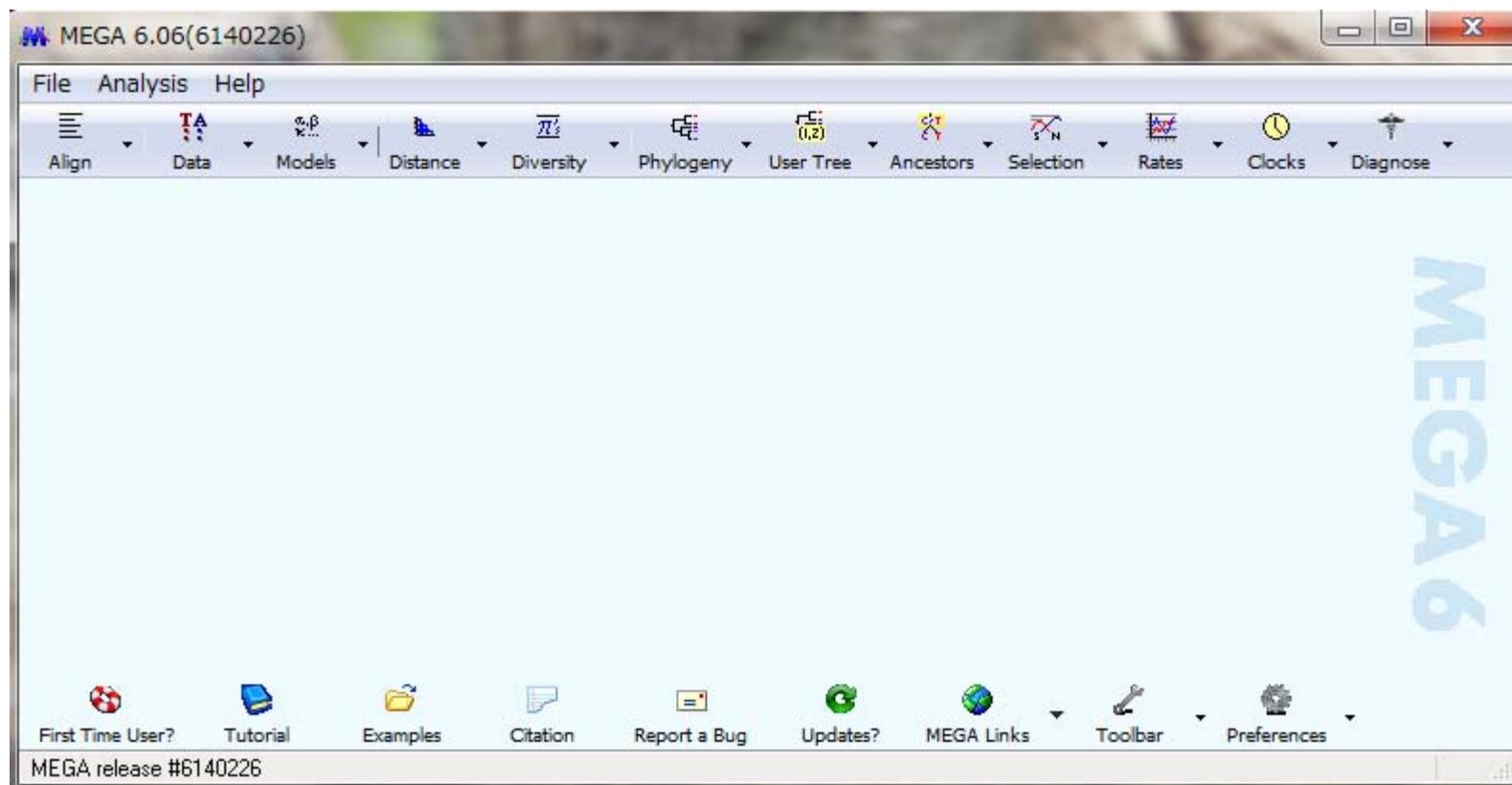
Amino acid sequences of desaturase (fasta format)

```
>HzeZ9_18
MPPQGQTGGSWVL YETDAVNEDTDAPVIVPPSAEKREWKIVWRNVILMGMLHIGGVYGA
Y LFLTTAMWRTCIFAVVLYICSGLGITAGAHLWAHKSYKARLPLRLMLTLFNTLAFQDA
V IDWARDHRMHMKYSETDADPHNATRGFFFHVGWLLVRKHPQIKAKGHTIDSDLKSDPI
L RFQKKYYLFLMPLVCFILPCYIPT-LWGESLNAYFVCSIFRYVYVLNVTWLVNSAAHL
W GAKPYDKNINPVETRPVSLVVLGEGFHNYHHTFPWDYKTAELGDYSLNLTKLFIDTMAA
I GWAYDLKTVSTDVIQKRVKRTGDGSHPVWGDDHEVHQADKKLAAIINPEKT
>TniZ9_18
MPPQGQTGGSWVL YETDAVNNTDAPVIVPPSAEKREWKIVWRNVILMGMLHIGGVYGA
Y LFLTKAMWLTDLFAFFLYLCSGLGITAGAHLWAHKSYKARLPLRLLLTLFNTLAFQDA
V IDWARDHRMHMKYSETDADPHNATRGFFFHVGWLLVRKHPQIKAKGHTIDSDLKSDPI
L RFQKKYYLTLMPICFILPSYIPT-LWGEAFNAFFVCSIFRYVYVLNVTWLVNSAAHL
W GSCKPYDKNINPVETRPVSLVVLGEGFHNYHHTFPWDYKTAELGDYSLNFTKMFIDFMA
I GWAYDLKTVSTDVIQKRVKRTGDGSHAVWGDDHEVHQEDKKLAAIINPEKT
>BmoZ9_18
MPPQRKQEASWVL YEADANNLPEDAPPVPPSAEKRPWKIVWRNVILFFILHVGGVYGGY
Y LFLFKAMWRTSIFAIFLYLCSGLGITAGAHLWAHKSYKARLPLRILLTIFNTIAFQDA
V VDWARDHRMHMKYSETDADPHNATRGFFFHVGWLLVRKHPQIKAKGHTDVNELRNDPI
L RFQKKYYQILMPLACFIMPTYVPT-LWGETVWNSFYVCAIFRYVYVLNITWLVNSAAHM
W GSCKPYDKNINPVETRPVSLVVLGEGFHNYHHTFPWDYKTAELGDYSLNLSKLFIDFMAK
I DWAYDLKTVSTDVIQKRTKRTGDGSHPVWGVDVGEVATEDKTTNLVNSKV
>EpoZ9_18
MPPQGQPAAWVLEESDATDDKDVAVALPPSAEKRKLSIVWRNVILFVLLHTGAVYGGY
Y LFFTAKMWWATKFFAFFLYLCSGLGITAGAHLWAHKSYKARLPLRILLTLFNTIAFQDSV
L DWARDHRMHMKYSETDADPHNATRGFFFHVGWLLVRKHPQIKAKGHTIDMSDLCSDPV
L RFQKKYYLTLMPLFCAFILPTYIPT-LWGESENAYFVAAIFRYCYVLNVTWLVNSAAHK
W GDRPYDKNINPVETKPVSLVVFGEFHNYHHTFPWDYKTAELGGYSLNISKLFIDTMAK
I GWAYDLKSVSPDIVEKRVKRTGDGSHEVWGDDKDVPAEQKAAATIINPEKT
.
```

desaturase.fasta

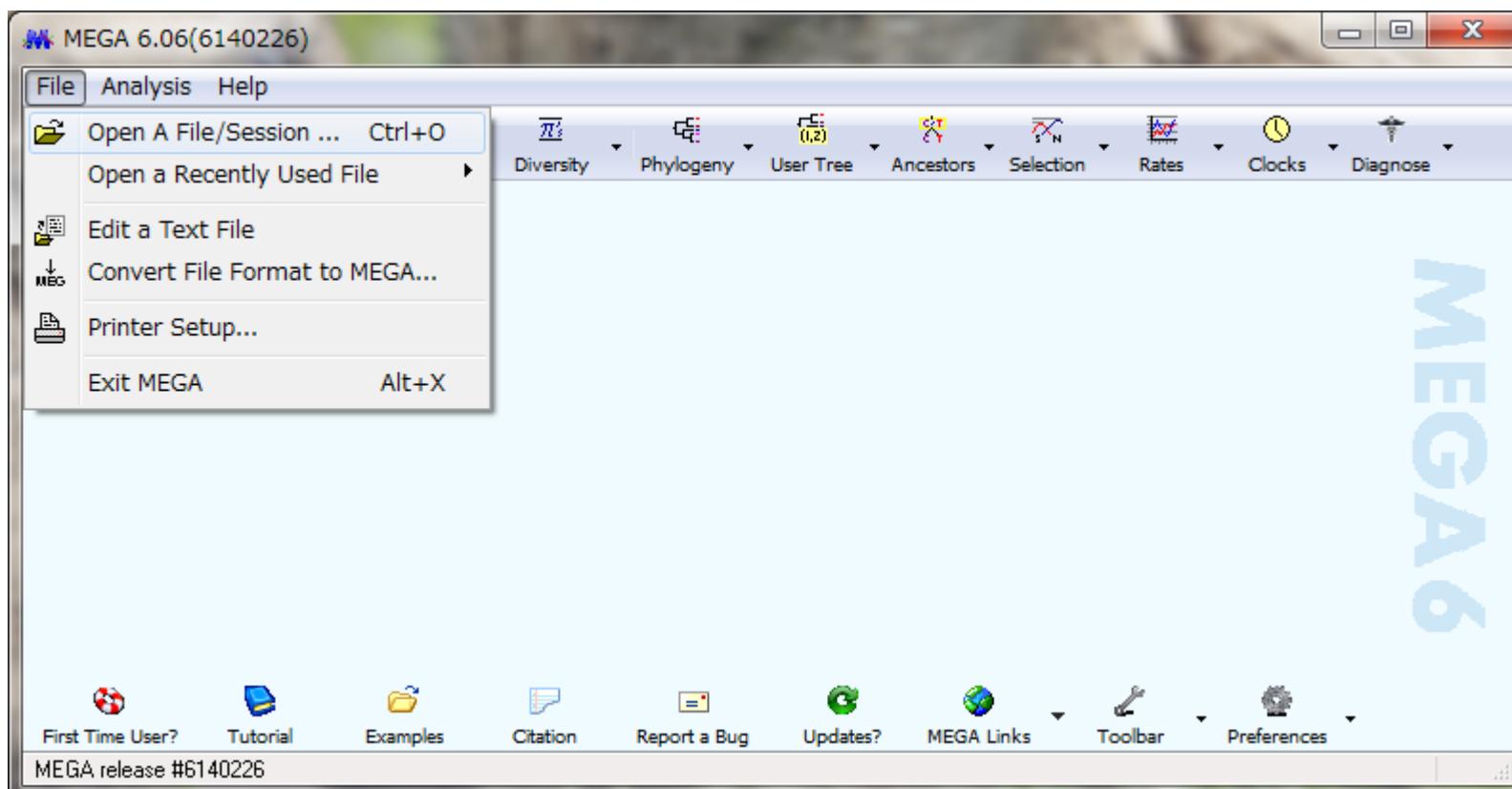
Start MEGA

Double click the shortcut



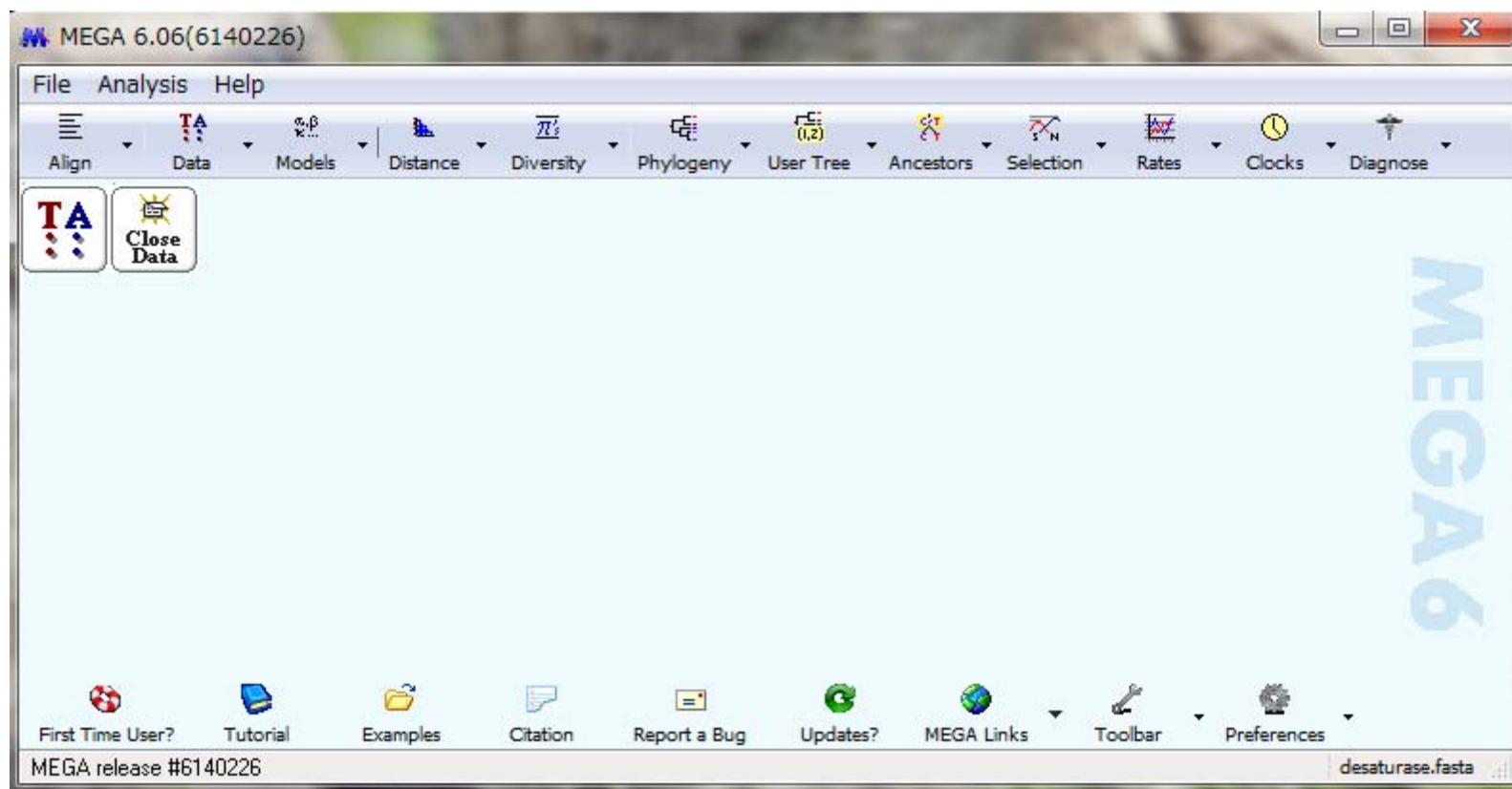
Open the file of amino acid sequences

[File][Open A File/Session]



Open the file of amino acid sequences

"Analyze" -> "Protein Sequence"



View the data

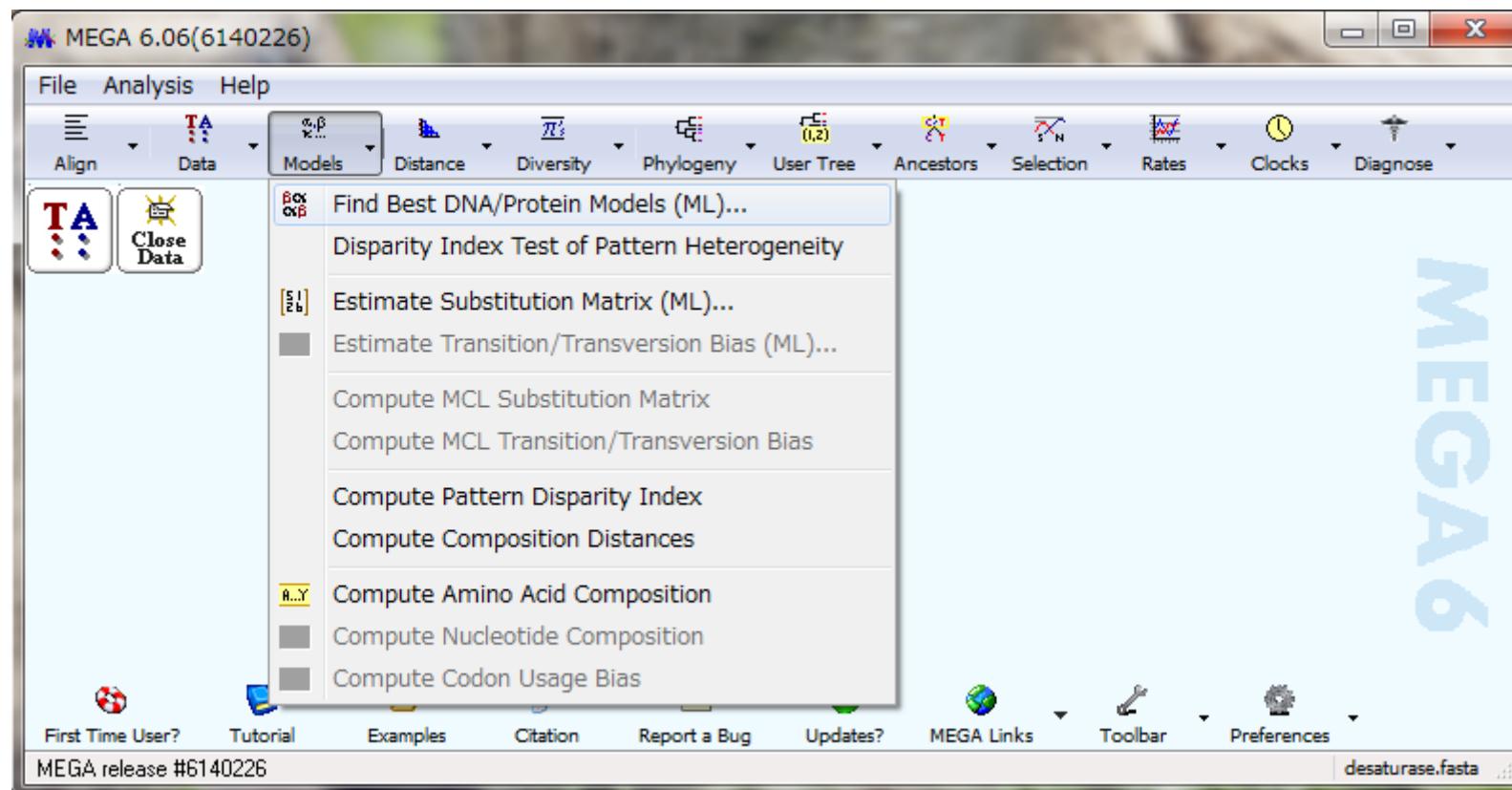
Double click



The screenshot shows the MEGA 6.06 software interface. The main window title is "MEGA 6.06(6140226)" and the sub-window title is "M6: Sequence Data Explorer". The menu bar includes File, Analysis, Data, and Help. The toolbar contains various icons for file operations like Open, Save, and Print, along with sequence analysis tools like UUC, C, V, Pi, S, and Special. The main area displays a sequence alignment table with 18 rows of data. Each row has a checkbox at the beginning and a sample name followed by a group identifier. The columns represent amino acid positions. A red box highlights the "Align" button in the toolbar, and a red arrow points from it to the alignment table.

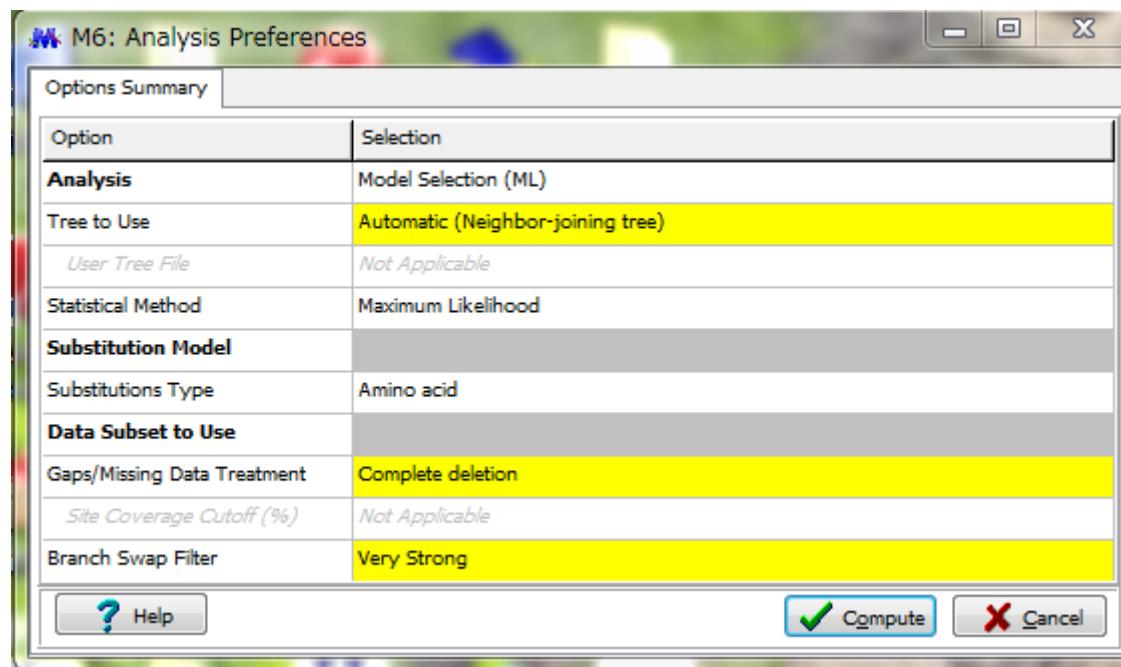
Comparing the models of amino acid replacements

[Models][Find Best DNA/Protein Models (ML)]



Comparing the models of amino acid replacements

List of options

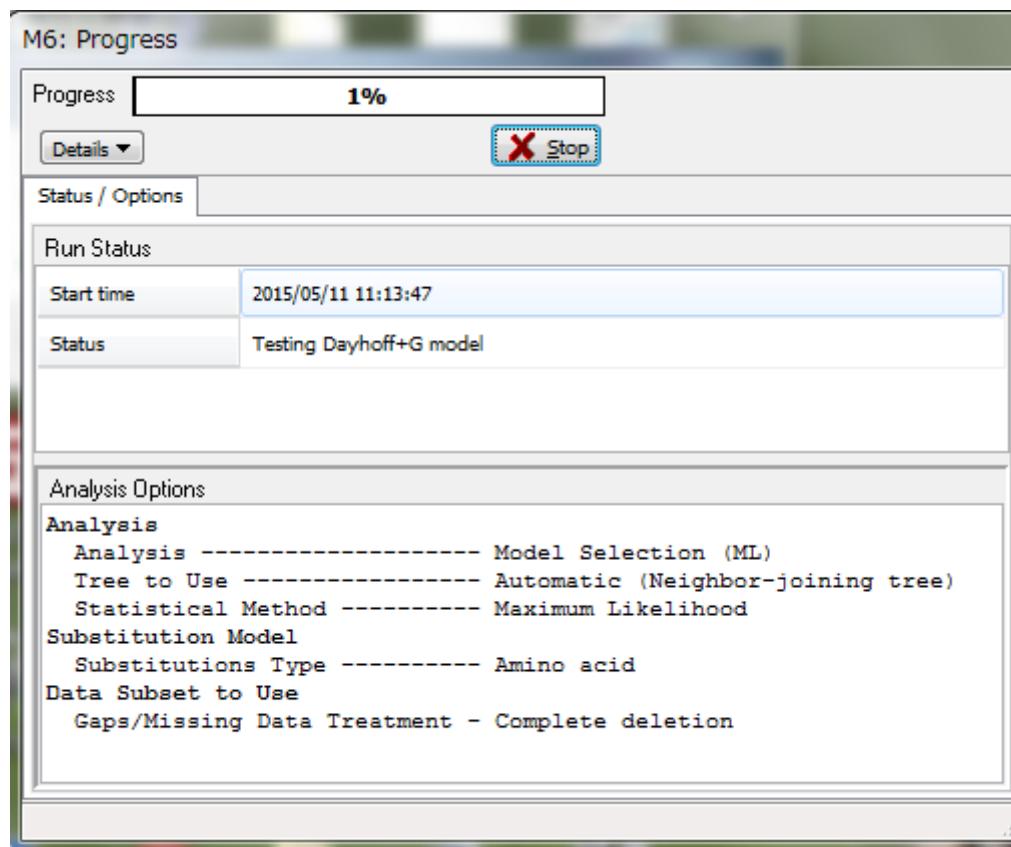


You can change the options of the items colored yellow.

Here, keep the default values, and click [Compute].

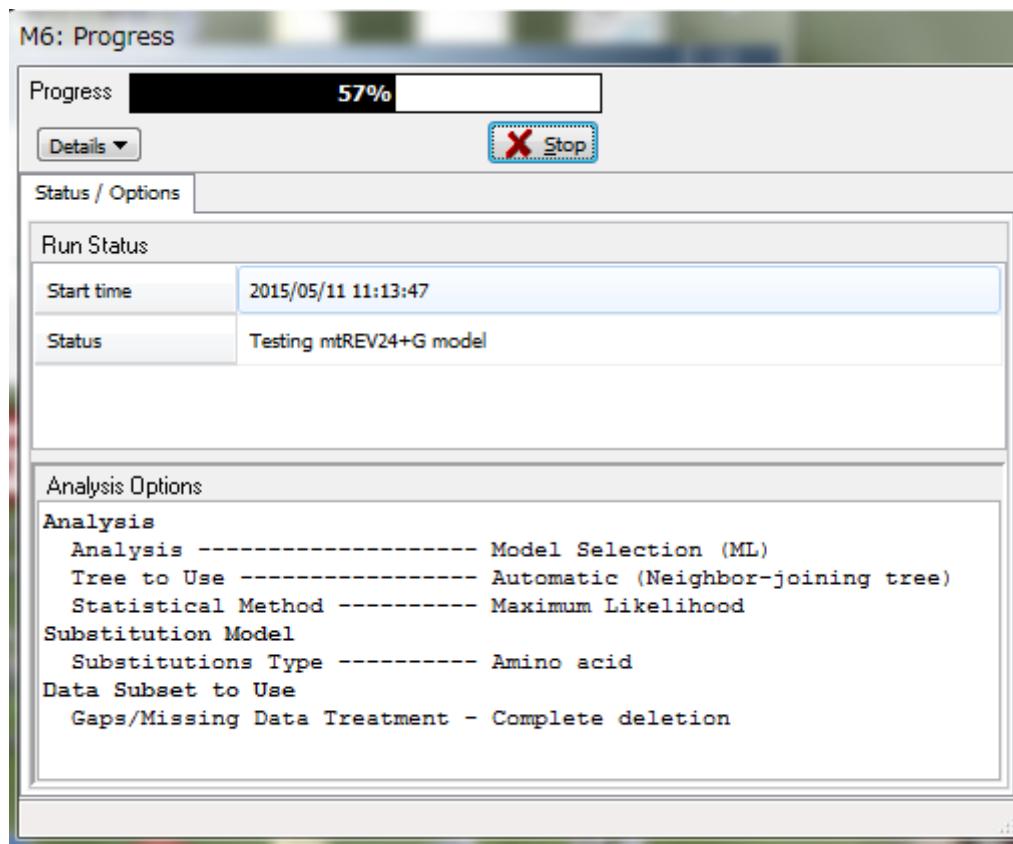
Comparing the models of amino acid replacements

You will see the progress of the analysis



Comparing the models of amino acid replacements

You will see the progress of the analysis



Because various models of aa replacements are applied, it takes some time.

Be patient..

Comparing the models of amino acid replacements

Once finished, you will see the list of models ranked with BIC.

MEGA Caption Expert: Find Best-Fit Substitution Model (ML)

File Edit View Help

XL CSV

Table. Maximum Likelihood fits of 48 different amino acid substitution models																
Model	Parameters	BIC	AICc	lnL	(+I)	(+G)	f(A)	f(R)	f(N)	f(D)	f(C)	f(Q)	f(E)	f(G)	f(H)	f(I)
LG+G	42	11957.554	11670.512	-5792.995	n/a	0.70	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022	0.022
LG+G+I	43	11960.402	11666.538	-5789.995	0.15	1.14	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022	0.022
WAG+G+I	43	12046.125	11752.261	-5832.857	0.17	1.37	0.087	0.044	0.039	0.057	0.019	0.037	0.058	0.083	0.024	0.024
WAG+G	42	12046.744	11759.702	-5837.590	n/a	0.75	0.087	0.044	0.039	0.057	0.019	0.037	0.058	0.083	0.024	0.024
cpREV+G	42	12068.733	11781.691	-5848.584	n/a	0.72	0.076	0.062	0.041	0.037	0.009	0.038	0.050	0.084	0.025	0.025
cpREV+G+I	43	12072.005	11778.141	-5845.797	0.16	1.17	0.076	0.062	0.041	0.037	0.009	0.038	0.050	0.084	0.025	0.025
rtREV+G	42	12072.014	11784.972	-5850.225	n/a	0.72	0.065	0.045	0.038	0.042	0.011	0.061	0.061	0.064	0.027	0.027
rtREV+G+I	43	12074.609	11780.745	-5847.099	0.15	1.18	0.065	0.045	0.038	0.042	0.011	0.061	0.061	0.064	0.027	0.027
JTT+G	42	12091.113	11804.071	-5859.774	n/a	0.70	0.077	0.051	0.043	0.051	0.020	0.041	0.062	0.075	0.023	0.023
JTT+G+I	43	12091.885	11798.021	-5855.737	0.16	1.23	0.077	0.051	0.043	0.051	0.020	0.041	0.062	0.075	0.023	0.023
LG+G+F	61	12102.453	11685.897	-5781.400	n/a	0.72	0.083	0.045	0.045	0.048	0.010	0.019	0.029	0.060	0.050	0.050
LG+G+I+F	62	12104.561	11681.194	-5778.030	0.15	1.19	0.083	0.045	0.045	0.048	0.010	0.019	0.029	0.060	0.050	0.050
rtREV+G+F	61	12149.273	11732.717	-5804.809	n/a	0.72	0.083	0.045	0.045	0.048	0.010	0.019	0.029	0.060	0.050	0.050
LG+I	42	12151.011	11863.969	-5889.723	0.25	n/a	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022	0.022

Comparing the models of amino acid replacements

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MEGA Caption Expert: Find Best-Fit Substitution Model (ML)

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Model	Parameters	BIC	AICc	lnL	(+I)	(+G)	f(A)	f(R)	f(N)	f(D)	f(C)	f(Q)	f(E)	f(G)	f(H)
LG+G	42	11957.554	11670.512	-5792.995	n/a	0.70	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
LG+G+I	43	11960.402	11666.538	-5789.995	0.15	1.14	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
WAG+G+I	43	12046.125	11752.261	-5832.857	0.17	1.37	0.087	0.044	0.039	0.057	0.019	0.037	0.058	0.083	0.024
WAG+G	42	12046.744	11759.702	-5837.590	n/a	0.75	0.087	0.044	0.039	0.057	0.019	0.037	0.058	0.083	0.024
cpREV+G	42	12068.733	11781.691	-5848.584	n/a	0.72	0.076	0.062	0.041	0.037	0.009	0.038	0.050	0.084	0.025
cpREV+G+I	43	12072.005	11778.141	-5845.797	0.16	1.17	0.076	0.062	0.041	0.037	0.009	0.038	0.050	0.084	0.025
rtREV+G	42	12072.014	11784.972	-5850.225	n/a	0.72	0.065	0.045	0.038	0.042	0.011	0.061	0.061	0.064	0.027
rtREV+G+I	43	12074.609	11780.745	-5847.099	0.15	1.18	0.065	0.045	0.038	0.042	0.011	0.061	0.061	0.064	0.027
JTT+G	42	12091.113	11804.071	-5859.774	n/a	0.70	0.077	0.051	0.043	0.051	0.020	0.041	0.062	0.075	0.023
JTT+G+I	43	12091.885	11798.021	-5855.737	0.16	1.23	0.077	0.051	0.043	0.051	0.020	0.041	0.062	0.075	0.023
LG+G+F	61	12102.453	11685.897	-5781.400	n/a	0.72	0.083	0.045	0.045	0.048	0.010	0.019	0.029	0.060	0.050
LG+G+I+F	62	12104.561	11681.194	-5778.030	0.15	1.19	0.083	0.045	0.045	0.048	0.010	0.019	0.029	0.060	0.050
rtREV+G+F	61	12149.273	11732.717	-5804.809	n/a	0.72	0.083	0.045	0.045	0.048	0.010	0.019	0.029	0.060	0.050
LG+I	42	12151.011	11863.969	-5889.723	0.25	n/a	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022

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MEGA Caption Expert: Find Best-Fit Substitution Model (ML)

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Proportion of invariant sites, shape parameter of the gamma distribution ...

Model	Parameters	BIC	AICc	lnL	(+I)	(+G)	f(A)	f(R)	f(N)	f(D)	f(C)	f(Q)	f(E)	f(G)	f(H)
LG+G	42	11957.554	11670.512	-5792.995	n/a	0.70	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
LG+G+I	43	11960.402	11666.538	-5789.995	0.15	1.14	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
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Comparing the models of amino acid replacements

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MEGA Caption Expert: Find Best-Fit Substitution Model (ML)

File Edit View Help

XL CSV

Proportion of invariant sites, shape parameter of the gamma distribution ...

Model	Parameters	BIC	AICc	lnL	(+I)	(+G)	f(A)	f(R)	f(N)	f(D)	f(C)	f(Q)	f(E)	f(G)	f(H)	
Best model	LG+G	42	11957.554	11670.512	-5792.995	n/a	0.70	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
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	WAG+G	42	12046.744	11759.702	-5837.590	n/a	0.75	0.087	0.044	0.039	0.057	0.019	0.037	0.058	0.083	0.024
	cpREV+G	42	12068.733	11781.691	-5848.584	n/a	0.72	0.076	0.062	0.041	0.037	0.009	0.038	0.050	0.084	0.025
	cpREV+G+I	43	12072.005	11778.141	-5845.797	0.16	1.17	0.076	0.062	0.041	0.037	0.009	0.038	0.050	0.084	0.025
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MEGA Caption Expert: Find Best-Fit Substitution Model (ML)

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Proportion of invariant sites, shape parameter of the gamma distribution ...

Model	Parameters	BIC	AICc	lnL	(+I)	(+G)	f(A)	f(R)	f(N)	f(D)	f(C)	f(Q)	f(E)	f(G)	f(H)	
Best model	LG+G	42	11957.554	11670.512	-5792.995	n/a	0.70	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
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	WAG+G															
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	LG+I	42	12151.011	11863.969	-5889.723	0.25	n/a	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022

Comparing the models of amino acid replacements

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MEGA Caption Expert: Find Best-Fit Substitution Model (ML)

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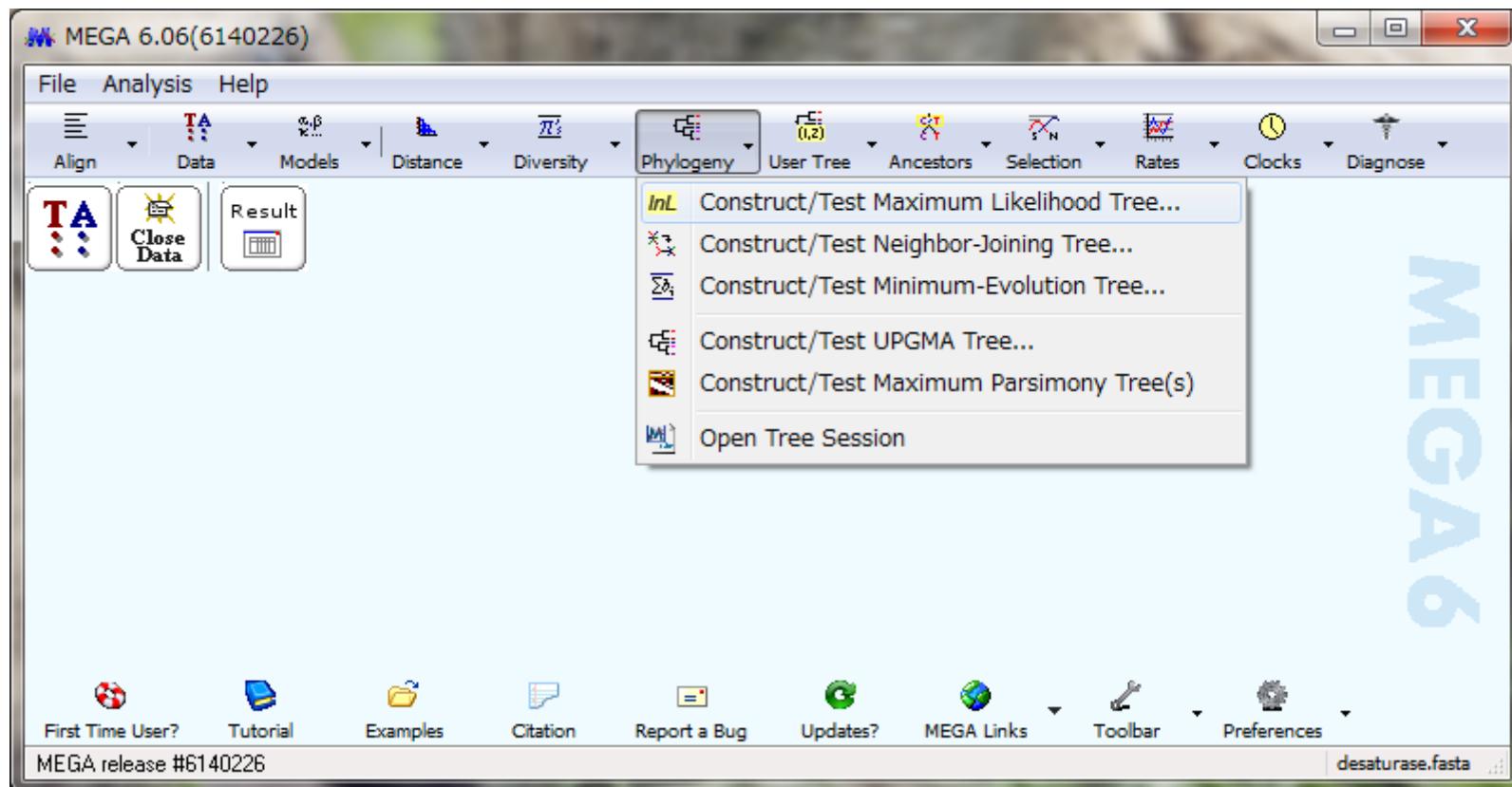
Proportion of invariant sites, shape parameter of the gamma distribution ...

Model	Parameters	BIC	AICc	lnL	(+I)	(+G)	f(A)	f(R)	f(N)	f(D)	f(C)	f(Q)	f(E)	f(G)	f(H)	
Best model	LG+G	42	11957.554	11670.512	-5792.995	n/a	0.70	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
	LG+G+I	43	11960.402	11666.538	-5789.995	0.15	1.14	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
	WAG+G+I	43	12046.125	11752.261	-5832.857	0.17	1.37	0.087	0.044	0.039	0.057	0.019	0.037	0.058	0.083	0.024
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	rtREV+G+I	43	12074.609	11780.745	-5847.099	0.15	1.18	0.065	0.045	0.038	0.042	0.011	0.061	0.061	0.064	0.027
	JTT+G	42	12091.113	11804.071	-5859.774	n/a	0.70	0.077	0.051	0.043	0.051	0.020	0.041	0.062	0.075	0.023
	JTT+G+I	43	12091.885	11798.021	-5855.737	0.16	1.23	0.077	0.051	0.043	0.051	0.020	0.041	0.062	0.075	0.023
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Maximum
likelihood
inference

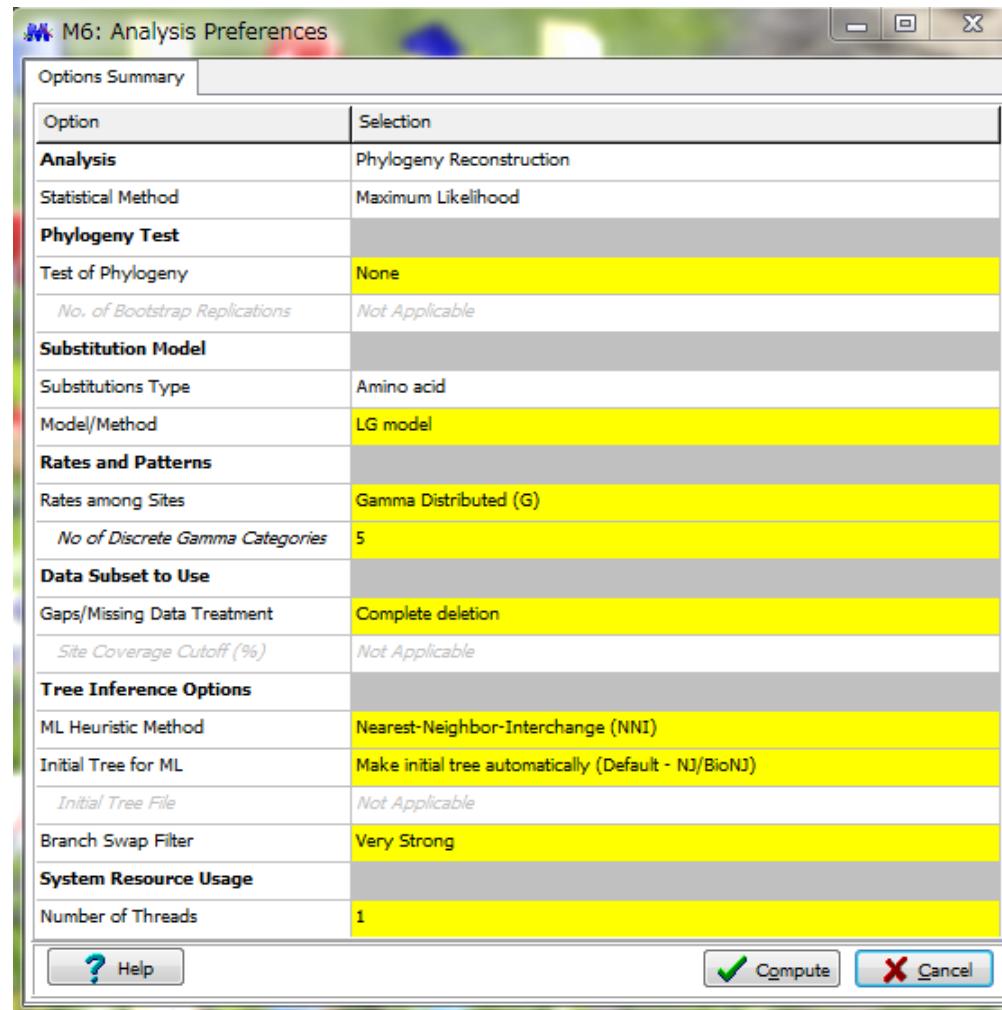
Constructing a phylogenetic tree by the best model

[Phylogeny][Construct/Test Maximum Likelihood Tree]



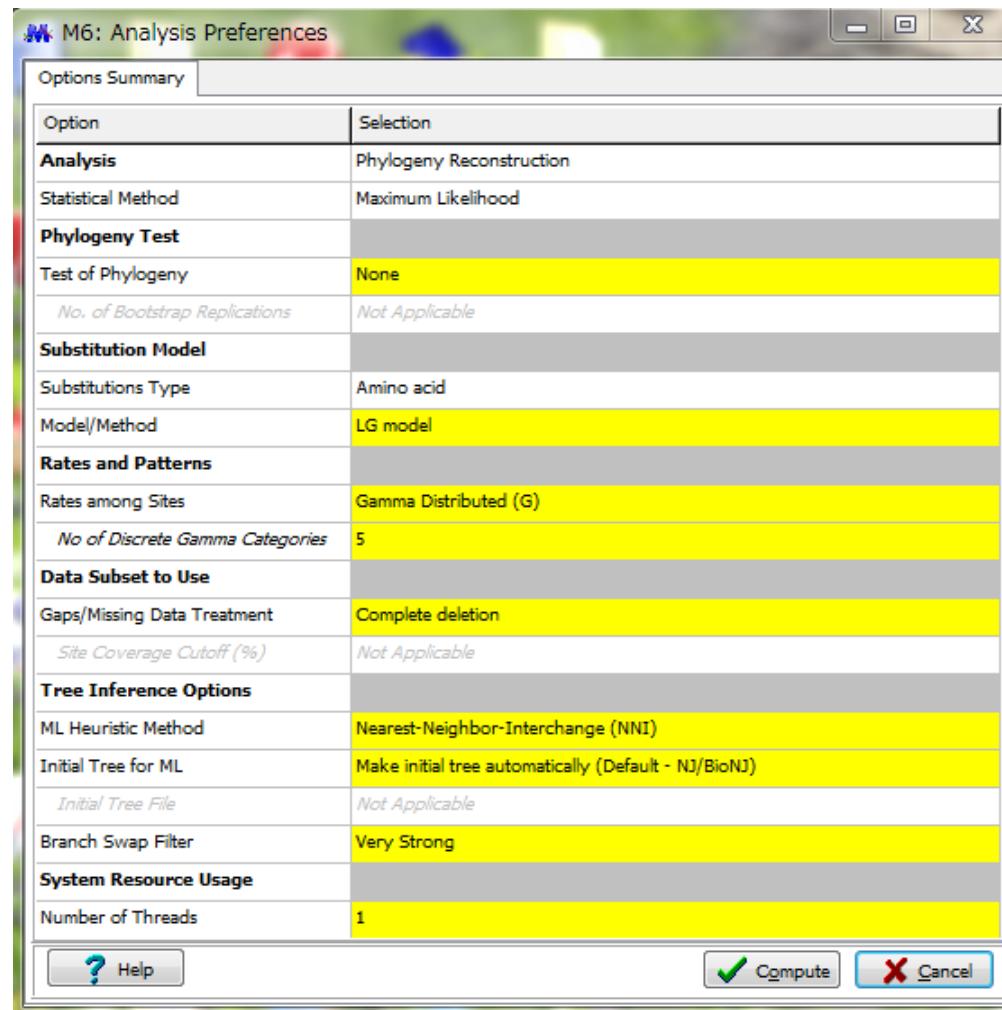
Constructing a phylogenetic tree by the best model

List of options



Constructing a phylogenetic tree by the best model

List of options



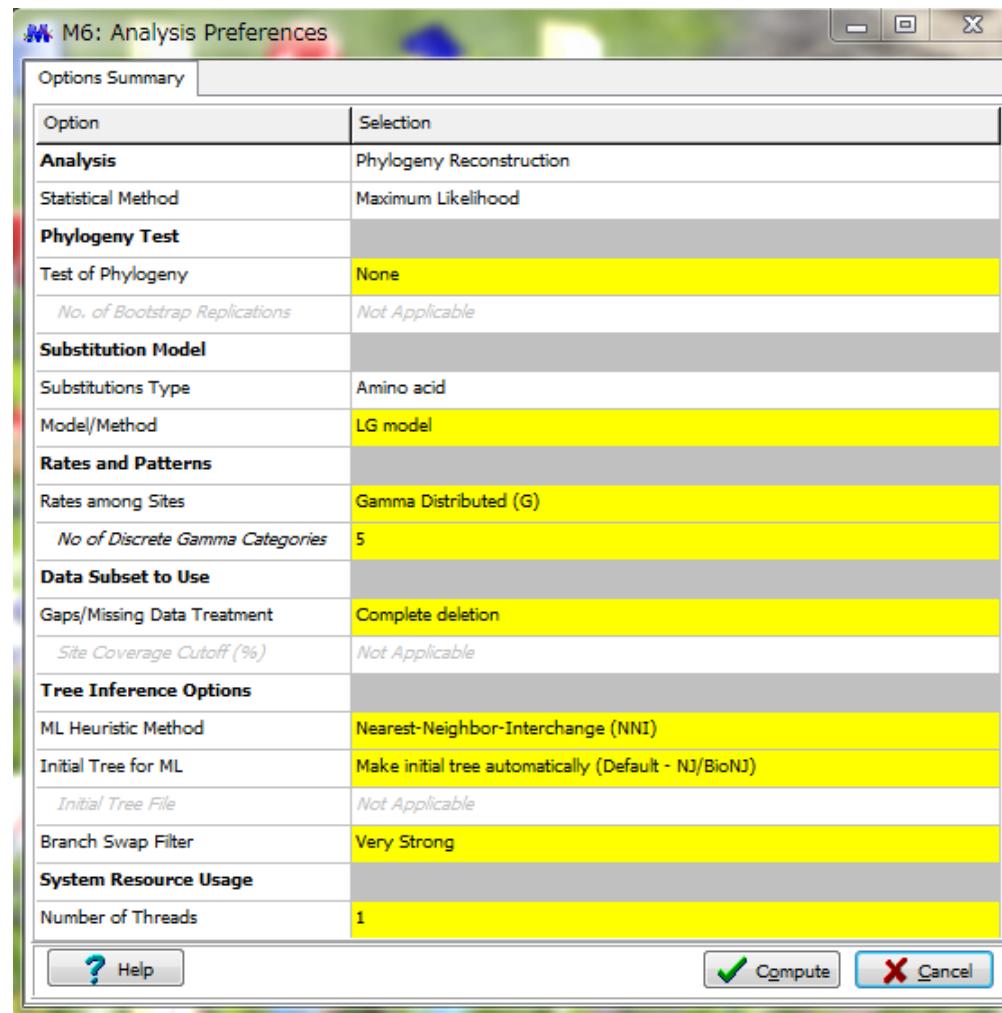
bootstrap (Y/N)

aa replacement matrix

Variable rate among sites

Constructing a phylogenetic tree by the best model

List of options



bootstrap (Y/N)

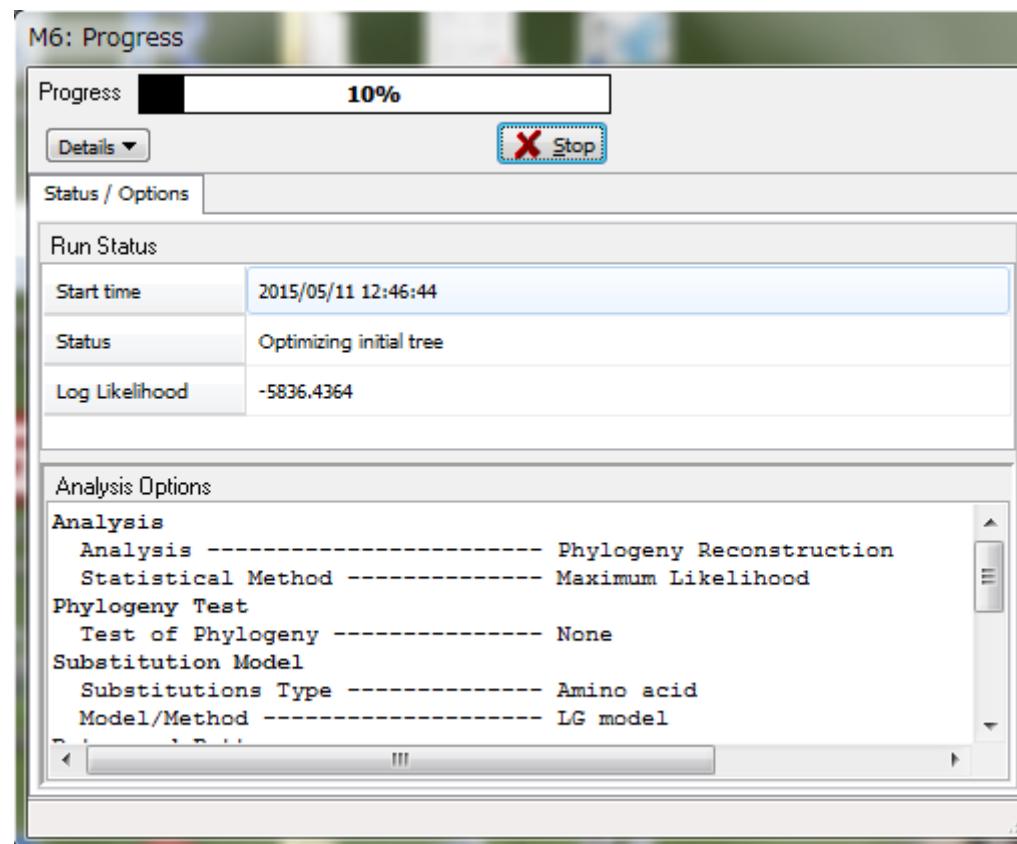
aa replacement matrix

Variable rate among sites

If ready, click Compute].

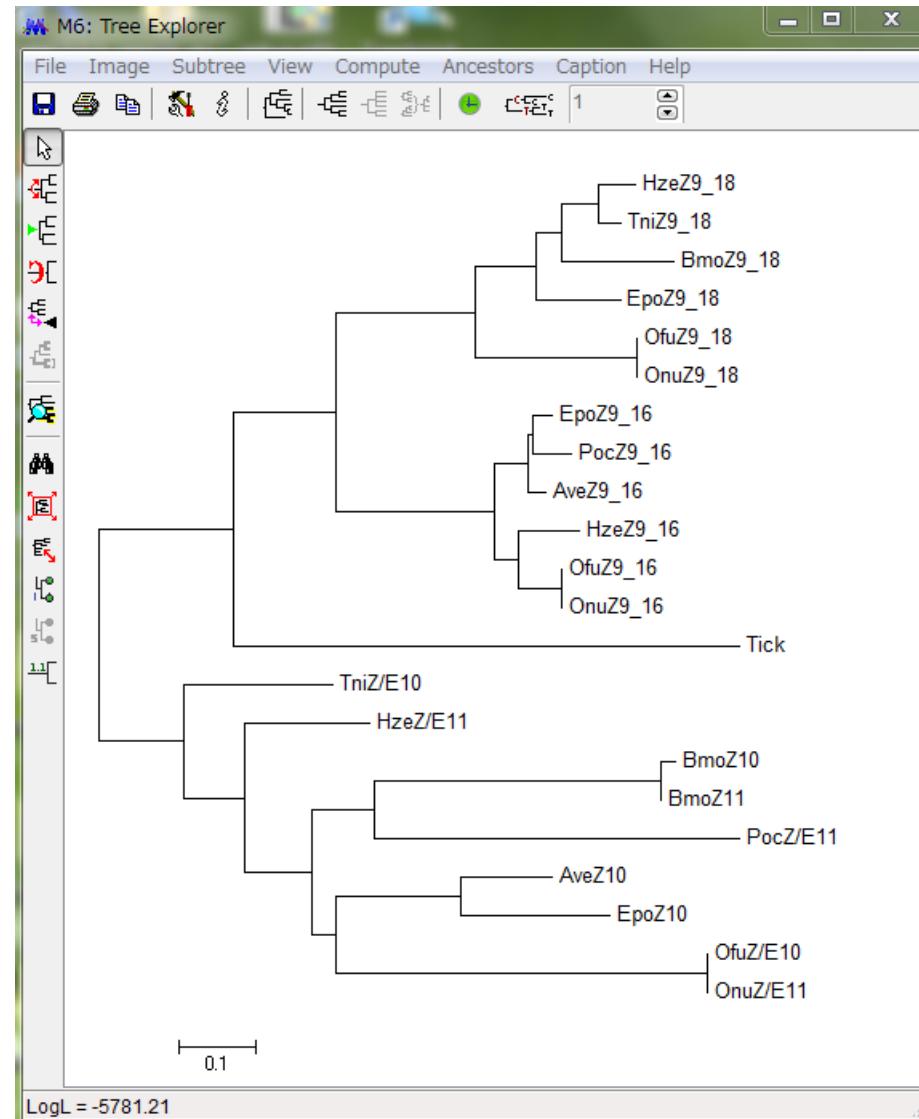
Constructing a phylogenetic tree by the best model

Computation starts ...



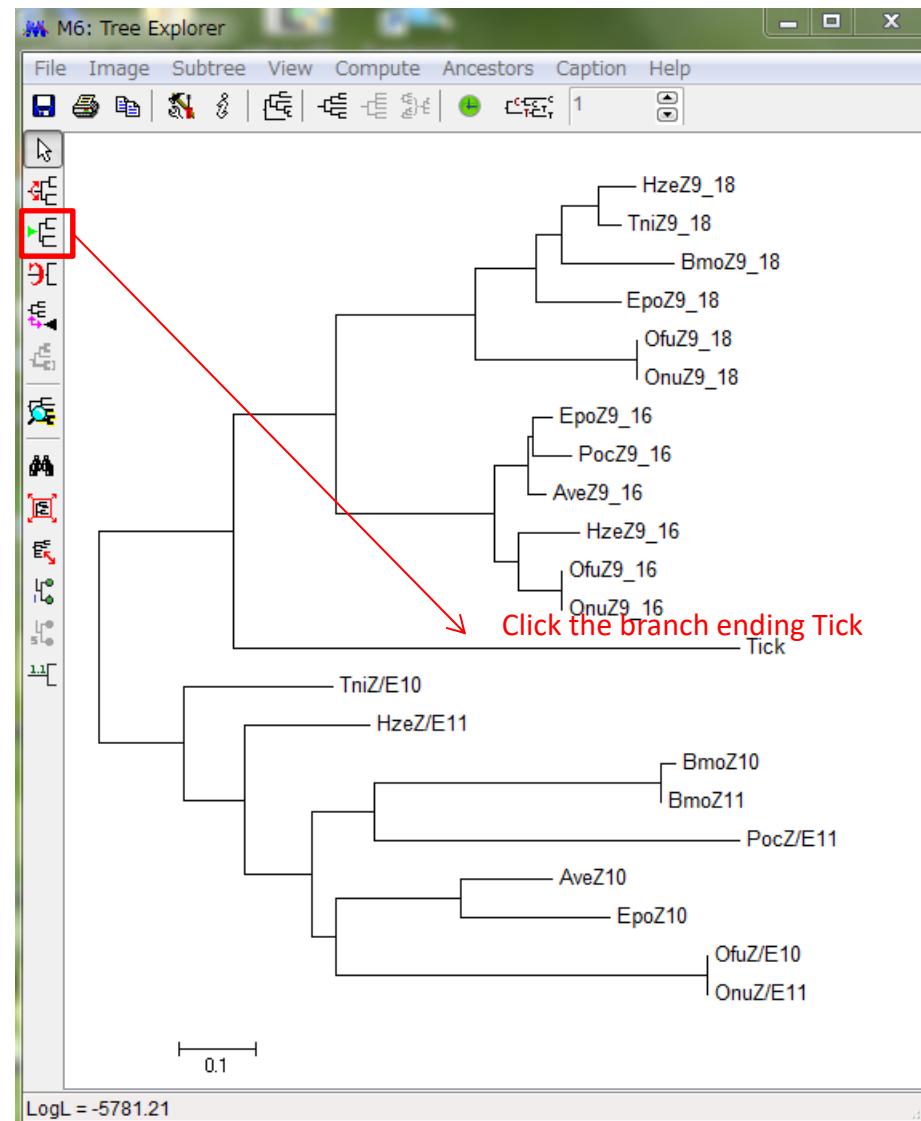
Constructing a phylogenetic tree by the best model

Once the phylogenetic tree is constructed, you will see the figure.



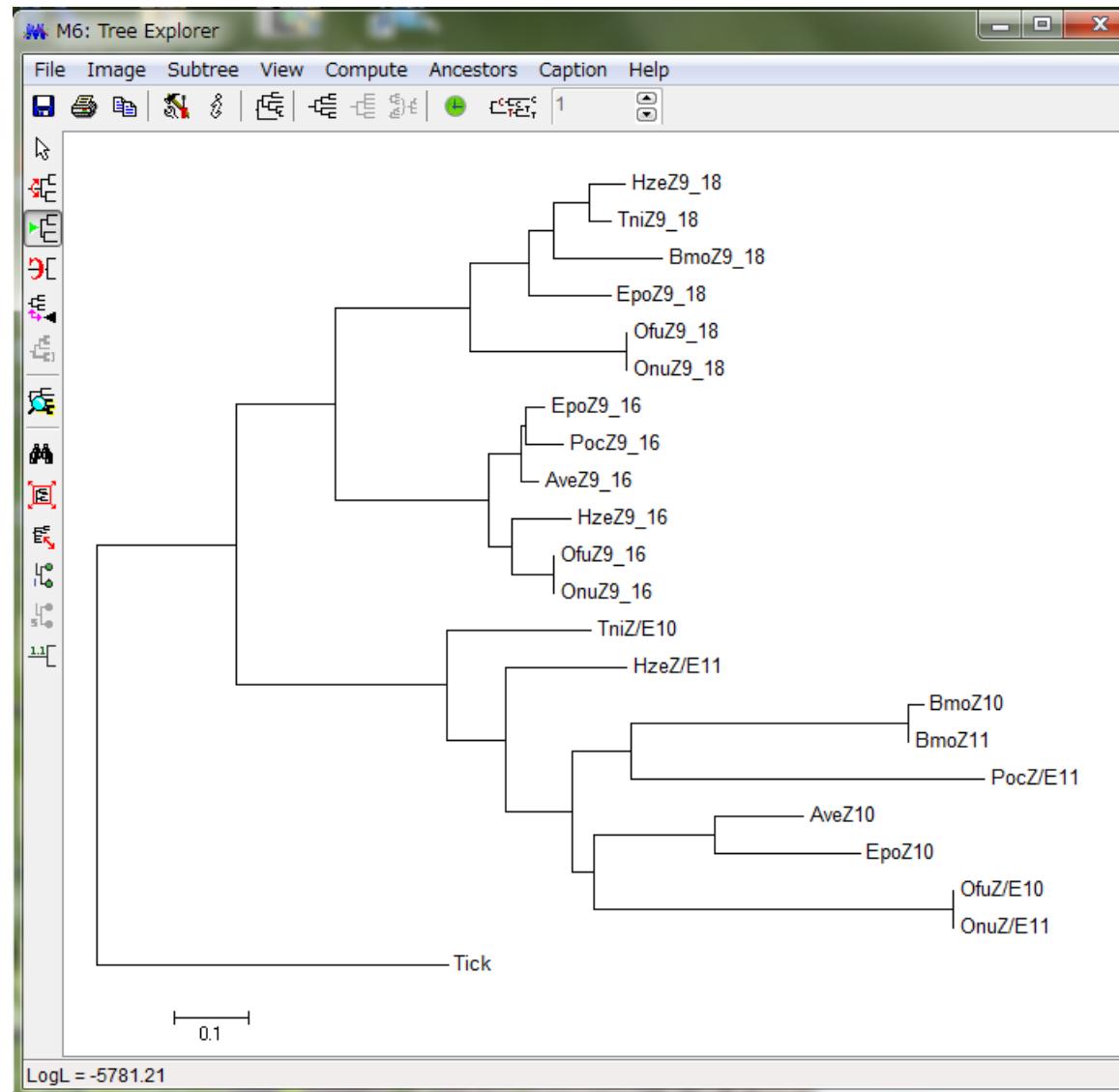
Constructing a phylogenetic tree by the best model

Click  and specify Tick as an outgroup



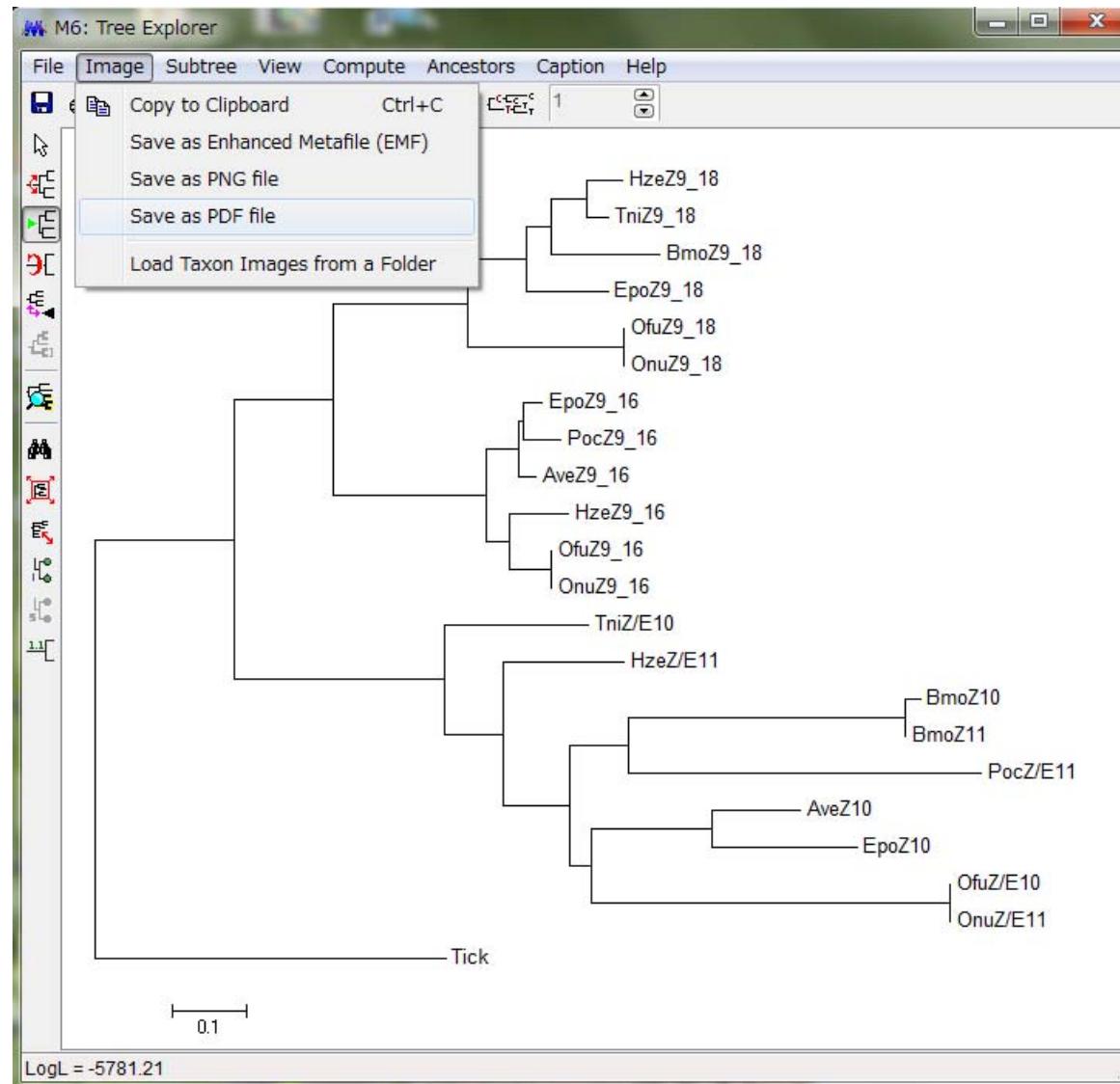
Constructing a phylogenetic tree by the best model

Rooted tree with Tick as an outgroup

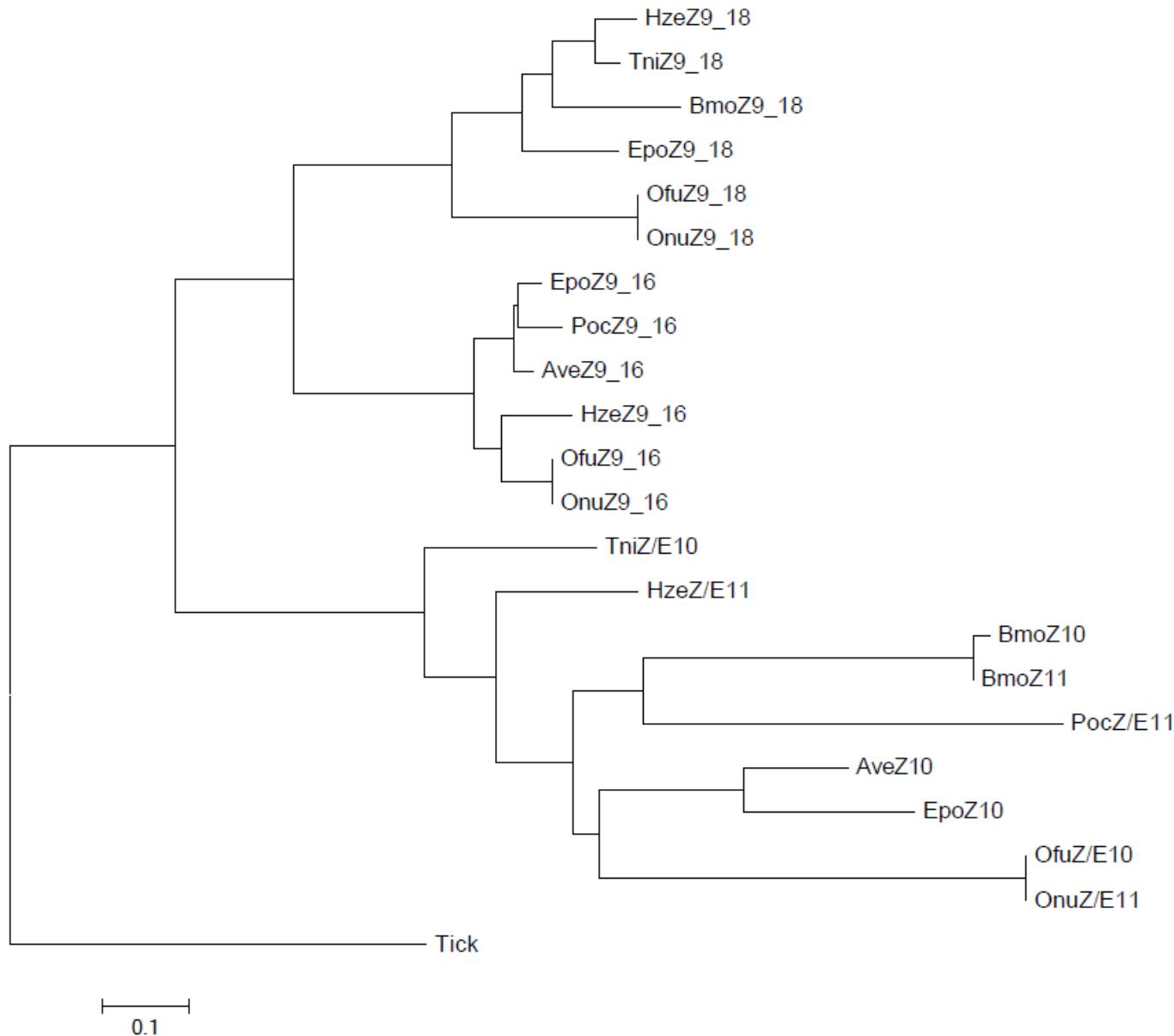


Save the figure as a file

[Image][Save as PDF file] (choice of format: clipboard, emf, png, pdf ...)

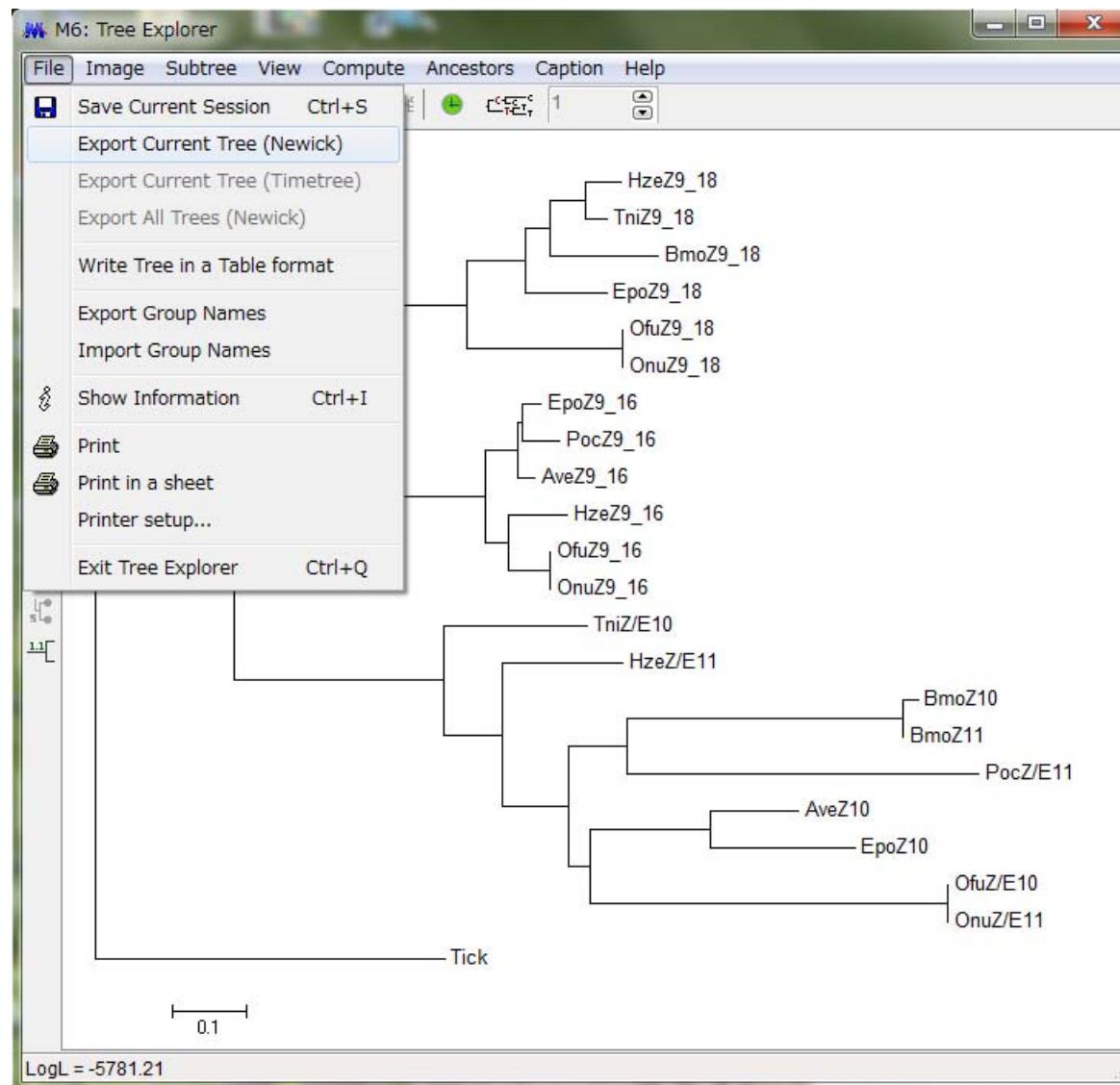


Saved image file



Save the tree information as a newick file

[File][Export Current Tree (Newick)]



Newick file

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**More thought
on the rate
heterogeneity
among sites**

Heterogeneity/homogeneity among sites

MEGA Caption Expert: Find Best-Fit Substitution Model (ML)

File Edit View Help

XL CSV

Table. Maximum Likelihood fits of 48 different amino acid substitution models

Model	Parameters	BIC	AICc	lnL	(+I)	(+G)	f(A)	f(R)	f(N)	f(D)	f(C)	f(Q)	f(E)	f(G)	f(H)
LG+G	42	11957.554	11670.512	-5792.995	n/a	0.70	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
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cpREV+G+I	43	12072.005	11778.141	-5845.797	0.16	1.17	0.076	0.062	0.041	0.037	0.009	0.038	0.050	0.084	0.025
...															
JTT+I+F	61	12399.213	11982.657	-5929.780	0.25	n/a	0.083	0.045	0.045	0.048	0.010	0.019	0.029	0.060	0.050
Dayhoff+I	42	12408.718	12121.676	-6018.576	0.25	n/a	0.087	0.041	0.040	0.047	0.034	0.038	0.050	0.089	0.034
LG	41	12442.188	12161.968	-6039.735	n/a	n/a	0.079	0.056	0.042	0.053	0.013	0.041	0.072	0.057	0.022
Dayhoff+I+F	61	12445.342	12028.786	-5952.844	0.25	n/a	0.083	0.045	0.045	0.048	0.010	0.019	0.029	0.060	0.050
WAG	41	12506.129	12225.909	-6071.705	n/a	n/a	0.087	0.044	0.039	0.057	0.019	0.037	0.058	0.083	0.024
cpREV	41	12526.120	12245.900	-6081.701	n/a	n/a	0.076	0.062	0.041	0.037	0.009	0.038	0.050	0.084	0.025
rtREV	41	12553.704	12273.484	-6095.493	n/a	n/a	0.065	0.045	0.038	0.042	0.011	0.061	0.061	0.064	0.027
...															

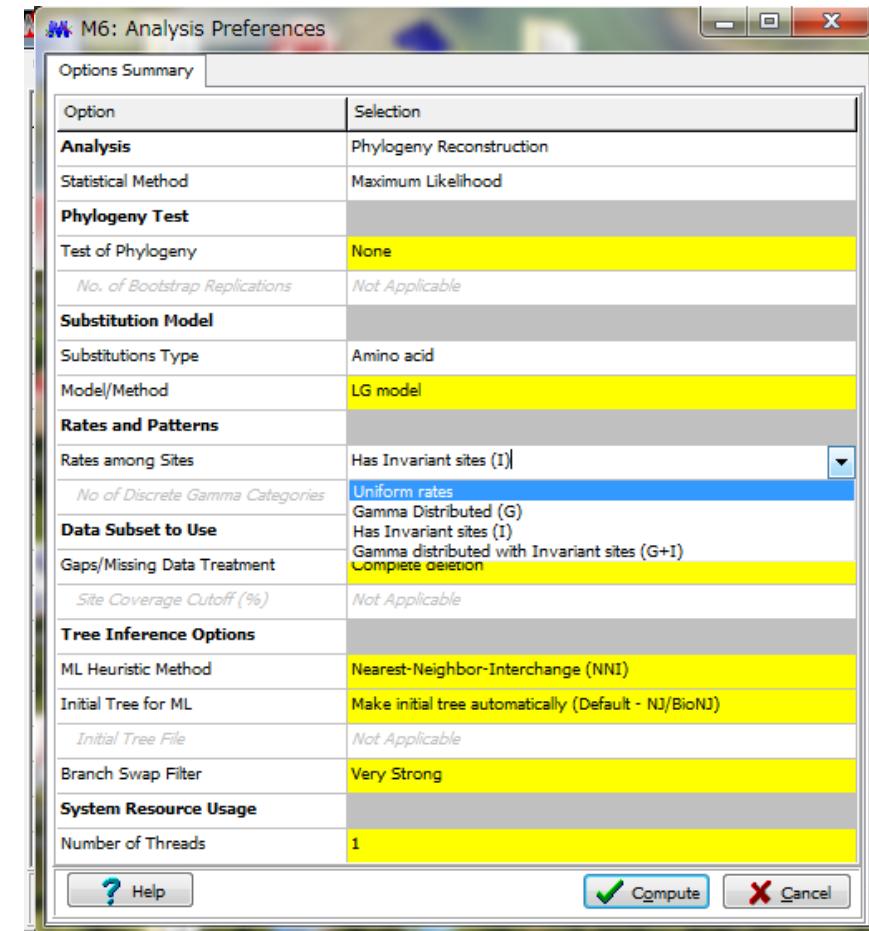
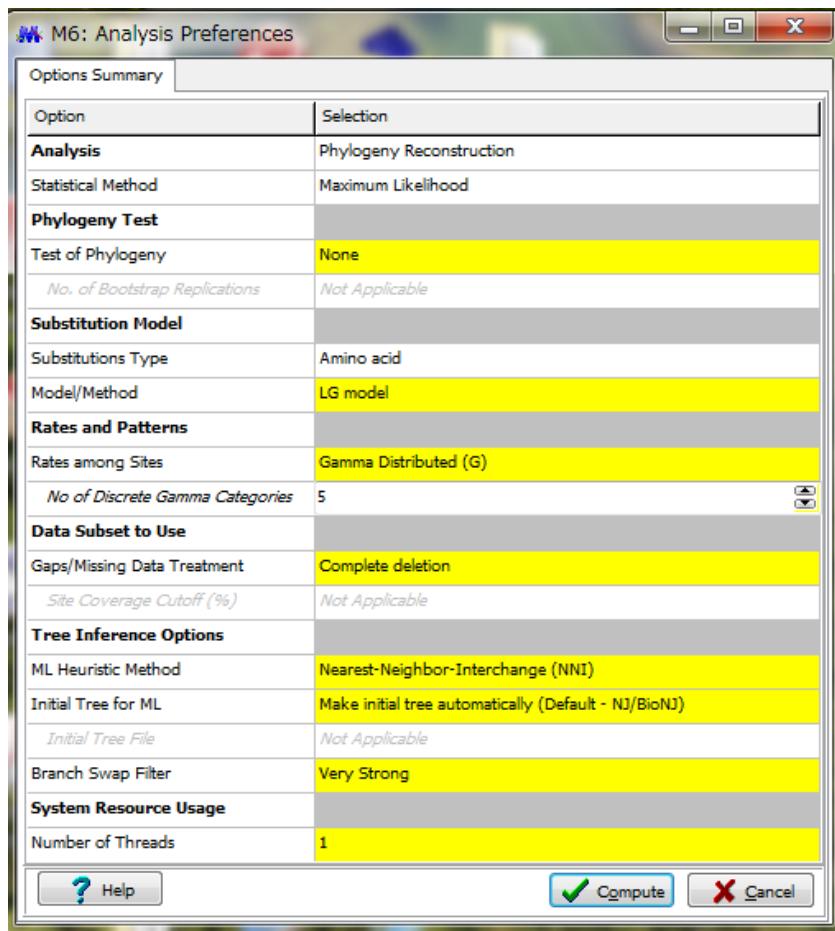
Best model

Assuming uniform rate among sites

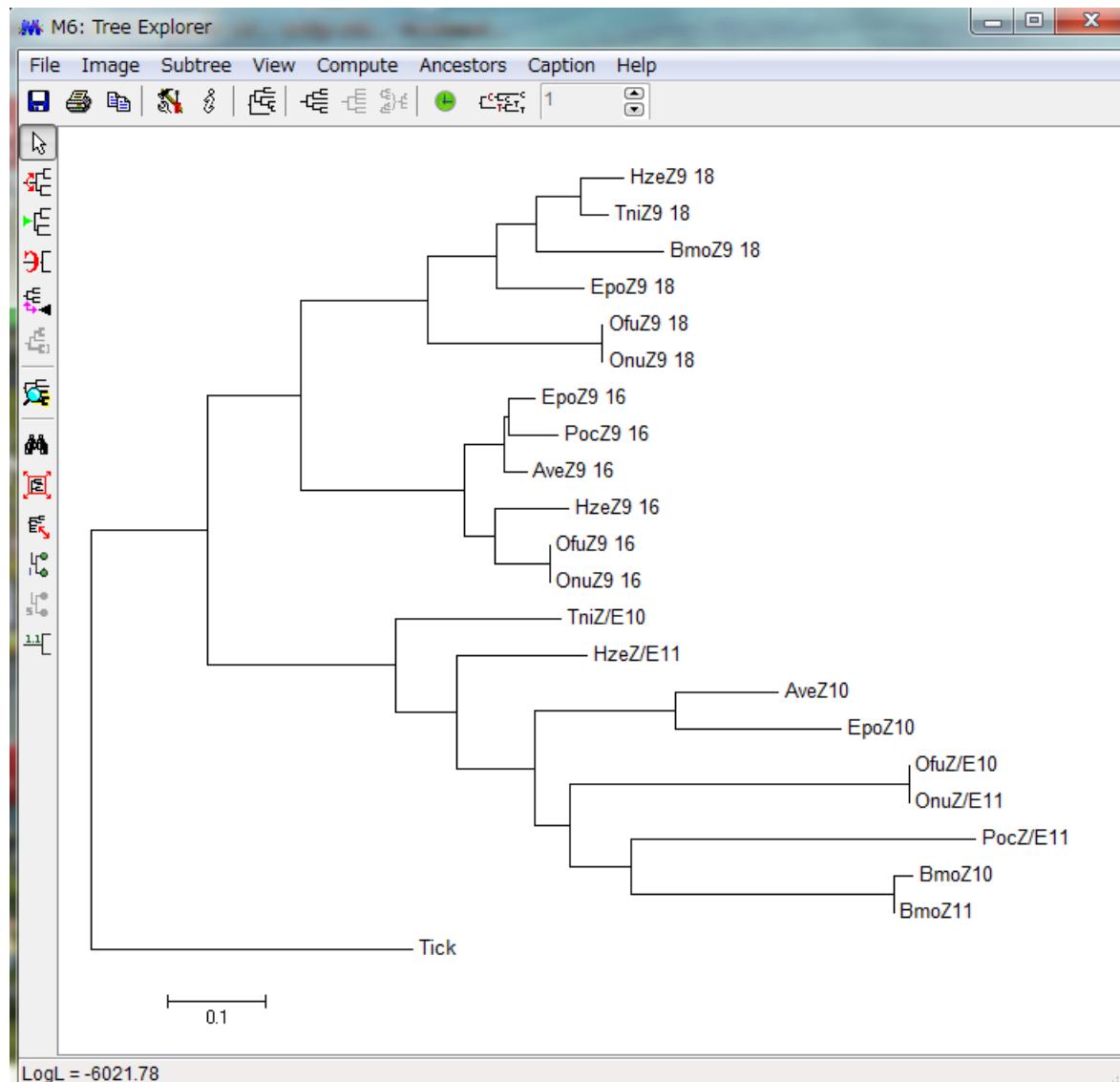
Effect of ignoring rate heterogeneity among sites

[Phylogeny][Construct/Test Maximum Likelihood Tree]

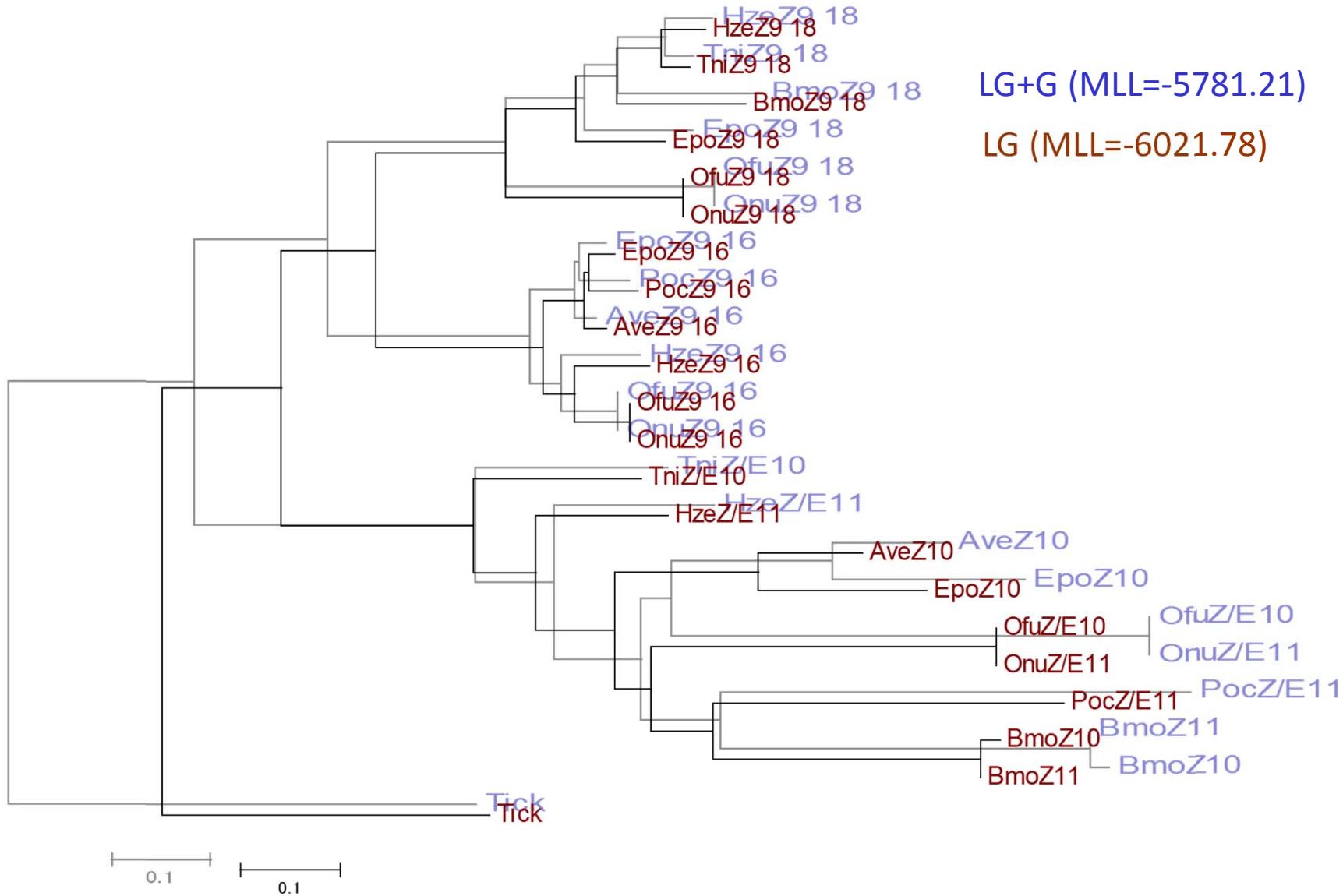
-> [Rates among Sites]: Choose “Uniform rates”



Effect of ignoring rate heterogeneity among sites



Effect of ignoring rate heterogeneity among sites



[coffee break]

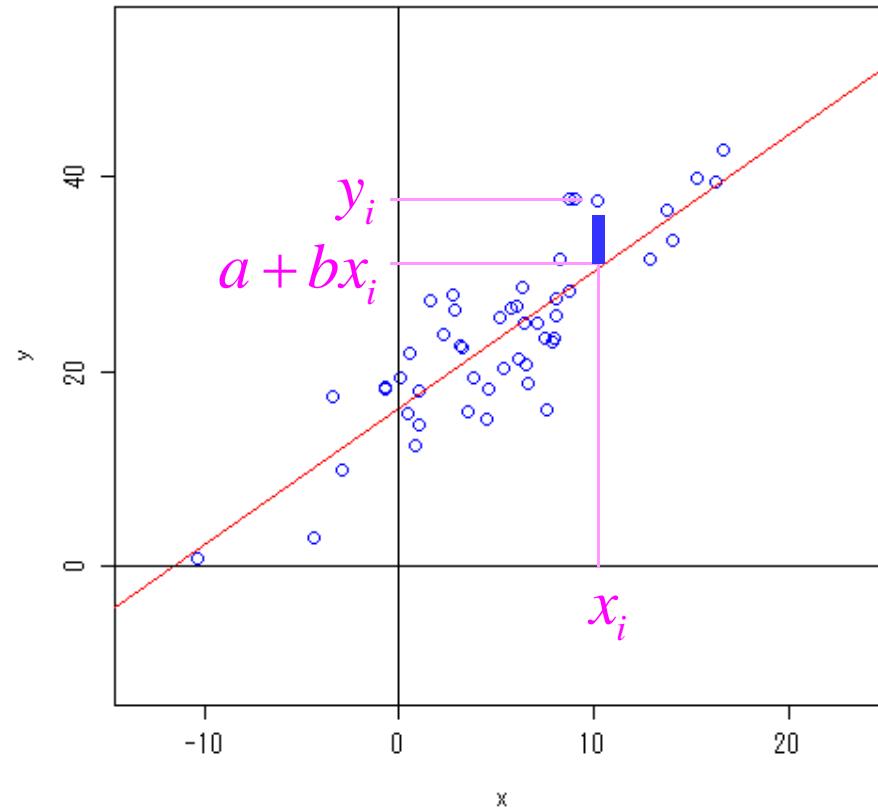
Maximum
likelihood
inference

Likelihood and maximum likelihood (ML) procedure

- Likelihood describes the probability of observing the data by a statistical model.
- Likelihood function: explicit representation of the likelihood in terms of parameters.
- Log likelihood: log of likelihood
- ML procedure: the method of estimating the parameters by maximizing the log likelihood value
 - MLE: the estimate by ML procedure
 - The variance of the MLE is obtained by the inverse of Fisher information quantity (minus the second derivative (Hessian) of the log likelihood function).

ML procedure for Regression analysis

$$y_i = a + bx_i + \varepsilon_i \quad i = 1, \dots, n$$



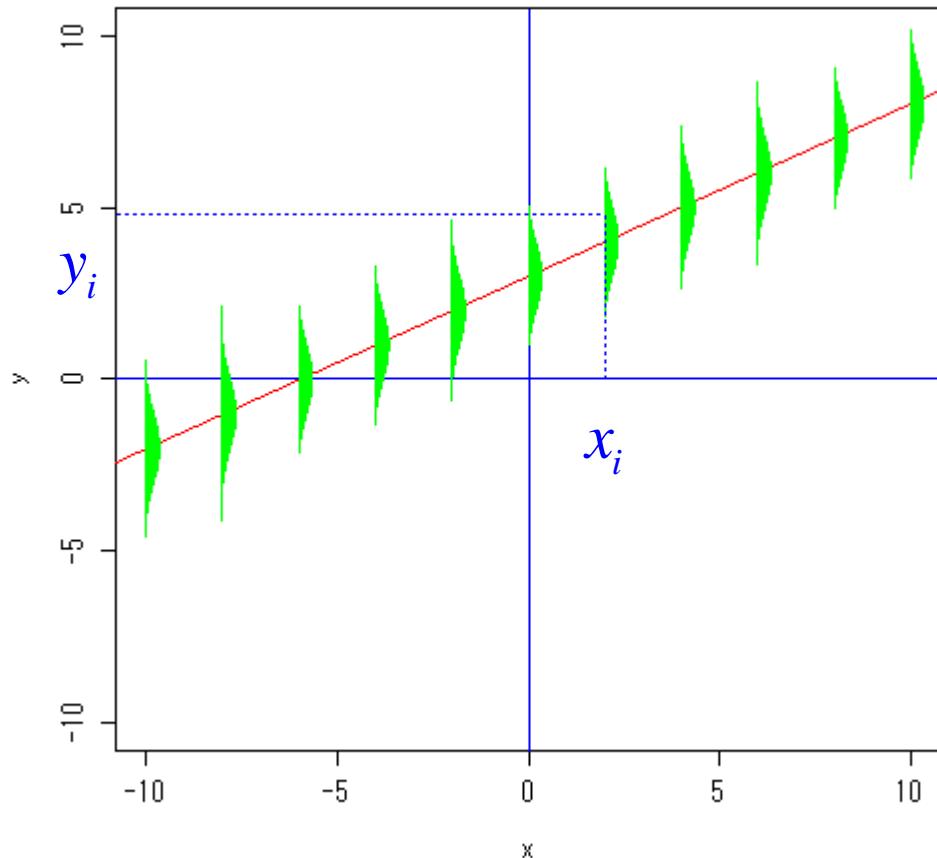
ML procedure for Regression analysis

Statistical model and the likelihood

$$L(a, b, s^2 | y_1, \dots, y_n)$$

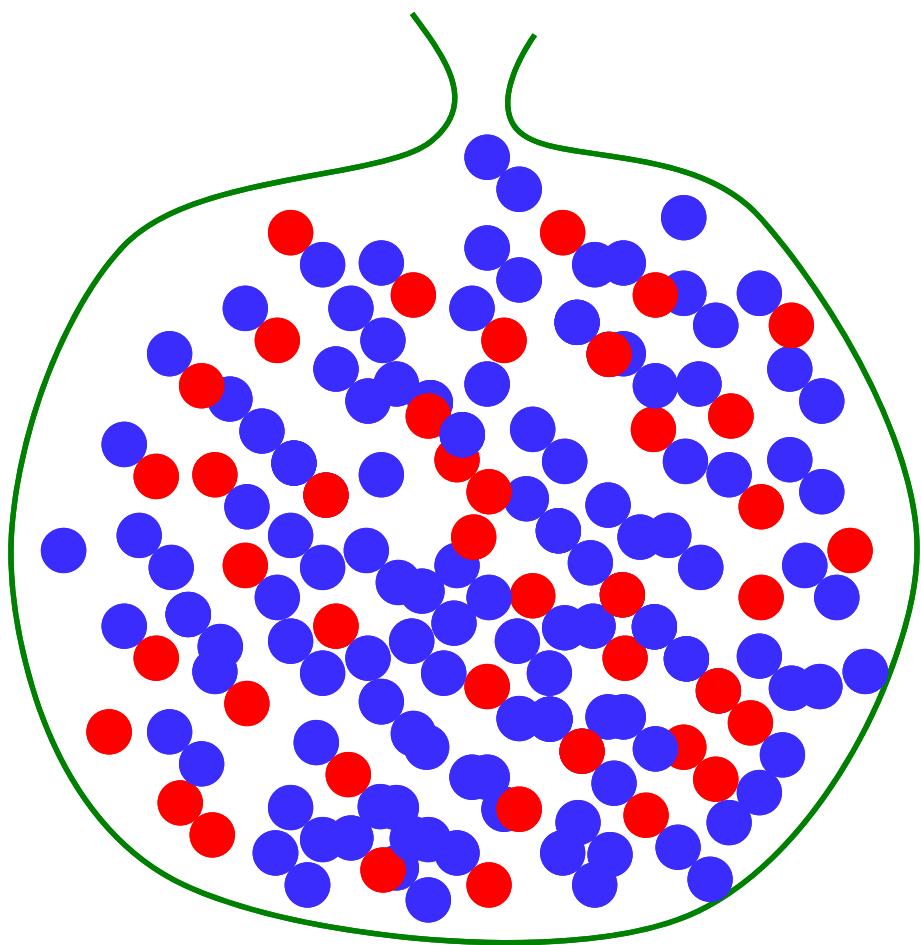
$$= p(y_1) \cdots p(y_n)$$

$$= \left(\frac{1}{\sqrt{2\pi}s^2} \right)^n \exp \left[-\sum_{i=1}^n \frac{(y_i - (a + bx_i))^2}{2s^2} \right]$$

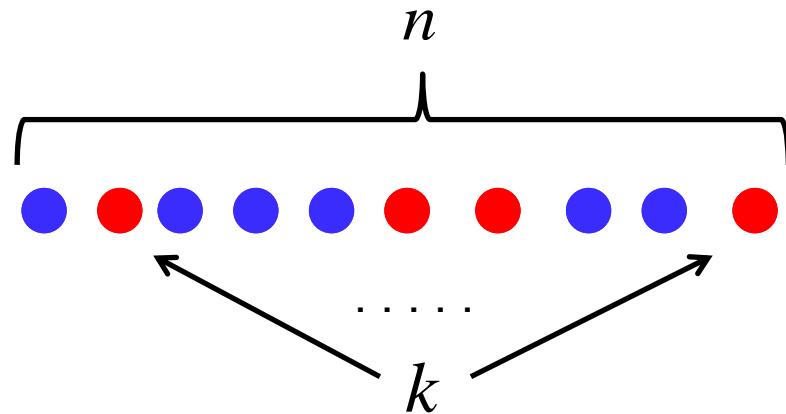
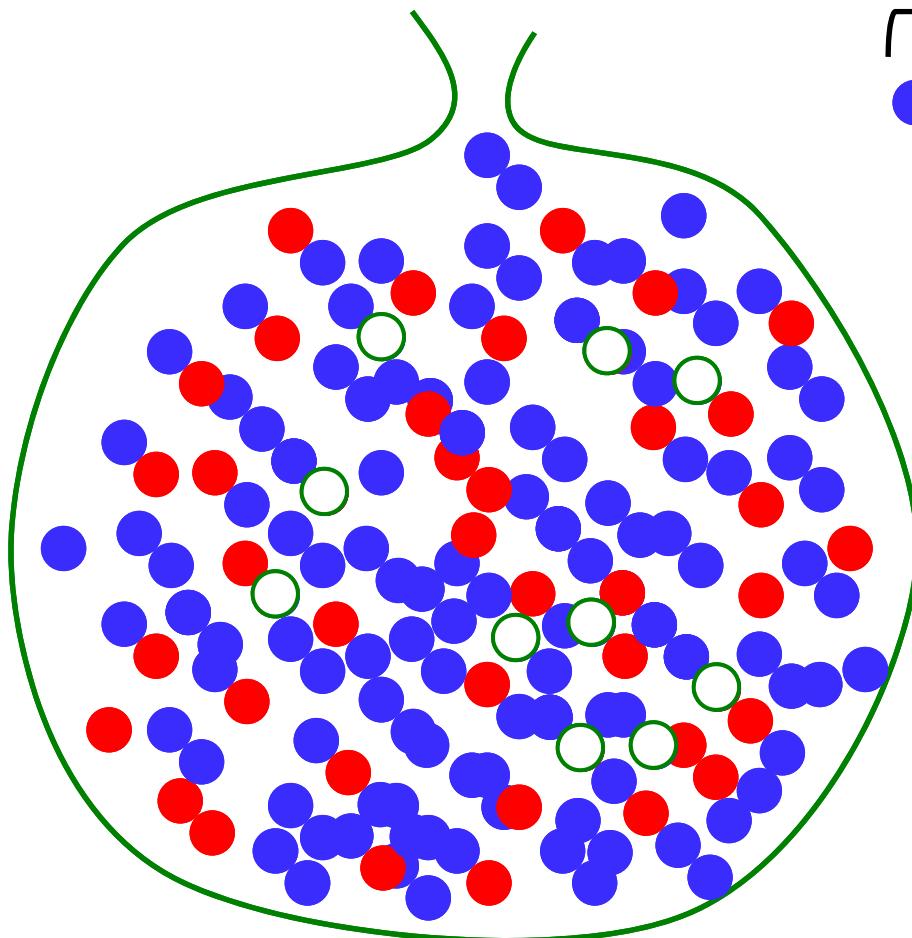


The least squares estimate is the ML estimate assuming normal distribution of error terms.

ML procedure for estimation of the probability



ML procedure for estimation of the probability



$$L = p^k (1-p)^{n-k}$$

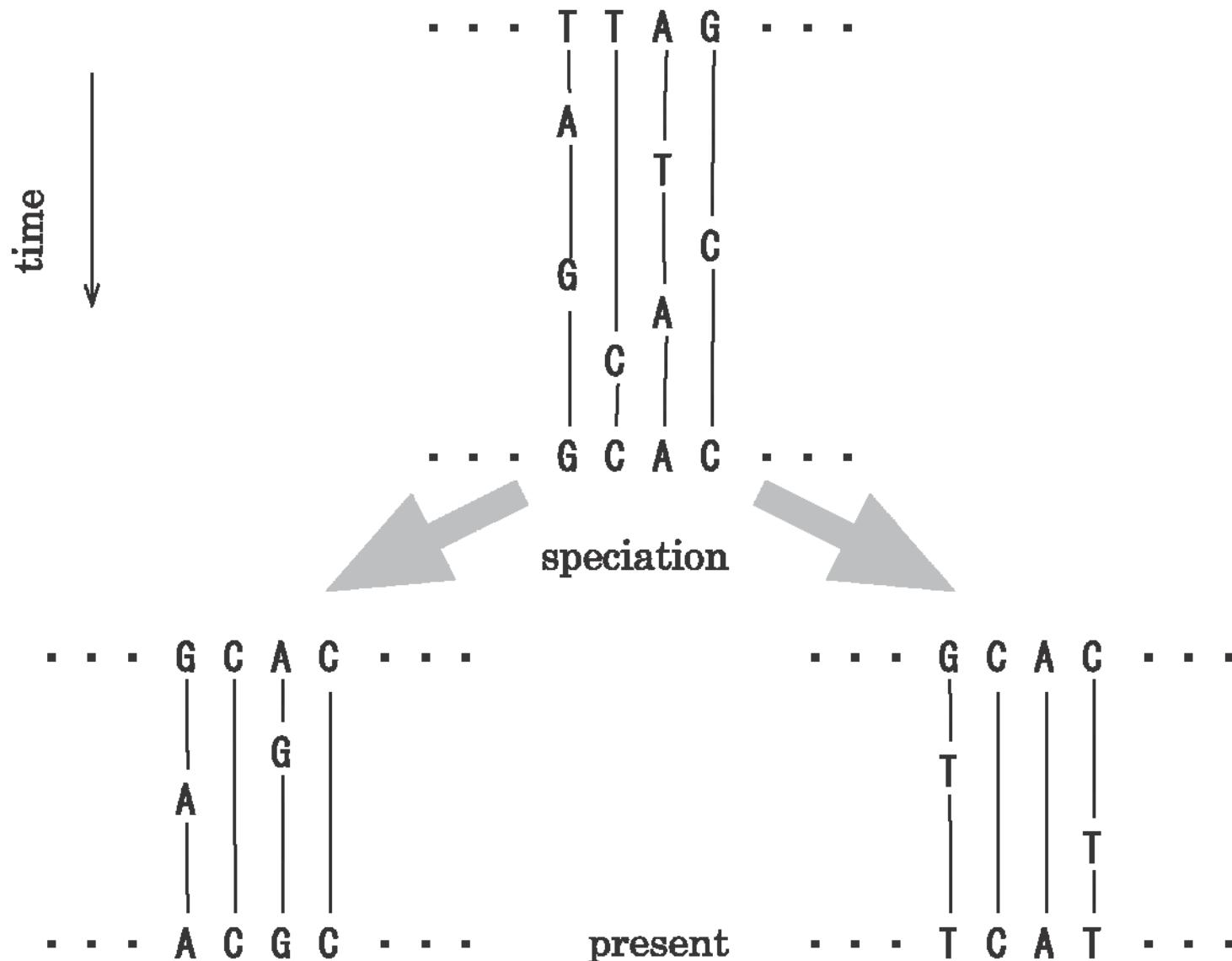
$$\lambda = \log L$$

$$= k \log p + (n - k) \log(1 - p)$$

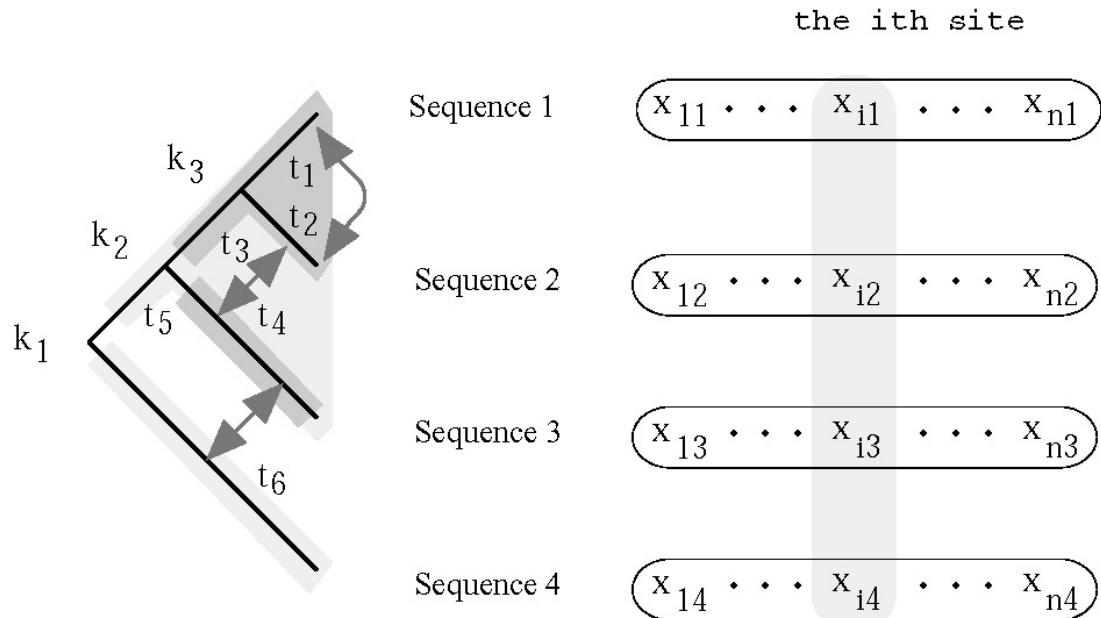
$$\frac{d\lambda}{dp} = \frac{k}{p} + \frac{n - k}{p - 1} = 0$$

$$\hat{p} = \frac{k}{n}$$

Statistical model of molecular evolution



Likelihood of a site (alignment column)



$$f(x_{i1}, \dots, x_{i4} | \lambda, t_1, \dots, t_6) = \sum_{k_1} \pi_{k_1} p_{k_1, x_{i4}}(t_6) \left(\sum_{k_2} p_{k_1, k_2}(t_5) \left(p_{k_2, x_{i3}}(t_4) \left(\sum_{k_3} p_{k_2, k_3}(t_3) (p_{k_3, x_{i2}}(t_2) p_{k_3, x_{i1}}(t_1)) \right) \right) \right)$$

$$l(\boldsymbol{\theta} | \mathbf{X}, T) = \sum_{h=1}^n \log f(\mathbf{X}_h | \boldsymbol{\theta}, T) = \sum_{h=1}^n \log \left[\sum_{Z_{i_0}} \pi_{Z_{i_0}} \prod_{j \in node(T) \setminus i_0} \sum_{Z_j} P_{Z_{anc(j)} Z_j} (t_{anc(j)}, j) \right]$$

$\mathbf{P}(t) = \exp(t\mathbf{R}) = \exp(\textcolor{red}{tr}\mathbf{R}_0)$

branch lengths

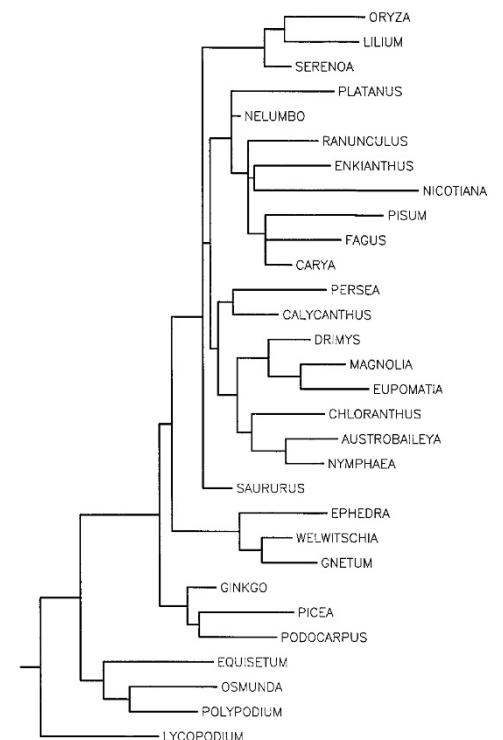
Likelihood of sequences

Species 1	X_{11}	...	X_{1q}	...	X_{1n}
	\vdots	\vdots	\vdots	\vdots	\vdots
Species p	X_{p1}	...	X_{pq}	...	X_{pn}
	\vdots	\vdots	\vdots	\vdots	\vdots
Species s	X_{s1}	...	X_{sq}	...	X_{sn}

$$l(\mathbf{t}, \mathbf{r} | \mathbf{X}) = \log f(\mathbf{X}_1 | \mathbf{t}, \mathbf{r}) + \dots + \log f(\mathbf{X}_q | \mathbf{t}, \mathbf{r}) + \dots + \log f(\mathbf{X}_n | \mathbf{t}, \mathbf{r})$$

$$\mathbf{P}(t) = \exp(t\mathbf{R}) = \exp(t\mathbf{R}_0)$$

branch lengths



Numerical optimization package makes ML easy

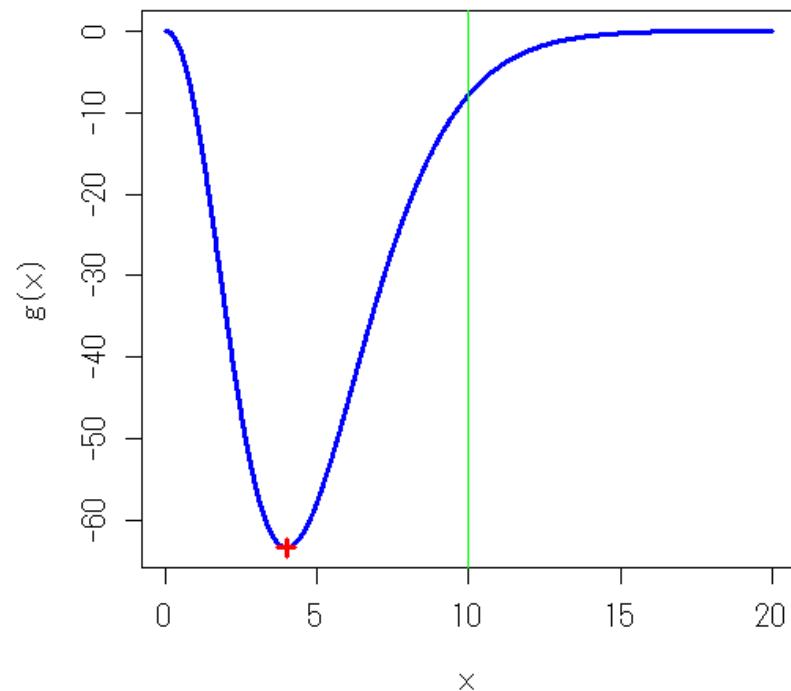
An example function

```
g <- function(x)
  { - exp(-x) * x^2*(x+2)^3}

x0 <- 10
xmin <- optim(x0, g, method="BFGS",
               hessian=T)
xmin
```

```
$par
[1] 4.000001
$value
[1] -63.29885
$hessian
[,1]
[1,] 13.18725
```

solution
Minimum value
Hessian (second derivative)



Once you describe a likelihood function, the computer calculates the MLE and its variance.

Logistic regression of age at sexual maturity

simulated data mature_age.txt

```
read.table("mature_age.txt")->mature_age  
round(mature_age, 3)
```

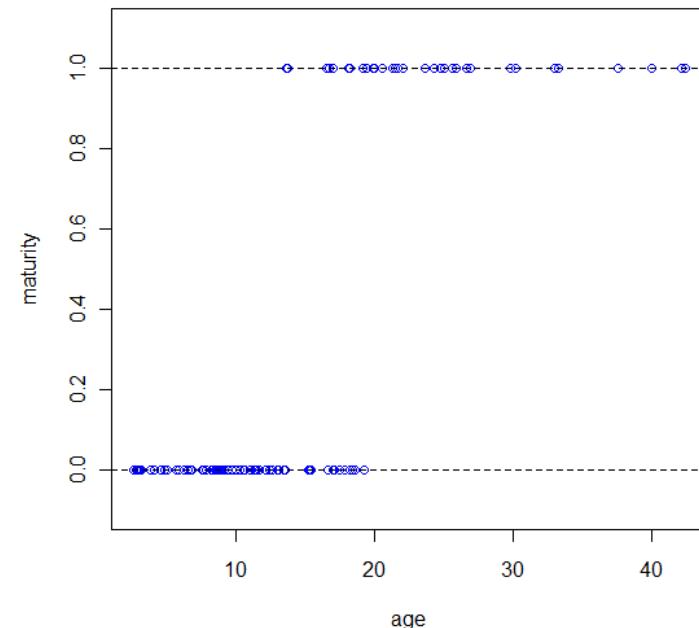
	age	maturity	
1	39.995	1	matured
2	12.427	0	immatured
3	25.069	1	matured
4	20.008	1	matured
5	19.420	1	matured
	

```
plot(maturity~age,mature_age)
```

Generalized linear model with binomial distribution

```
maturity.glm <-  
  glm(maturity~age,binomial,mature_age)  
summary(maturity.glm)
```

	Estimate	Std. Error	z value	Pr(> z)	
(Intercept)	-10.2050	2.4413	-4.18	2.91e-05	***
age	0.5730	0.1377	4.16	3.18e-05	***



$$p = \frac{1}{1 + \exp(10.205 - 0.573x)}$$

Logit transformation and reparametrization

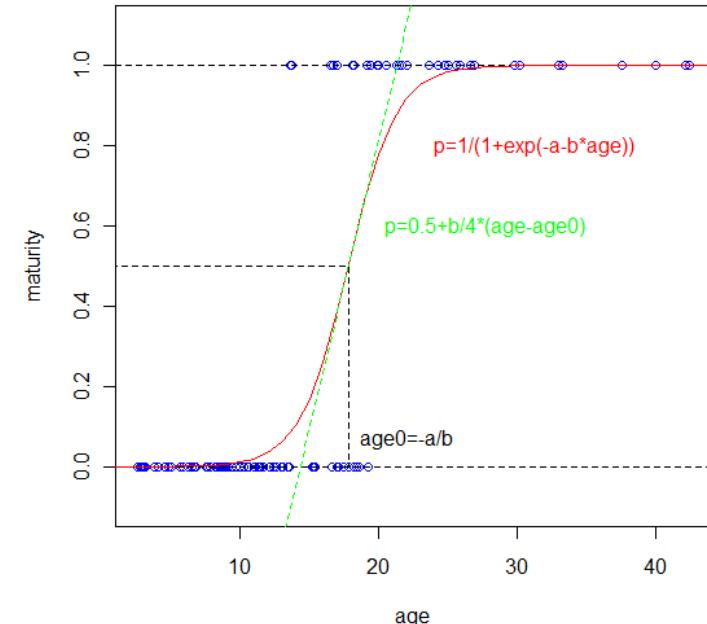
Modeling the logit-transformed variable

$$\log \frac{p}{1-p} = a + bx$$

$$p = \frac{1}{1 + \exp(-a - bx)} \quad \text{Logistic function}$$

Characterization

- $p = 0.5$, when $x = -\frac{a}{b}$
- Well approximated by a line $p = 0.5 + \frac{b}{4} \left(x + \frac{a}{b} \right)$ at $\left(-\frac{a}{b}, 0.5 \right)$



Reparametrization $age0 = -a/b$, $c = b/4$ matches biological characters.

$$p = \frac{1}{1 + \exp(-4c(x - age0))}$$

Maximum likelihood inference by R

```
nlike.glm <- function(theta,data){  
  c <- theta[1]; age0 <- theta[2]  
  n <- dim(data)[1]  
  x <- data[,1]; y <- data[,2]  
  p <- 1/(1+exp(-4*c*(x-age0)))  
  loglike <- sum(dbinom(y,size=1,prob=p,log=T))  
  return(-loglike)  
}  
  
theta0 <- c(0,0)  
theta_est <- optim(theta0,nlike.glm,data=mature_age,hessian=T)  
theta_est  
est <- theta_est$par  
se <- sqrt(diag(solve(theta_est$hessian)))  
mle <-  
data.frame(estimate=est,standard_error=se,row.names=c("c" , "age0" ))  
round(mle,4)
```

	estimate	standard_error
c	0.1433	0.0344
age0	17.8097	0.6923

Fitting K80 model to mtDNA sequences of human and chimpanzee

```
mtCDNA_human.nuc
```

```
CTACCCGCCGCACTGATCATTCTATTCCCCCTTATTGATCCCCACCTCAAATAT  
CTCATACAACCGACTAATTACCACCCAAACATGACTAACAACTAACCTCAAACAA  
ATGATAGCCATAACACAACACTAAAGGACGAACCTGATCTTACTAGTATCCTTAATC
```

```
mtCDNA_chimp.nuc
```

```
TTACCCGCCGCACTGATCAATCATTCTATTCCCCCTTACTGGTCCCCACTTCTAAACAT  
CTCATACAACCGACTAATTACCACCCAAACATGACTAACCTGACCTCAAACAA  
ATAATAACTATACACAGCACTAAAGGACGAACCTGATCTCTACTAGTATCCTTAATC
```

```
scan( "mtCDNA_human.nuc" ,what= " " ) ->human  
human <- strsplit(human,split= " " )  
human <- unlist(human)
```

```
[1] "C" "T" "A" "C" "C" "G" "C" "G" "C" "G" "C" "A" "G" "T" "A" "C"  
[17] "T" "G" "A" "T" "C" "A" "T" "T" "C" "T" "A" "T" "T" "T" "C" "C"  
[33] "C" "C" "C" "T" "C" "T" "A" "T" "T" "G" "A" "T" "C" "C" "C" "C"  
[49] "A" "C" "C" "T" "C" "A" "A" "A" "T" "A" "T" "C" "T" "C" "A"
```

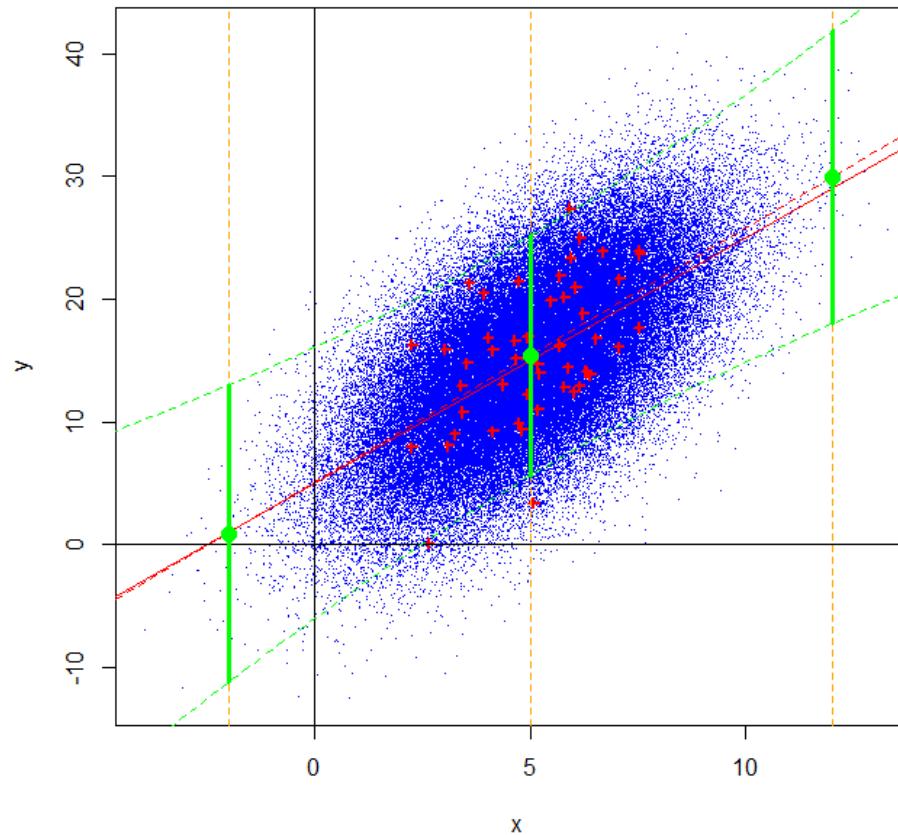
```
table(human,chimp)
```

		chimp			
		A	C	G	T
human	A	2954	17	141	16
	C	18	3163	4	374
	G	165	5	1110	2
	T	15	310	2	2411

	estimate	standard error
t	0.0037	0.0002
alpha	27.7425	1.9271

Kimura80_pair_ML.R

Precision of the prediction and the complexity of the model



Complex models may improve the fitting to the data. However, the variance of the estimates become large, if the model include too many parameters beyond the amount of information in the data.

Model selection based on the precision of the prediction

Variable selection of multiple regression

$$y_i = a + b_1 x_{1i} + \dots + b_k x_{ki} + \varepsilon_i \quad (i = 1, \dots, n)$$

Final prediction error FPE :

$$FPE = \frac{n+k+1}{n-k-1} \hat{\sigma}^2$$

$$\hat{\sigma}^2 = \frac{1}{n} \sum_{i=1}^n (y_i - \hat{a} - \hat{b}_1 x_{1i} - \dots - \hat{b}_k x_{ki})^2$$

Prediction error is obtained by discounting the sum of squared residuals

By considering the predictive power for general models, Akaike (1974) derived a criteria defined by

$$AIC = -2 \times (\text{maximum likelihood value}) + 2 \times (\text{number of parameters})$$

In the special case of regression analysis,

$$AIC = n \log \hat{\sigma}^2 + 2(k + 2) \approx \log FPE - n \log n$$

**Plot trees by
ape**

Read newick file

```
library(ape)
read.tree("desaturase_tree.nwk")->tree
summary(tree)
```

```
Phylogenetic tree: tree

Number of tips: 22
Number of nodes: 20
Branch lengths:
  mean: 0.122944
  variance: 0.024444867
  distribution summary:
    Min.     1st Qu.      Median      3rd Qu.      Max.
0.000000000 0.02457463 0.05800963 0.16716395 0.66505495
  No root edge.
  First ten tip labels: HzeZ9_18
                           Tniz9_18
                           BmoZ9_18
                           EpoZ9_18
                           Ofuz9_18
                           Onuz9_18
                           EpoZ9_16
                           PocZ9_16
                           AveZ9_16
                           HzeZ9_16
  No node labels.
```

Look at the content

names(tree)

```
[1] "edge"           "edge.length"    "Nnode"        "tip.label"
```

tree\$edge

```
      [,1] [,2]  
[1,]   23   24  
[2,]   24   25  
[3,]   25   26  
. . .
```

tree\$edge.length

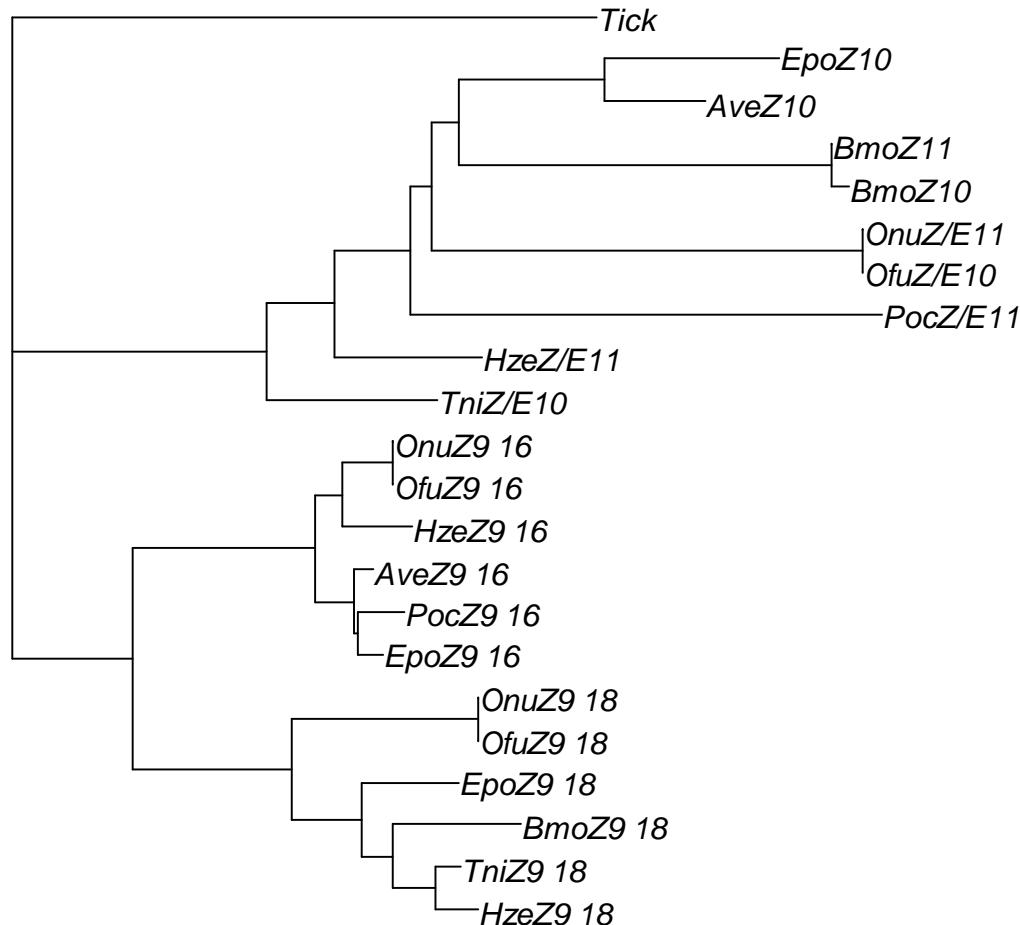
```
[1] 0.13716450 0.18089948 0.08089457 0.03392684 0.04898512 0.04775221  
[7] 0.02881577 0.14738616 0.11048854 0.21265592 0.00000000 0.00000000  
. . .
```

tree\$tip.label

```
[1] "HzeZ9_18"  "TniZ9_18"  "BmoZ9_18"  "EpoZ9_18"  "OfuZ9_18"  "OnuZ9_18"  
[7] "EpoZ9_16"  "PocZ9_16"  "AveZ9_16"  "HzeZ9_16"  "OfuZ9_16"  "OnuZ9_16"  
[13] "TniZ/E10"  "HzeZ/E11"  "PocZ/E11"  "OfuZ/E10"  "OnuZ/E11"  "BmoZ10"  
[19] "BmoZ11"    "AveZ10"   "EpoZ10"    "Tick"
```

Plot the tree

```
plot(tree)
```



Check the structure of tip-labels and node labels

```
tree0 <- tree  
summary(tree0)
```

```
Phylogenetic tree: tree  
  
Number of tips: 22  
Number of nodes: 20  
Branch lengths:  
  mean: 0.122944  
  variance: 0.02444867  
  distribution summary:  
    Min.     1st Qu.      Median      3rd Qu.      Max.  
0.00000000 0.02457463 0.05800963 0.16716395 0.66505495  
  . . .
```

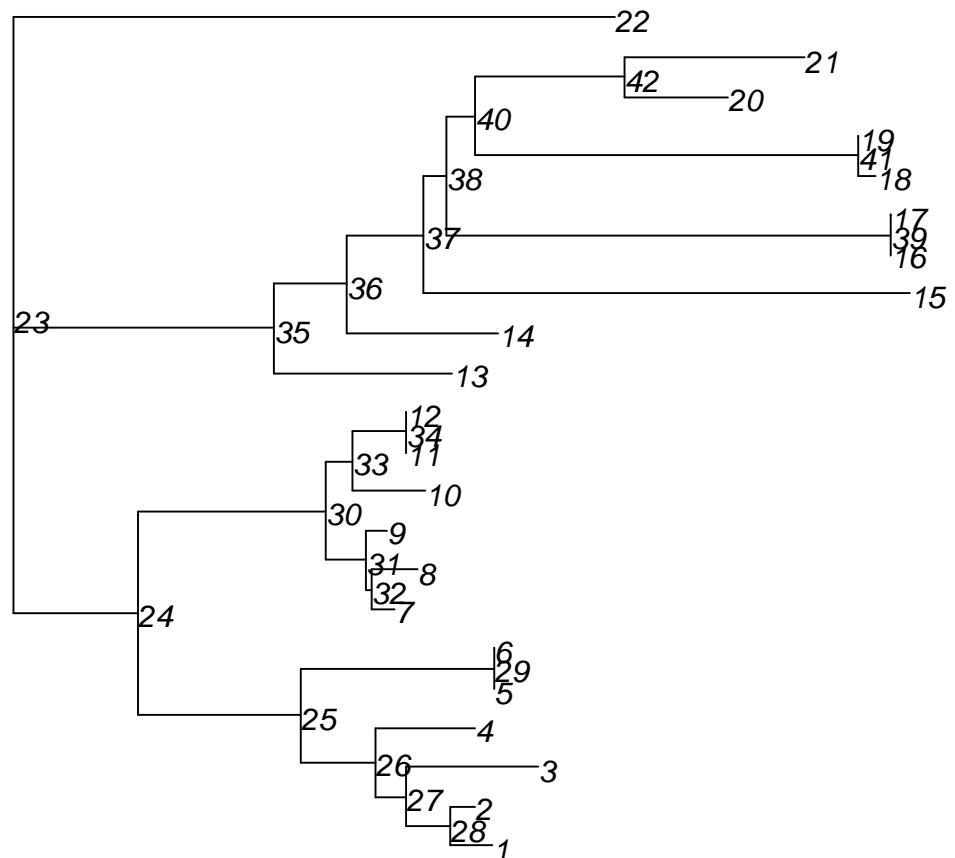
```
tree0$tip.label <- 1:22  
tree0$node.label <- 22+1:20
```

Check the structure of edges

```
plot(tree0, show.node.label=T)
```

```
tree$edge
```

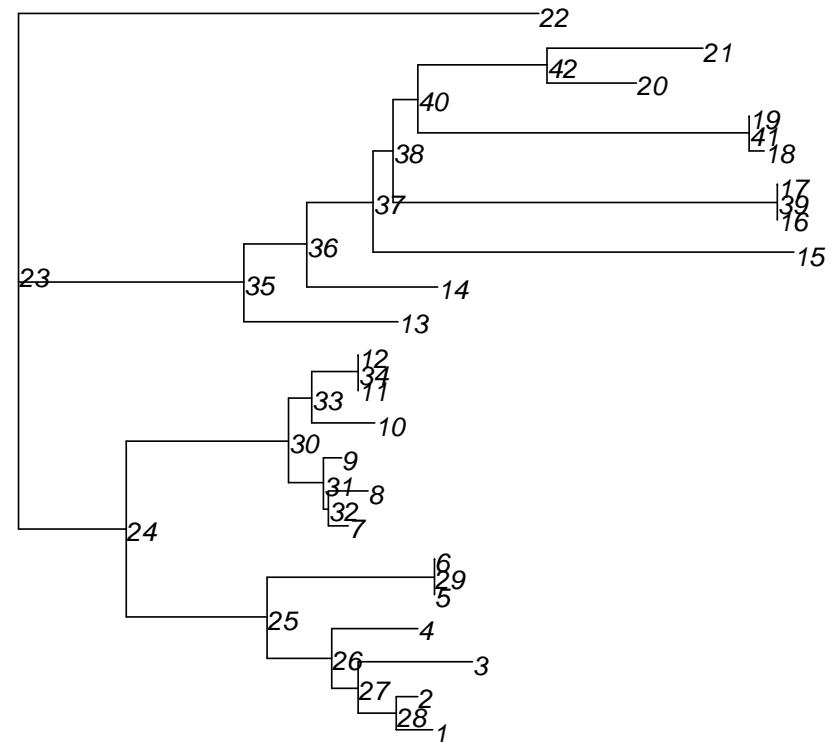
[,1]	[,2]
[1,]	23 24
[2,]	24 25
[3,]	25 26
[4,]	26 27
[5,]	27 28
[6,]	28 1
[7,]	28 2
[8,]	27 3
[9,]	26 4
[10,]	25 29
[11,]	29 5
[12,]	29 6
[13,]	24 30
[14,]	30 31
[15,]	31 32
[16,]	32 7
[17,]	32 8
[18,]	31 9
...	



Define the colors of edges for the three groups

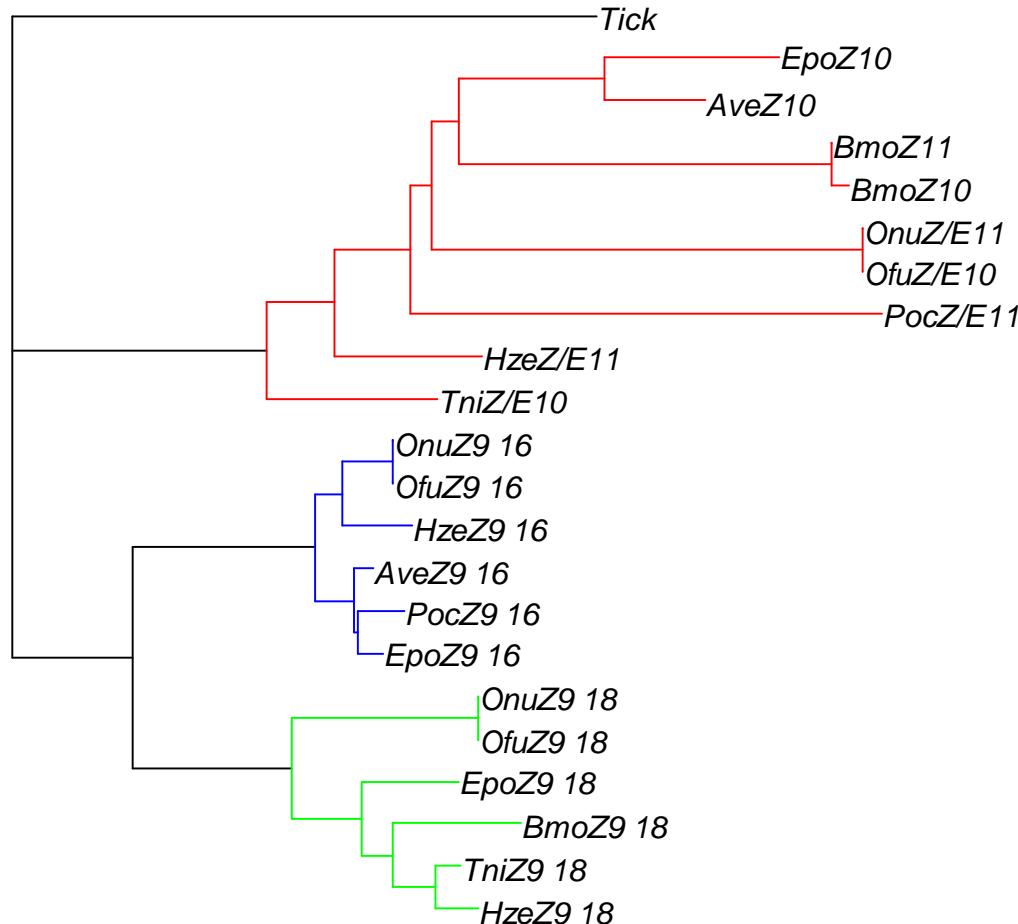
```
edges1 <- which.edge(tree, group=tree$tip.label[1:6])
edges2 <- which.edge(tree, group=tree$tip.label[7:12])
edges3 <- which.edge(tree, group=tree$tip.label[13:21])

edge.colors <- rep("black",dim(tree$edge)[1])
edge.colors[edges1] <- "green"
edge.colors[edges2] <- "blue"
edge.colors[edges3] <- "red"
```



Plot the tree with colors for the three groups

```
plot(tree, edge.color=edge.colors)
```



Assignment 1

Construct the phylogenetic tree of desaturase by the models; LG+G, LG, WAG+G, WAG. Compare the values of AICc and BIC, and interpret the difference of the phylogenetic trees. Please also try ape.

Please send the word file / pdf file named **agri1_name.doc** / **agri1_name.pdf** to:

Hirohisa Kishino (kishino@lrbm.ab.a.u-tokyo.ac.jp).

Here, “name” should be replaced by your name.

Deadline: 26 May 2019 (Sunday)